

**UNIVERSIDADE FEDERAL DE SANTA MARIA  
CENTRO DE CIÊNCIAS DA SAÚDE  
PROGRAMA DE PÓS-GRADUAÇÃO EM REABILITAÇÃO  
FUNCIONAL**

**Bárbara Santos Luccas Duarte**

**Efeitos da Crioterapia em Marcadores Bioquímicos, Histológicos e Funcionais  
após a compressão do nervo isquiático em ratos.**

**Santa Maria, RS  
2018**

**Bárbara Santos Luccas Duarte**

**Efeitos da Crioterapia em Marcadores Bioquímicos, Histológicos e Funcionais  
após a compressão do nervo isquiático em ratos.**

Dissertação apresentada ao Curso de Pós-Graduação em Reabilitação Funcional, da Universidade Federal de Santa Maria (UFSM/RS), como requisito parcial para obtenção do título de **Mestre em Reabilitação Funcional**.

**Orientador: Prof. Dr. Gustavo Orione Puntel**

**Santa Maria, RS  
2018**

Duarte, Bárbara Santos Luccas  
Efeitos da Crioterapia em Marcadores Bioquímicos,  
Histológicos e Funcionais após a compressão do nervo  
isquiático em ratos. / Bárbara Santos Luccas Duarte.-  
2018.  
47 p.; 30 cm

Orientador: Gustavo Orione Puntel  
Coorientador: Felix Alexandre Antunes Soares  
Dissertação (mestrado) - Universidade Federal de Santa  
Maria, Centro de Ciências da Saúde, Programa de Pós  
Graduação em Reabilitação Funcional, RS, 2018

1. Traumatismos dos Nervos Periféricos 2. Crioterapia  
3. Nociceptividade 4. Funcionalidade 5. Estresse  
Oxidativo I. Puntel, Gustavo Orione II. Soares, Felix  
Alexandre Antunes III. Título.

Sistema de geração automática de ficha catalográfica da UFSM. Dados fornecidos pelo  
autor(a). Sob supervisão da Direção da Divisão de Processos Técnicos da Biblioteca  
Central. Bibliotecária responsável Paula Schoenfeldt Patta CRB 10/1728.

**Bárbara Santos Luccas Duarte**

**Efeitos da Crioterapia em Marcadores Bioquímicos, Histológicos e Funcionais  
após a compressão do nervo isquiático em ratos.**

Dissertação apresentada ao Curso de Pós-Graduação em Reabilitação Funcional, da Universidade Federal de Santa Maria (UFSM/RS), como requisito parcial para obtenção do título de **Mestre em Reabilitação Funcional**.

**Aprovado em 27 de julho de 2018:**

---

**Gustavo Orione Puntel, Dr. (UFSM)**  
(Presidente/orientador)

---

**Luis Ulisses Signoi, Dr. (UFSM)**

---

**Cristiane Lenz Dalla Corte, Dra. (UNIPAMPA)**

Santa Maria, RS  
2018

## AGRADECIMENTOS

*Agradeço, primeiramente, a Deus por estar sempre guiando meu caminho, abençoando meus passos, me dando força, coragem e colocando pessoas especiais ao meu lado, sem as quais eu não chegaria até aqui.*

*Ao meu esposo Felipe, meu infinito agradecimento, por sempre estar ao meu lado me dando o suporte necessário para que eu siga sempre em frente mesmo com as adversidades da vida, por compartilhar momentos, por sempre acreditar no meu potencial e por sempre ser essa pessoa incrível da qual eu aprendo todos os dias e que só me faz crescer.*

*À minha família: principalmente a minha Avó, por todos valores a mim transmitidos, por me ensinar a sempre a ter fé, por ser meu exemplo de mulher e por me ensinar o que é o amor. Ao meu avô, minha mãe e minhas tias pelo amor, carinho, dedicação, compreensão e pelos valores a mim transmitidos. Ao meu pai, irmã e Dani, por mesmo longe me incentivarem a dar o melhor de mim e sempre acreditarem na minha capacidade.*

*À Liese e Luise, que estiveram ao meu lado durante esses dois anos e me apoiaram até aqui, com vocês aprendi o valor de uma amizade verdadeira.*

*Ao meu orientador, Professor Gustavo, pela oportunidade, acolhimento, disponibilidade de seu tempo, orientação, atenção e confiança.*

*Ao meu Co-orientador, Professor Felix, por me aceitar em seu laboratório, pelas orientações, por compartilhar seu conhecimento e oferecer todas as condições para que este trabalho se concretizasse.*

*Aos colegas de laboratório, em especial a Andrezza, Rodrigo e Diane pela parceria, amizade, risadas e principalmente pela paciência e ajuda em todos os momentos que precisei.*

*Aos colegas do PPGRF especialmente ao Gustavo, Edneia, Murilo e Tainara pela amizade que formamos, com certeza vocês tornaram este processo mais leve.*

*Aos demais professores, colegas e funcionários deste Programa de Pós-Graduação, agradeço a disposição para me ajudar e a contribuição, de alguma forma, para a realização do meu trabalho e para a minha formação.*

*Enfim, agradeço à Universidade Federal de Santa Maria e ao Programa de Pós-Graduação em Reabilitação Funcional, a possibilidade de realização deste curso.*

## RESUMO

### **EFEITOS DA CRIOTERAPIA EM MARCADORES BIOQUÍMICOS, HISTOLÓGICOS E FUNCIONAIS APÓS A COMPRESSÃO DO NERVO ISQUIÁTICO EM RATOS.**

AUTORA: Bárbara Santos Luccas Duarte

ORIENTADOR: Gustavo Orione Puntel

Os nervos periféricos são responsáveis por levar informações sensoriais da pele, músculos e outros órgãos ao sistema nervoso central (SNC) e informações motoras do SNC para músculos somáticos e órgãos efetores controlados pelo sistema autônomo. A ocorrência de lesões neurais periféricas pode acarretar comprometimentos funcionais de caráter motor e sensitivo nos territórios neurais envolvidos. A crioterapia constitui uma técnica terapêutica extensivamente utilizada como forma de analgesia, no entanto seus efeitos sobre modelos animais experimentais de compressão neural periférica são pouco estudados. O presente estudo objetivou avaliar os efeitos da crioterapia sobre as alterações, bioquímicas (indicativas de estresse oxidativo) e comportamentais (indicativas de alterações comportamentais sensoriais algícas ou motoras) após desenvolvimento de uma lesão por compressão do nervo isquiático em ratos. Os animais foram inicialmente submetidos a avaliação basal do teste de von Frey, 72 horas após o início, os grupos Lesão do Nervo Periférico (PNL), PNL + Crioterapia (Cryo) e Sham foram submetidos ao procedimento cirúrgico de compressão do nervo ciático ou cirurgia simulada. Após 24 horas, o tratamento com 20 minutos de crioterapia foi iniciado uma vez ao dia por 15 dias em todos os grupos, com exceção do Grupo Controle e Sham. Após início do tratamento os testes comportamentais foram repetidos em 7 e 15 dias. No 15ºdia de tratamento os animais foram eutanasiados e amostras de tecido sanguíneo e muscular coletado para análise, estresse oxidativo e marcadores de dano. Os resultados deste estudo demonstram que a crioterapia foi capaz melhorar a função ciática de ratos submetidos a lesão do nervo em 7 dias. Além disso através de testes bioquímicos demonstramos que a lesão aumentou o dano oxidativo e a crioterapia foi capaz de atenuar estes níveis.

Palavras Chaves: Traumatismos dos Nervos Periféricos, Crioterapia, Nociceptividade, Funcionalidade, Estresse Oxidativo

## ABSTRACT

### EFFECTS OF CRYOTHERAPY ON BIOCHEMICAL, HISTOLOGICAL AND FUNCTIONAL MARKERS AFTER SCIATIC NERVE COMPRESSION IN RATS.

Author: Bárbara Santos Luccas Duarte  
Adviser: Gustavo Orione Puntel

Peripheral nerves are responsible for conducting sensory information from skin, muscles and other organs to the central nervous system (CNS) and motor information from the CNS to somatic muscles and effector organs controlled by the autonomic system. The occurrence of peripheral neural lesions can lead to functional and motor impairment in the neural territories involved. Cryotherapy is a therapeutic technique extensively used as a form of analgesia. However its effects on experimental animal models of peripheral neural compression are poorly studied. The present study aimed to evaluate the effects of cryotherapy on oxidative stress and behavioral sensorial or painful motor changes after a sciatic nerve compression injury in rats. The animals were firstly submitted to the von Frey test. After 72 hours the groups Peripheral Nerve Lesion (PNL), PNL + Cryo (Cryo) and Sham were submitted to surgical sciatic nerve compression or surgery simulated. After 24 hours, the 20-minute cryotherapy treatment was started once a day for 15 days in all groups, with the exception of the Control and Sham Group. The tests were repeated in 7 and 15 days after treatment. On the 15<sup>th</sup> day the animals were euthanized and samples of blood and muscle tissue collected for analysis, oxidative stress and damage markers. The results demonstrate that cryotherapy was able to improve the sciatic function of rats subjected to nerve damage at the day 7. In addition, through biochemical tests, we demonstrated that the lesion increased oxidative damage and cryotherapy was able to attenuate these levels.

**Keywords:** Peripheral Nerve Injuries, Cryotherapy, Nociception, Functionality, Oxidative Stress.

## **LISTA DE SIGLAS E/OU ABREVIATURAS**

**ERO:** Espécies Reativas de Oxigênio

**CK:** Creatina Quinase

**DCF-RS:** diclorofluoresceína oxidada

**LDH:** Lactato Desidrogenase

**LNP:** Lesão do Nervo Periférico

**MTT:** Metil Tetrazólio

**SNC:** Sistema Nervoso Periférico

**SNP:** Sistema Nervoso Central

## SUMÁRIO

1 INTRODUÇÃO.....	9
1.1 ASPÉCTOS MORFOLÓGICOS E FUNCIONAIS DO NERVO PERIFÉRICO APÓS LESÃO .....	9
1.2 ALTERAÇÕES BIOQUÍMICAS APÓS LESÃO DO NERVO PERIFÉRICO .....	11
1.3 CRIOTERAPIA .....	12
1.4 OBJETIVOS .....	13
1.4.1 Objetivo Geral .....	13
1.4.2 Objetivos Específicos .....	13
2 DESENVOLVIMENTO .....	14
2.1 ARTIGO CIENTÍFICO .....	15
3. CONCLUSÃO.....	43
REFÊRENCIAS BIBLIOGRÁFICAS: .....	44

## 1 INTRODUÇÃO

### 1.1 ASPÉCTOS MORFOLÓGICOS E FUNCIONAIS DO NERVO PERIFÉRICO APÓS LESÃO.

O Sistema Nervoso Periférico (SNP) é o tecido nervoso situado fora do Sistema Nervoso Central (SNC) e inclui receptores sensitivos, nervos e seus gânglios associados e plexos nervosos (De Graaff, 2003; JUNQUEIRA; CARNEIRO, 1999). Tem a função de estabelecer comunicação e transmitir informações entre o SNC e órgãos da sensibilidade e gânglios e músculos, esta comunicação é executada pela condução nervosa estabelecida por meio de fibras eferentes e aferentes. (JUNQUEIRA; CARNEIRO, 1999). A garantia de uma alta velocidade de condução se dá pela presença de uma bainha de mielina em torno das fibras nervosas e o calibre axônico (ROSSO; YOUNG; SHAHIN, 2017).

Deste modo fibras eferentes são aquelas que saem do SNC conduzindo a informação e estimulando ou ativando a musculatura, ditas assim fibras motoras; já aquelas que conduzem o estímulo ao SNC, são fibras aferentes chamadas de fibras sensitivas (DANGELO; FANTTINI, 2000). No entanto, a maioria dos Nervos levam fibras dos dois tipos, sendo chamados de nervos mistos (JUNQUEIRA; CARNEIRO, 1999). A contração muscular acontece através de nervos motores, cada nervo origina numerosos ramos que se ramificam no tecido conjuntivo.

Lesões nos nervos periféricos (LNP) resultam em perda parcial ou total de funções motoras, sensoriais e autonômicas do membro envolvido, e diminuem substancialmente a velocidade de condução nervosa (RODRÍGUEZ; VALERO-CABRÉ; NAVARRO, 2004; ROSSO; YOUNG; SHAHIN, 2017). Isso ocorre, pois uma das reações mais elementares do SNP é a degeneração Walleriana, que se da quando a continuidade da fibra nervosa é interrompida através de eventos traumáticos, tóxicos, isquêmicos ou metabólicos (DUBOVÝ, 2011). Além disso, LNP promove a degeneração dos axônios e bainhas de mielina distalmente à lesão, refletindo na diminuição de estímulos nervosos que irá repercutir diretamente na perda funcional/motora e/ou sensitiva do membro dependendo do nível e local da lesão (GORDON; TYREMAN; RAJI, 2011; RODRÍGUEZ; VALERO-CABRÉ; NAVARRO, 2004; FERRIGNO, 2005)

Após a LNP, a porção distal do membro inervado pelo nervo ferido sofrerá gradual atrofia e fibrose podendo evoluir de forma irreversível se este não reparado o mais precocemente possível, por este motivo atualmente muitas pesquisas buscam estratégias para promover a regeneração de fibras proximais; minimizando a atrofia da função efetora e de manutenção enquanto aguarda o crescimento (ZHANG et al., 2017). Além das alterações funcionais, lesões no SNP podem vir a contribuir no desenvolvimento de dor neuropática, podendo assim ser associada a disfunção sensorial e alterações na comunicação do SNP e SNC.(GIARDINI et al., 2017) Mais de 70% da população brasileira que procuram um consultório médico por diferentes motivos tem a dor como um deles (ROCHA et al., 2007). A Associação Internacional para o Estudo da Dor (IASP) (MERSKEY; BOGDUK (EDS), 1994) “*define a dor como uma experiência sensorial e emocional desagradável associada a dano real ou potencial de tecidos ou descrita em termos de tal dano*”. A dor pode vir a ser definida como dois tipos, “Nociceptiva” ou “Neuropática” (SCHESTATSKY, 2009).

A Dor Neuropática é definida por um estado de má adaptação provocada por alterações funcionais e estruturais das vias sensitivas centrais e periféricas que produzem modificações no processamento das informações nociceptivas (TEIXEIRA, 2003), pessoas com dor neuropática podem apresentar características clínicas comuns, como dores espontâneas e dores evocadas, alodinia mecânica e ainda, hiperalgesia secundária que se dá pelo comprometimento dos tecidos neurais e não neurais, podendo estar associada à sensibilização central (BENNETT et al., 2006; DETLOFF et al., 2014; KRAYCHETE; GOZZANI; KRAYCHETE, 2008). Além dos fatores fisiológicos, pacientes com neuropatia podem apresentar também irritabilidade, alteração do padrão do sono, alteração do humor e isolamento social, o que reflete diretamente no cotidiano e qualidade de vida (VALL; BRAGA, 2005). Sabe-se também que citocinas pró-inflamatórias estão diretamente associadas a modulação da dor, e sua expressão está ligada a dor neuropática (CHEN, 2012; DETLOFF et al., 2014; KRAYCHETE; GOZZANI; KRAYCHETE, 2008).

Algumas modalidades de tratamento conservador vem sendo estudada para modulação da dor, utilizando técnicas como de mobilização, exercícios físicos e crioterapia (DETLOFF et al., 2014; GIARDINI et al., 2017; KARVAT et al., 2016).

## 1.2 ALTERAÇÕES BIOQUÍMICAS APÓS LESÃO DO NERVO PERIFÉRICO

Quando o SNP é exposto a uma lesão, o potencial de regeneração deste é muito superior ao do SNC, isso é devido as diferentes respostas das células da glia, alterações moleculares e celulares que são ativadas no local da lesão (FARONI et al., 2014; IDE, 1996). Diversos mediadores inflamatórios que desempenham funções essenciais na degeneração waleriana são regulados através da expressão de citocinas/quimiocinas no coto distal após lesões nervosas (DUBOVÝ, 2011). Citocinas são proteínas moduladoras da resposta inflamatória que promovem interações entre células em pequenas distâncias (Kraychete et al., 2008) as citoquinas inflamatórias como (fator de necrose tumoral- $\alpha$  (TNF- $\alpha$ ), IL-1, IL-6 e muitos outros) são produzidas principalmente por macrófagos e mastócitos e têm um importantes papel na resposta inflamatória, incluindo ativação do endotélio e leucócitos na indução da resposta de fase aguda (MEDZHITOV, 2008). A produção de citoquinas pró-inflamatórias, tais como IL-1, IL-6 e TNF- $\alpha$  liberam uma série de espécies reativas no local da inflamação, levando ao estresse oxidativo exagerado (BISWAS, 2016; SONG et al., 2014). No entanto, a inflamação e o estresse oxidativo são eventos fisiopatológicos que estão inter-relacionados (BISWAS, 2016; LEE; SONG; YEUM, 2015).

Komirishetty et al (2017) mostraram que o estresse oxidativo / nitrosativo após compressão do nervo ciático também desempenhou efeitos importantes na patogênese da neuropatia periférica. O estresse oxidativo é um desequilíbrio das reações pró-oxidantes/antioxidantes resultantes das reações metabólicas que utilizam oxigênio, os mecanismos antioxidantes são contornados ocorrendo dano oxidativo. (VALKO et al., 2007; MOTTA, 2011) O excesso de espécie reativa de oxigênio (ERO) pode reagir com os lipídios celulares, proteínas e/ou DNA, danificando ou inibindo sua função normal, a sua formação pode causar danos significativos nas células. Sua formação é mantida em quantidades mínimas por mecanismos antioxidantes visando o equilíbrio redox.(VALKO et al., 2007; MOTTA, 2011) Alguns estudos implicam o estresse oxidativo em várias doenças humanas, bem como no processo de envelhecimento, e indicam também que o estresse oxidativo é uma das principais causas de dano neural, uma vez que esse dano presente irá modular moléculas antioxidantes, que desempenham um papel importante na regeneração da lesão do nervo periférico. (LANZA et al., 2012; QIU et al., 2014; VALKO et al., 2007). Além de marcadores bioquímicos, alguns estudos também associam a alguns achados histológicos e discutem esta relação, Qiu et

al. (2014), registrou a presença de organelas inchadas comprovando a lesão do nervo periférico, e o aumento da expressão das principais enzimas antioxidantes, indicando assim a ocorrência do estresse oxidativo.

Sendo assim muitas modalidades de tratamento tem sido investigada para inibição dos efeitos do estresse oxidativo e investigação de demais possíveis envolvimentos para diminuição de dano neural após lesões do nervo periférico.

### 1.3 CRIOTERAPIA

A técnica de crioterapia é bastante utilizada por profissionais da área da saúde. Um dos motivos são o baixo custo e fácil aplicação, podendo ser utilizada para analgesia, manutenção do edema e na recuperação pós-trauma (GUIRRO; ABIB; MÁXIMO, 1999). Segundo Uchôa; Freitas, (2006) A característica do tratamento através da aplicação do frio se dá pela diminuição de movimento molecular, e é utilizado com o objetivo de diminuir a temperatura local fazendo com que a área de tratamento reaja com uma série de respostas locais e sistêmicas. Após 15 a 20 minutos de aplicação do gelo ocorre uma diminuição do metabolismo levando a limitação da formação de edema, da liberação da histamina, da ativação de neutrófilos, da ação colagenase, de leucócitos sinoviais e maior drenagem linfática (BRANCACCIO et al., 2005).

Muitas publicações discutem os efeitos da crioterapia, porém (GUTIÉRREZ ESPINOZA; LAVADO BUSTAMANTE; MÉNDEZ PÉREZ, 2010) em um estudo de revisão sistemática trazem que existe uma quantidade escassa de publicações de alta qualidade metodológica que a abordam com objetivo de utiliza-la como ferramenta terapêutica no gerenciamento da dor. A variação da temperatura tem grandes efeitos no sistema nervoso periférico, entre os eles, sabe-se que causa um retardo na condução das fibras nervosas contribuindo com o processo de analgesia (FARIAS et al., 2010; RUTKOVE, 2001). Entretanto, dois fatores podem influenciar nos efeitos desta terapia na recuperação do nervo e modulação da dor, que são, a temperatura e o tempo de exposição ao método (HOOSHMAND; HASHMI; EM, 2004). E ainda, relaciona o grau de hipotermia ao dano nervoso, pois acredita-se que a esta diminui a velocidade do impulso quimioelétrico em virtude da solidificação mielínica interrompendo assim a condutibilidade dos nervos mielínicos. Porém, quando exposto a temperaturas muito

baixas, pode evoluir com processo de degeneração axonal distal ao nível da lesão, mas a porção proximal do axônio permanece preservada, essa característica mantém a possibilidade de regeneração do nervo (HSU; STEVENSON, 2015)

Com base na pesquisa da literatura é possível observar que o gelo diminui a velocidade de condução nervosa podendo assim contribuir no processo de analgesia. Entretanto é possível observar também que essa alteração da condutibilidade do nervo pode causar ou induzir processos de hiperalgesia ou danos nervosos quando expostos de maneira inadequada. A crioterapia quando utilizada por um longo período de tempo pode trazer outras características de dor, principalmente de dor neuropática (JU et al., 2012).

Sendo assim mais estudos que investiguem os efeitos variados nos diferentes limites de tempo com esta terapia são necessários. Existe uma variedade de estudos envolvendo o uso da crioterapia como recurso terapêutico, no entanto os resultados são controversos e inconclusivos, isso se dá pelo amplo número de protocolos utilizados com tempo de aplicação variado (GUIRRO; ABIB; MÁXIMO, 1999; GUTIÉRREZ ESPINOZA; LAVADO BUSTAMANTE; MÉNDEZ PÉREZ, 2010). Outro fator determinante para se estabelecer a relevância da técnica se dá por poucos estudos que investiguem os possíveis efeitos negativos desta terapia (RUTKOVE, 2001), tais como as alterações bioquímicas e funcionais, decorrentes do uso incorreto deste recurso, sobre a funcionalidade de nevos periféricos.

## 1.4 OBJETIVOS

### **1.4.1 Objetivo Geral**

Estudar os efeitos da crioterapia sobre as alterações bioquímicas e comportamentais após a compressão cirúrgica do nervo isquiático em ratos.

### **1.4.2 Objetivos Específicos**

- Analisar os efeitos da compressão cirúrgica do nervo isquiático sobre as alterações bioquímicas indicativas de dano oxidativo no tecido muscular do território de inervação do mesmo, a partir da quantificação da viabilidade

celular, dos níveis de diclorofluoriceina oxidada (DCF-RS), da peroxidação lipídica.

- Analisar os níveis plasmáticos de atividade das enzimas creatina quinase (CK) e lactato desidrogenase (LDH) determinados pela compressão cirúrgica do nervo isquiático;
- Analisar as alterações comportamentais motoras de marcha e de percepção sensorial dos animais submetidos à compressão cirúrgica do nervo isquiático;
- Analisar as alterações histopatológicas relacionadas ao processo de cicatrização do nervo isquiático seguintes à compressão cirúrgica do mesmo;
- Investigar os efeitos da crioterapia sobre os objetivos específicos supracitados;

## 2 DESENVOLVIMENTO

O desenvolvimento desta dissertação está apresentado sob a forma de artigo científico. Os itens Materiais e Métodos, Resultados, Discussão dos Resultados e Referências Bibliográficas, encontram-se no próprio artigo. O artigo encontra-se na formatação para publicação da revista científica *Plos One*.

## 2.1 ARTIGO CIENTÍFICO

Full title:

**EFFECTS OF LONG-TERM THERAPEUTIC COLD TREATMENT ON  
BIOCHEMICAL AND FUNCTIONAL MARKERS AFTER SCIATIC NERVE  
COMPRESSION IN RATS.**

**EFFECTS OF LONG-TERM THERAPEUTIC COLD TREATMENT ON  
BIOCHEMICAL AND FUNCTIONAL MARKERS AFTER SCIATIC NERVE  
COMPRESSION IN RATS.**

Bárbara S.L. Duarte<sup>1</sup>, Andrezza B.V. Furtado<sup>1</sup>, Diane D. Hartmann<sup>2</sup>,  
Rodrigo P. Martins<sup>2</sup>, Dèbora F. Gonçalves<sup>2</sup>, Pamela C. da Rosa<sup>2</sup>, Felix A.  
Soares<sup>2</sup>, Gustavo O. Puntel<sup>1,2</sup>.

<sup>1</sup>Centro de Ciências da Saúde, Programa de Pós-graduação em Reabilitação Funcional,  
Universidade Federal de Santa Maria, Santa Maria, RS, Brazil.

<sup>2</sup>Centro de Ciências Naturais e Exatas, Programa de Pós-graduação em Ciências  
Biológicas: Bioquímica Toxicológica, Universidade Federal de Santa Maria,  
Santa Maria, RS, Brazil

## ABSTRACT

Peripheral nerves are responsible for conducting sensory information from skin, muscles and other organs to the central nervous system (CNS) and motor information from the CNS to somatic muscles and effector organs controlled by the autonomic system. The occurrence of peripheral neural lesions can lead to functional and motor impairment in the neural territories involved. Cryotherapy is a therapeutic technique extensively used as a form of analgesia. However its effects on experimental animal models of peripheral neural compression are poorly studied. The present study aimed to evaluate the effects of cryotherapy on oxidative stress and behavioral sensorial or painful motor changes after a sciatic nerve compression injury in rats. The animals were firstly submitted to the von Frey test. After 72 hours the groups Peripheral Nerve Lesion (PNL), PNL + Cryo (Cryo) and Sham were submitted to surgical sciatic nerve compression or surgery simulated. After 24 hours, the 20-minute cryotherapy treatment was started once a day for 15 days in all groups, with the exception of the Control and Sham Group. The tests were repeated in 7 and 15 days after treatment. On the 15<sup>th</sup> day the animals were euthanized and samples of blood and muscle tissue collected for analysis, oxidative stress and damage markers. The results demonstrated that cryotherapy was able to improve the sciatic function of rats subjected to nerve damage at the day 7. In addition, through biochemical tests, we demonstrated that the lesion increased oxidative damage and cryotherapy was able to attenuate these levels.

**Keywords:** Peripheral Nerve Injuries, Cryotherapy, Nociception, Functionality, Oxidative Stress.

## 1           **INTRODUCTION**

2

3           The integrity and balance between the structures related with movements are  
4 responsible for the functional ability of the human's body members. There is a  
5 regulatory role of the Central Nervous System (CNS) that obtains sensory information,  
6 integrates and processes the signals received, and produces the most adequate response  
7 for its preservation and regulating comfort to each position and action. Such information  
8 is received and transmitted through the afferent and efferent fibers of the Peripheral  
9 Nervous System (PNS)(1). The peripheral nerve is the extension of the CNS, being  
10 responsible for the motor and sensory function of the organism. The basic functional  
11 unit of the peripheral nerve is the neuron, consisting of a cellular body and a nerve fiber.  
12 This functional unit of the PNS demands the intimate association of two different cell  
13 types: the Schwann cells (SCs) with the neuronal axons (1,2)

14           PNS lesions are commonly associated with mechanical trauma, and less frequent  
15 after surgical procedures, resulting in acute traumatic compression or crushing (3,4).

16           However, the PNS has an intrinsic capacity for regeneration and repair that is related to  
17 factors such as age, mechanism of injury and location of the lesion(3). The functional  
18 loss resulting from the trauma is influenced by several mechanisms, which may be  
19 reversible. As a result of peripheral lesions, an inflammatory response occurs, leading  
20 to changes in vascular permeability, increased blood flow, swelling and hyperalgesia(5).

21           However, existing treatments for accelerated recovery and nerve regeneration are not  
22 fully available (4). Currently, there are many difficulties both in diagnosis and in  
23 treatment, with little capacity to predict lesions in which recovery is even possible(4).

24           For better recovery and rehabilitation of these lesions, many conservative modalities are  
25 being studied (6).

26           Cryotherapy is one of the conservative modalities of treatment commonly used  
27   in clinical practice for inflammatory response control (pain relief, edema and  
28   hyperthermia reduction, prevention of functionality lost). Commonly used by athletes,  
29   and population in general, the mostly employed methods of cryotherapy consist in  
30   application of wet ice or submersion of body treated areas in cold water. The differences  
31   of treatment protocols consists basically on the duration of each application or the  
32   global period in days after the lesion onset (7). It is known that a 30 minutes treatment  
33   produces twice the physiological effect of a 15 minute treatment, and can produce a  
34   different result under the tissue (7). In this sense, it is perceived that the action of the  
35   treatment by cryotherapy is beneficial. However, negative effects such as the  
36   solidification and hardening of the myelinic lipid, leading to an impairment of the  
37   afferent and efferent nerve stimulus conduction, were already observed depending on  
38   the time of application of the tissue (8),

39           The sciatic nerve is the most important motor nerve, and one important sensorial  
40   nerve, of animal's lower/hind limb. Experimental models of chronic constriction injury  
41   (CCI) already demonstrated impairs in motor skill abilities and sensorial behavior.  
42   Moreover, morphological changes indicative of the sciatic nerve degeneration were  
43   already observed. (5,9) However, biochemical changes suggestive of an oxidative stress  
44   of the skeletal muscle innervated by them is stills not well described.

45           Although the beneficial effects of cryotherapy on reducing the oxidative damage  
46   of skeletal muscle after injuries such as contusion and distension were already described  
47   (10,11) , there is no consensus about cryotherapy effects on experimental models of  
48   nerve injury. Therefore, the present work aimed to investigate for the first time the  
49   effects of cryotherapy on oxidative damage in the skeletal muscle represented by an

50 experimental model of CCI in rats. In addition, also in motor changes and sensory  
51 behavior resulting from CCI.

52

## 53 Materials and methods

### 54 Animals and reagents

55 Wistar male rats, three to four months old weighing between 200 and 250g, were  
56 used for this study, from the central laboratory of the Federal University of Santa Maria  
57 (UFSM). The animals were kept in boxes with free access to food and water, in a room  
58 with controlled temperature ( $22 \pm 2^\circ\text{C}$ ) and with a light / dark cycle of 12 h. All  
59 experimental procedures were conducted according to the ethical principles in animal  
60 experimentation elaborated by the Brazilian College of Animal Experimentation  
61 (COBEA), and according to principles determined in the procedures law for the  
62 scientific use of animals n ° 11,794, of October 08, 2008 . The detailed design of the  
63 experimental procedures of the lesion were submitted 63 to the Committee of Ethics and  
64 Animal Welfare of the Federal University of Santa 64 Maria (UFSM), All the procedure  
65 were in accordance with the guidelines of the 65 Committee on Care and Use of  
66 Experimental Animal Resources of the Federal 66 University of Santa Maria, Brazil  
67 (UFSM; 4185290915)

### 68 Experimental groups

69 The animals (n=29) were randomized and divided into five main homogeneous groups:

- 70 1. Control Group (Ctrl) (n = 5): No surgical intervention or treatment.
- 71 2. Sham group (Sham) (n = 6): submitted to the surgical procedure without  
72 compression of the Sciatic Nerve.

- 73           3. Peripheral nerve injury group (PNI) (n = 6): submitted to surgical  
74           procedure and compression of the Sciatic Nerve.
- 75           4. Cryotherapy group (Cryo) (n = 6): submitted to 20 minutes of daily  
76           cryotherapy session without any surgical procedure involved.
- 77           5. Peripheral Nerve Injury and Cryotherapy Group ( PNI+Cryo) (n = 6):  
78           submitted to the surgical procedure with compression of the Sciatic  
79           Nerve and 20 minutes of cryotherapy daily session.

80           **Study design**

81           The animals were initially submitted to a basal evaluation of the von Frey test,  
82           72 hours after control, the PNI, PNI + Cryo and Sham groups underwent the surgical  
83           procedure of sciatic nerve compression or sham surgery. After 24 hours the treatment  
84           with 20 minutes of cryotherapy was started once a day for 15 days in all the groups  
85           except for the Control and Sham Group.

86           On the seventh day after the start of the experiments, all animals underwent  
87           behavioral and functional analyzes (von-frey and IFC) to evaluate the mechanical  
88           allodynia test and sciatic function index. On the 15th day of cryotherapy, the animals  
89           were again submitted to functional and behavioral tests and were immediately  
90           euthanized after the tests. Then the tissues (blood, gastrocnemius muscle and sciatic  
91           nerve) were collected for biochemical and histological analyzes.

92           The animals were euthanized immediately after the cryotherapy application  
93           protocol. Blood was collected by cardiac puncture in a pre-heparinized syringe, and  
94           both gastrocnemius muscles were quickly removed, weighed, and placed on ice.  
95           Samples of skeletal muscle tissue were homogenized in 10 min in 10 volumes of 10mM  
96           cold Tris (pH 7.4) and centrifuged at 4000 xg for 10 min to produce the low-speed  
97           floating fraction that was used for different biochemical assays at all essay. Whole

98 blood samples were centrifuged at 1500 xg for 10 min for plasma separation which was  
99 used for biochemical analysis.

100

101 **Induction of sciatic nerve injury by chronic constriction**  
102 **injury (CCI)**

103 The rats were anesthetized (n = 18) with ketamine hydrochloride (i.p. 40mg / kg)  
104 associated with xylazine hydrochloride (i.p.10mg / kg). The common sciatic nerve was  
105 exposed to the medial thigh level by incision through the femoral biceps, proximally to  
106 the trifurcation of the sciatic tissue releasing tissue along approximately 7 mm of the  
107 sciatic nerve only, distal to the major trochanter of the femoral bone. The sciatic nerve  
108 was exposed through the femoral biceps muscle, and then its 3 terminal branches (the  
109 sural, common peroneal and tibial) were identified. Four wired ligatures (4.0 black silk)  
110 were tied around the sciatic nerve as described by Bennett GJ, Xie YK (1988) (12)  
111 Stricture of the sciatic nerve was controlled by ligation when a small, brief contraction  
112 occurred in the muscle surrounding the exposed sciatic nerve. The muscle, adjacent  
113 fascia and skin incisions were closed with 4/0 silk thread, and the animal was returned  
114 to its recovery cage. Sham surgeries (n =6) involved the exposure of the sciatic nerve  
115 and its branches with the same procedures, but without creating any constriction /  
116 lesion.

117

118 **Cryotherapy application protocol**

119 After 24 hours of the PNI, the cryotherapy sections were performed individually  
120 for each animal during 20 minutes by using a disposable container with water and ice in  
121 order to kept the temperature at 5 °C (the temperature was monitored by a

122 analogical/digital thermometer submerged in the container along the entire period of  
123 treatment). The right hind limb of the animal was submerged until the medial region of  
124 the knee joint accordingly for Karvat et al., 2016 (6). This protocol was repeated daily  
125 for 15 consecutive days, always between 10 AM and 1PM hours. It is important to  
126 emphasize that this treatment protocol was innovator in the treatment of a peripheral  
127 nerve injury in rats.

128

## 129 **Sensorial and motor behavioral tests**

130 The evaluation of mechanical allodynia and motor was performed using the Von  
131 Frey and Sciatic Function Index (SFI) tests.

132 Von Frey was used to measure the 50% paw withdrawal threshold, the 50% threshold  
133 was determined using the method proposed by Dixon WJ. (1988) (13). To perform the  
134 tests the rats were allocated in individual stainless steel chambers with a wire mesh  
135 floor. Seven von Frey filaments (Remington's Sensory-Touch Sensors, Remington  
136 Medical Equipment Ltd., ON, Canada) were applied to the plantar surfaces of the right  
137 hind paw. The stimulation with the filaments was performed 3 consecutive times.  
138 Removal of the hind paw was considered a positive response. The results were analyzed  
139 with their respective baseline results.

140 The SFI proposed by De Medinaceli (9) was used to evaluate the motor function  
141 in order to evaluate the functional recovery of the sciatic nerve after injury and  
142 cryotherapy application. The animals were placed on a wooden path so they could only  
143 walk forward, with a dark shelter at the end. The animals were placed to walk on a  
144 white paper, the paws will be stamped so that the appropriate measures were taken for  
145 analysis. The following calculation was based on the equation proposed by (14): SFI =  
146  $283.3 \times [(EPL-NPL) / NPL] + 109.5 \times [ETS-NTS] / NTS + 13.3 \times [-EITS] / NITS - 8.8$ .

147 PL = Print length (distance from the heel to the third toe).  
148 TS = Toe extension (distance from first to fifth toe)  
149 ITS = Spread of the middle finger (distance from the second to the fourth toe).  
150 E = Paw of the experiment.  
151 N = Contralateral leg.

152

### 153 **Biochemical analisis in skeletal muscle**

#### 154 **Oxidized dichlorofluorescein (DCF-RS) levels**

155 DCF-RS levels were measured according to MYHRE (15) with some  
156 modifications. Aliquots of skeletal muscle homogenate (50 µL) were added to a  
157 medium containing Tris-HCl buffer (10 mM; pH 7.4) and DCFH-DA (1µM). After  
158 DCFH-DA addition, the medium was incubated in the dark for 1h until fluorescence  
159 measurement procedure (excitation at 488nm and emission at 525 nm and both slit  
160 widths used were at 1,5 nm). DCF-RS levels were determined using a standard curve of  
161 DCF-RS and the results were corrected by mg of protein.

#### 162 **Measurement of 3-(4,5-Dimethylthiazol - 2 -ye)- 2,5- Diphenyl-** 163 **tetrazolium Bromide (MTT) reduction levels**

164 Aliquots of skeletal muscle homogenate (90 µL) were added to a medium  
165 containing 1 mg/mL of MTT and were incubated in the dark for 60 min at 37 °C. Then,  
166 900 µL of DMSO were added. Formazan levels were measured spectrophotometrically  
167 at 570 nm and 630nm and results were corrected by the protein content as proposed by  
168 Mosmann (16).

#### 169 **Thiobarbituric Acid Reactive Substances (TBARS)**

170           TBARS levels were determined according to the method described by Ohkawa  
171       et al. Aliquots of 500 µL of supernatant fraction obtained after blood sample  
172       precipitation or 200 µL of skeletal muscle S1 were added to color reaction. TBARS  
173       levels were measured at 532 nm using a standard curve of MDA and corrected by the  
174       protein content.

175       **Catalase (CAT)**

176           Catalase enzyme activity was measured in the skeletal muscle homogenate  
177       according to the method described by Aebi. A sample of skeletal muscle was added in a  
178       medium containing potassium phosphate buffer (TFK 50 mM, pH 7.4) and H<sub>2</sub>O<sub>2</sub> (1  
179       mM). The CAT kinetic analysis was initiated after the hydrogen peroxide  
180       H<sub>2</sub>O<sub>2</sub> addition; colorimetric reaction was measured at 240 nm.

181       **Protein quantification**

182           The protein content was estimated by the Bradford method (17) using bovine  
183       serum albumin (BSA) as the standard.

184       **Biochemical analysis in plasma after 15 days**

185       **Activity of the creatine kinase enzyme (CK)**

186           CK activity was determined spectrophotometrically in plasma samples using  
187       diagnostic kits (Labtest Diagnóstica S. A. Lagoa Santa, MG).

188       **Activity of the enzyme lactate dehydrogenase (LDH)**

189           LDH activity was determined spectrophotometrically from muscle tissue  
190       samples by the use of diagnostic kits (Labtest Diagnóstica S. A. Lagoa Santa, MG).

191       **Statistical analysis**

192 Graphpad Prism 5.0 (San Diego, CA, USA) was used for all analyzes. Data were  
193 expressed as mean and standard deviation (SD). Significance was assessed by Two-  
194 Way Variance Analysis (ANOVA) followed by Bonferrni's post-hoc test for behavioral  
195 analysis of mechanical allodynia. The significance of the SFI was evaluated by analysis  
196 of variance MANOVA followed by the Bonferrni post-hoc test. The other results,  
197 significance was assessed by analysis of one-way variance followed by the Tukey post-  
198 hoc test.

199 **Results**

200 **Motor and sensorial behavorial tests**

201 The SFI measurement data are shown in Figure 1A. In this study we observed  
202 that 7 days after the injury there was a significant loss of motor function in the PNI  
203 group when compared to the Ctrl group ( $p > 0.001$ ), indicating that the surgery was  
204 correctly performed. This hypothesis asserts when we compare the PNI group to Sham,  
205 we observed that the PNI group had a significant loss of function when compared to  
206 Sham ( $p > 0.001$ ), thus indicating success in surgery and sham surgery.

207 However, 15 days after the injury there was a recovery of function in the PNI  
208 group compared to Ctrl ( $p > 0.05$ ). This same pattern was demonstrated after 15 days in  
209 the Sham group when compared to PNI after 15 days ( $p > 0.05$ ). We thus suggest that  
210 this fact is reasserted by the intrinsic re-emergence of the nerve.

211 In 7 days after the injury, PNI + Cryo group presented recovery of nerve  
212 function ( $p > 0.001$ ), the result was not statistically significant after 15 days, however, it  
213 is still possible to observe improvement of the function compared to the surgery group.

214 **FIGURE 1A: Walking track analysis to evaluate functional recovery.**  
215 Cryotherapy was performed for 15 consecutive days for 20 minutes, gait alteration was  
216 evaluated 7 and 15 days of ice application. Data are expressed as  $\pm$  SEM and two-way

217 RM ANOVA was performed after the Bonferroni test. N = 6. \* p <0.05 and \*\*\* p  
218 <0.001 vs. ctrl, # p <0.05 and ### p <0.001 vs PNI + Cryo, \$ p <0.05 vs Cryotherapy  
219 group.

220 The results of the sensorial analysis through Von Frey were demonstrated in  
221 Figure 1B. No statistical difference was observed between the groups for nociception.  
222 However, it was possible to observe that the PNI group lowered the threshold  
223 throughout the 15 days while the PNL + Cryo group increased its threshold in the first 7  
224 days and maintained in the next 7 days.

225 **FIGURE 2B: Analysis of the paw withdrawal index after 7 and 15 days of**  
226 **injury and cryotherapy treatment.** Cryotherapy treatment decrease (g) removal paw  
227 after PNI Cryotherapy was performed for 15 consecutive days for 20min, sensitive body  
228 change that was investigated by the Frey test. The basal sensorial limit was performed  
229 prior to the application of cryotherapy and surgery and the sensory alteration was  
230 evaluated after 7 days and 15 days after application. Data are expressed as SEM and  
231 two-way ANOVA was performed after the Bonferroni test. N = 6

232

### 233 **Oxidative stress markers in skeletal muscle**

234 Figure 2 shows the effect of cryotherapy on skeletal muscle levels of DCFH-RS  
235 in rats exposed to sciatic nerve injury. Data are expressed as mean ± SEM (n = 6) and  
236 analyzed by One-Way ANOVA, followed by the Tukey test where appropriate. The PNI  
237 and Sham groups significantly increased DCF-RS levels when compared to the control  
238 group (p> 0.01 and p> 0.05). There was no difference in Cryo and PNI + Cryo groups  
239 when compared to control, but compared to PNI and Sham there was a lower level of  
240 DCF-RS (p> 0.05 and p> 0.01).

241           **FIGURE 2: Effect of Cryotherapy (5° C) on DCF-RS oxidation in**  
242           **gastrocnemius muscle in rats exposed to injury to the sciatic nerve.** The rats were  
243           exposed to CCI and were treated with Cryotherapy (5° C) for 20 min for day in 15 days.  
244           Data are expressed as mean ± SEM (n = 6) and were analyzed by One-Way ANOVA,  
245           followed by Tukey test when appropriated. \*p<0.05 \*\* p < 0,01 compared to Control  
246           group. #p<0.05 ## p < 0,01 compared to PNI + Cryo group. \$ p<0.05 \$\$p<0,01  
247           compared to cryotherapy group.

248           However, oxidative damage measure by Thiobarbituric Acid Reactive  
249           Substances (TBARS) did not demonstrate significantly difference in gastrocnemius  
250           muscle levels with cryotherapy treatment, the results were demonstrated in figure 3.

251           **FIGURE 3: Effect of Cryotherapy on of Thiobarbituric Acid Reactive**  
252           **Substances (TBARS) levels in gastrocnemius muscle in rats exposed to CCI.** The  
253           rats were exposed to surgical injury in sciatic nerve and treated with Cryotherapy (5° C)  
254           for 20 minutos for day in 15 days. Data are expressed as mean ± SEM (n = 6) and were  
255           analyzed by One-Way ANOVA, followed by Tukey test when appropriated.

256           In addition, MTT levels decrease in PNI group compared to Control (p>0.05) in  
257           gastrocnemius muscle after the Sciatic injury (Fig. 4).

258           **FIGURE 4: Effect of Cryotherapy in MTT activity, in gastrocnemius**  
259           **muscle in rats exposed to sciatic nerve injury.** The rats were exposed to PNI in sciatic  
260           nerve and treated with Cryotherapy (5° C) for 20 minutes a day during 15 days. Data  
261           are expressed as mean ± SEM (n = 6) and were analyzed by One-Way ANOVA,  
262           followed by Tukey test when appropriated. \*p<0.05 compared to Control group

263           **Enzymatic antioxidant mechanism (CAT)**

264 Effect of cryotherapy on catalase enzyme activity (CAT) was demonstrated in figure 5.  
265 Fifteen days after the injury, the CAT enzyme activity of the treated group was higher  
266 compared to the control ( $p < 0.05$ ) and surgery ( $p < 0.05$ ).

267 **FIGURE 5: Effect of Cryotherapy in catalase enzymeactivity (CAT) in**  
268 **gastrocnemius muscle exposed to sciatic nerve injury.** The rats were exposed to  
269 surgical injury in sciatic nerve and treated with Cryotherapy (5° C) for 20 minutes a day  
270 during 15 days. Data are expressed as mean  $\pm$  SEM ( $n = 6$ ) and were analyzed by One-  
271 Way ANOVA, followed by Tukey test when appropriated. \* $p < 0.05$  compared to  
272 Control group. # $p < 0.05$  compared to surgery group. \$  $p < 0.05$  compared to cryotherapy  
273 group.

274 **Skeletal muscle injury markers in plasma**

275 Cryotherapy treatment during 15 days after CCI did not alter plasmatic creatine kinase  
276 levels (Fig. 6), this result explained because this enzyme returns basal levels at 7 days  
277 after damage. Nevertheless, cryotherapy application increased LDH activity in  
278 treatment group without injury (Fig. 7).

279 **FIGURE 6: Effect of cryotherapy (5 ° C) on creatine kinase (CK) activity in**  
280 **plasma of rats exposed to sciatic nerve injury.** Rats were exposed to CCI in sciatic  
281 nerve and treated with cryotherapy (5 °C) for 20 minutes for 15 days. Data are  
282 expressed as mean  $\pm$  SEM ( $n = 6$ ) and analyzed by One-Way ANOVA, followed by  
283 Tukey's test when appropriate.

284 **FIGURE 7: Effect of cryotherapy (5 ° C) on lactate dehydrogenase (LDH) enzyme**  
285 **activity in plasma of rats exposed to sciatic CCI.** Rats were exposed to surgical injury  
286 in sciatic nerve and treated with cryotherapy (5 °C) for 20 minutes for 15 days. Data are  
287 expressed as mean  $\pm$  SEM ( $n = 6$ ) and analyzed by One-Way ANOVA, followed by  
288 Tukey's test when appropriate. \*  $p < 0.001$  to the control group.

289 **Discussion**

290 In the present study, we evaluated the cryotherapy (once daily for 20 minutes for  
291 15 days) effects on biochemical and functional alterations in rats submitted to peripheral  
292 nerve injury (PNI) of the sciatic nerve. To the best of our knowledge, there is no other  
293 studies that seek to assess the long-term effects of cryotherapy on the treatment of a  
294 peripheral nerve injury. Through this study, we demonstrated that the application of  
295 cryotherapy for 15 days, determined an improvement of the motor function of the sciatic  
296 nerve associated with significant biochemical but not significant sensorial changes.

297 The SFI evaluates the improvement of motor function, with values varying from  
298 0 (normal nerve function) to 100 (complete loss of nerve function) and is a reliable  
299 measure to evaluate the condition of the sciatic nerve in rats (9).

300 In this study, we observed that rats that underwent peripheral nerve compression  
301 had their motor function impaired (Fig. 1A). This is because with nerve compression the  
302 axonal nervous stimulus conduction and also the blood supply of the nervous fiber are  
303 impaired (5). In addition, the myelin sheath thickness around the fibers could be  
304 damaged after a constriction injury of the nerve leading to an impaired speed of nervous  
305 stimulus conduction. In fact, the impact of traumatic events on nervous fibers  
306 functionality depends on the size and thickness of the myelin sheath, since that its  
307 greater size means a greater neuronal conduction velocity (5,18). For this reason,  
308 compression lesions lead to a reduction in conduction velocity, resulting in severe  
309 physiological consequences. Furthermore, a traumatic nerve damage could result in the  
310 neuronal cell body ruptures with the consequent degeneration of axons distal to the  
311 lesion, determining denervation of target organs (19,20).

312 We observed that the application of cryotherapy PNI rats improved sciatic  
313 function in the first seven days compared to the group that had undergone compression

314 and had not been treated (Fig 1A). We suggest that this phenomenon can happen  
315 because the peripheral nervous system is highly temperature sensitive (18). When a  
316 cooling process occurs, there is an increase in the threshold of excitation of the nerve  
317 cells as a function of time, which is inversely proportional to the decrease of the  
318 impulse. Therefore, the sensory nerves react with an increase in the duration of the  
319 action potential leading to the depolarization of the fibers and consequently the decrease  
320 of pain (21). In another study investigating the effects of temperature on peripheral  
321 nerves, the authors observed that, when temperature increased, the duration, distal  
322 latency, area and amplitude of the sensory nerve action potential decreased, increasing  
323 the neuronal stimulus conduction velocity (22).

324 In contrast, when we analyzed the gait functionality after 15 days, we observed  
325 that the PNI + Cryo was not able to bring about significant improvement of the sciatic  
326 function, however, there was an improvement of the PNI group function. We report that  
327 this improvement was probably due to the fact that the time threshold was higher  
328 resulting in intrinsic nerve regeneration, which may have influenced the result when  
329 compared to the treated group. This finding corroborates with other studies that report  
330 that from the 3rd week after nerve compression the regeneration process begins and  
331 consequently the neuronal function improves (8).

332 In this study, we tried to verify a link between the mechanisms of functional  
333 impairment and biochemical changes indicative of oxidative stress in the skeletal  
334 muscle innervated by the sciatic nerve that underwent the compression injury. Some  
335 studies have reported that the excessive production of reactive oxygen species (ROS),  
336 which characterize an oxidative stress status, is involved in the induction of neuropathic  
337 pain (23,24). It is known that after compression of the sciatic nerve occurs an oxidative /  
338 nitrosative stress, which contributes to the genesis of peripheral neuropathy. Moreover,

339 oxidative stress is recognized as one of the main causes of neural damage, not only due  
340 to increased ROS levels but also due to an impairment of antioxidant systems, which  
341 play an important role in the regeneration of peripheral nerve injury (25–27). The  
342 cryotherapy protocol tested in this study determined a reduction of the oxidative  
343 damage in skeletal muscle as were indicated by a decreased DCF-RS levels production  
344 (Fig. 2).

345 According to Siqueira et al (2016), the application of cryotherapy during the  
346 phase of destruction after a skeletal muscle lesion can reverse the increase of DCF-RS  
347 and TBARS levels (28). In another study involving cryotherapy after a skeletal muscle  
348 damage, the authors suggest that excessive ROS levels could not only induce DCF-RS  
349 formation, but also trigger a complex cascade of reactions leading to lipid peroxidation  
350 (increased levels of TBARS) (10).

351 In agreement with this, we have demonstrated here that cryotherapy attenuated  
352 the increase of DCF-RS levels after PNI in rats (restoring control levels), reinforcing its  
353 antioxidant effects. However, we did not observe a significant lipid peroxidation in  
354 skeletal muscle in response to the PNI.

355 It is known that excessive levels of ROS production can cause changes in the  
356 structure and function of enzymes, and as a consequence, defects in some important  
357 enzymatic antioxidant systems (such as catalase) (10). LDH is a cytosolic key enzyme  
358 involved in the energetic metabolism of several cells, such as the myocite. This enzyme  
359 has the role to catalyze the reversible dehydrogenation of lactate, converting it into  
360 pyruvate (29). Our results demonstrated that there was an increase in plasma LDH  
361 activity in the group that only performed cryotherapy (Fig. 7), indicating an increase in  
362 rupture of myocite membranes and the leakage of this enzyme from cytosol to  
363 circulatory system. We understand that this unexpected result means that cryotherapy,

364 when applied for prolonged periods (20 minutes, daily for 15 days) in situations where  
365 there is no skeletal muscle lesion, can be harmful to myocytes membranes integrity.

366 Mitochondria play an essential role for survival and cellular function, including  
367 energy production, redox control, calcium homeostasis, metabolic and biosynthetic  
368 factors(30). Them are also the main source of ROS, which have a fundamental role in  
369 normal cellular functioning and also in cell death mechanisms (30). In this study we  
370 evaluated the skeletal muscle mitochondria functionality via measuring the  
371 methyltetrazolium salt (MTT) reduction to formazan (Fig. 4). This chemical reaction is  
372 associated with dehydrogenase enzyme activities, which are mainly located in  
373 mitochondria and reflects indirectly its viability. Therefore, a decreased MTT reduction  
374 activity means a reduced cellular viability (31), We observed that rats underwent to the  
375 nerve compression had lower levels of MTT reduction when compared to the control  
376 group animals (Fig 4). The other groups maintained the levels of cellular viability  
377 without statistical difference to control, depicting that cryotherapy could contribute to  
378 maintain the myocyte viability of PNI injured rats.

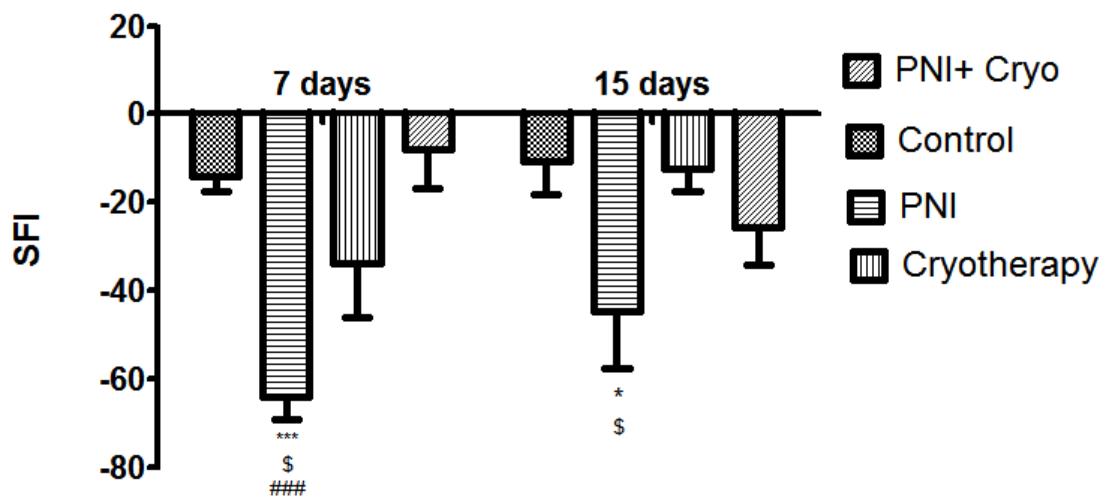
379 The increase in the antioxidant catalase activity of the PNI + Cryo treated rats  
380 suggests that cryotherapy determined an increase the antioxidant status of these animals.  
381 (Fig 5). This result corroborates with the literature that shows different protocols of  
382 cryotherapy are able to increase this enzyme activity (32).

383 The results of this study indicate that the experimental model of PNI in rats  
384 determined a significant impairment of motor behavior that was associated with  
385 significant changes in biochemical markers of oxidative stress. We note that  
386 cryotherapy has shown that it may be a potential treatment resource for pain  
387 management. However, most of the studies interrupt the treatment in the acute phase  
388 after injury, and we do not have clarity of what happens functionally and

389 physiologically after this period. For this reason, we sought to advance the treatment  
 390 period to 15 days in order to study and deepen the effects that cryotherapy may have on  
 391 the peripheral and muscular nervous system and to understand its mechanism of action  
 392 in the treatment of pain, functional aspects and repercussions in the biochemical  
 393 mechanisms.

394

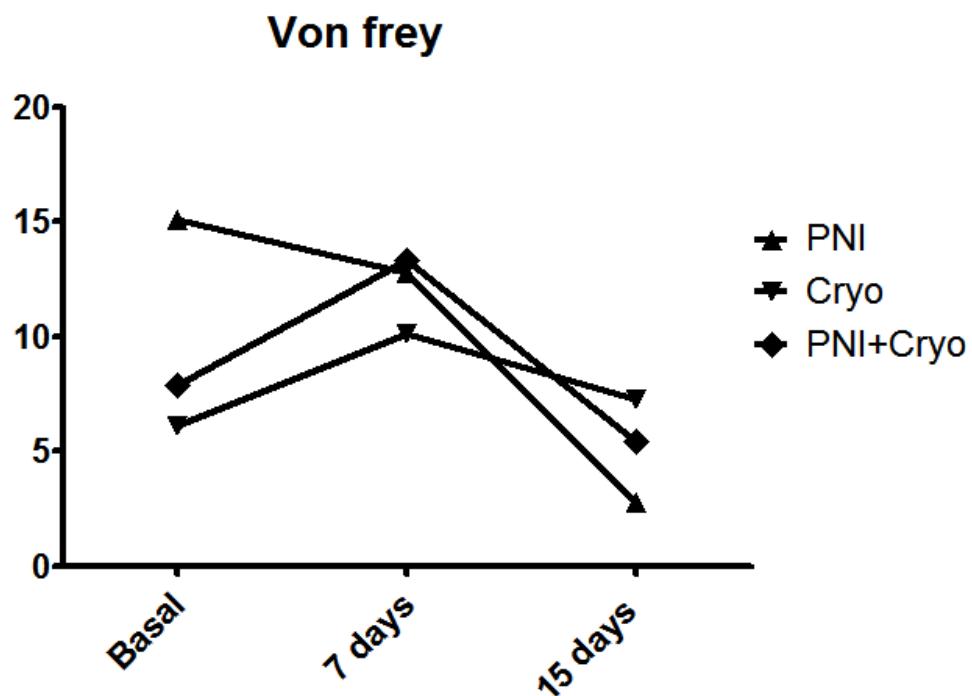
395



396

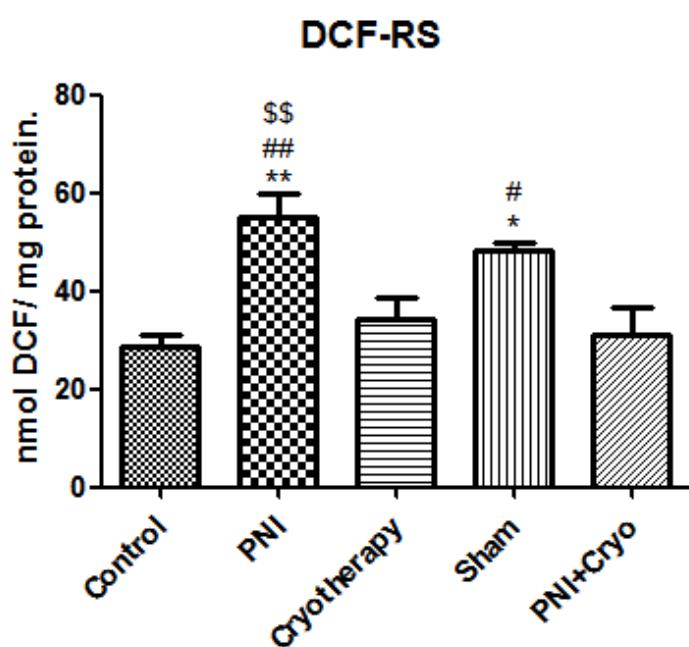
397 Fig 1A

398



399

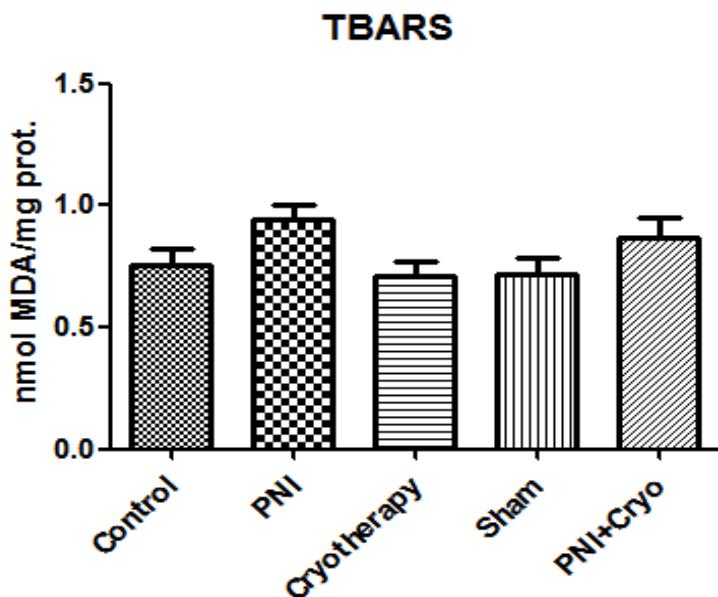
400 Fig 1B



401

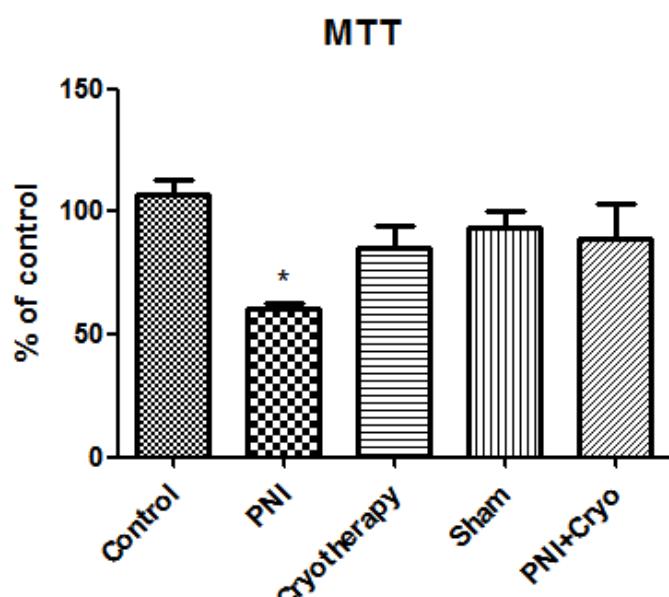
402 Fig 2

403



404

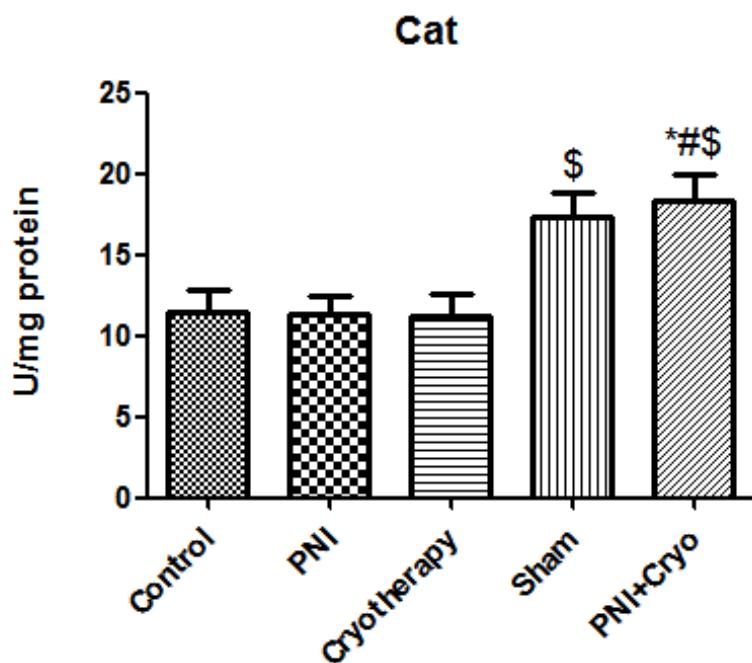
405 Fig 3



406

407 Fig 4

408



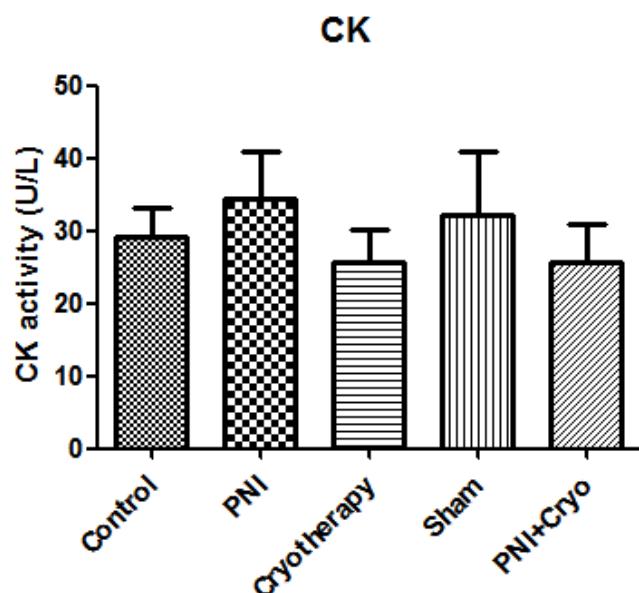
409

410 Fig 5

411

412

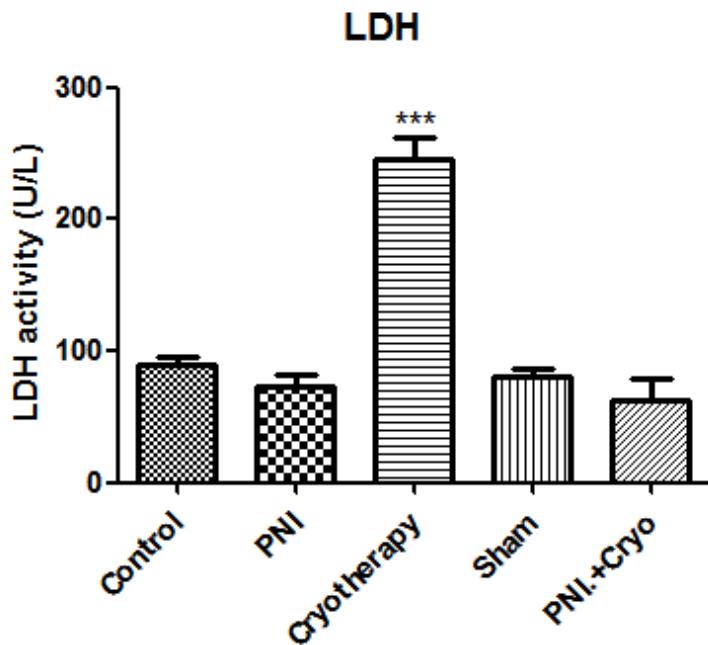
413



414

415

416 Fig 6



417

418

419 Fig 7

420 **Acknowledgments**421 The authors express their gratitude to CAPES, CNPq, PPGRF-UFSM and PPGBTOX –  
422 UFSM.

423

424 **References**

- 425 1. Pardini Freitas P. Reabilitação da mão. In: ATHENEU, editor. Reabilitação da  
426 Mão. 1º. São Paulo-SP; 2006. p. 211–55.
- 427 2. Canclini L, Wallrabe H, Di Paolo A, Kun A, Calliari A, Sotelo-Silveira JR, et al.  
428 Association of Myosin Va and Schwann cells-derived RNA in mammal  
429 myelinated axons, analyzed by immunocytochemistry and confocal FRET  
430 microscopy. Methods. 2014;66(2):153–61.

- 431 3. Faroni A, Mobasseri SA, Kingham PJ, Reid AJ. Peripheral nerve regeneration:  
432 Experimental strategies and future perspectives. *Adv Drug Deliv Rev* [Internet].  
433 2014;82–83:160–7. Available from:  
434 <http://www.ncbi.nlm.nih.gov/pubmed/25446133>
- 435 4. Sundem L, Chris Tseng KC, Li H, Ketz J, Noble M, Elfar J. Erythropoietin  
436 Enhanced Recovery After Traumatic Nerve Injury: Myelination and Localized  
437 Effects. *J Hand Surg Am.* 2016;41(10):999–1010.
- 438 5. Dubový P. Wallerian degeneration and peripheral nerve conditions for both  
439 axonal regeneration and neuropathic pain induction. *Ann Anat.* 2011;193(4):267–  
440 75.
- 441 6. Karvat J, Kakihata CMM, Vieira L, Antunes JS, Ribeiro L de FC, Bertolini GRF.  
442 Evaluation of nociception and edema in experimental sciatic nerve compression  
443 model in Wistar rats treated with cryotherapy. *Rev Dor* [Internet]. 2016;17.  
444 Available from: <http://www.gnresearch.org/doi/10.5935/1806-0013.20160073>
- 445 7. Enwemeka CS, Allen C, Avila P, Bina J, Konrade J, Munns S. Soft tissue  
446 thermodynamics before, during, and after cold pack therapy. *Med Sci Sports*  
447 *Exerc* [Internet]. 2002;34(1):45–50. Available from:  
448 <http://www.ncbi.nlm.nih.gov/pubmed/11782646>
- 449 8. Saueressig F, Xavier LL, Bagatini PB, Nique PS, Da Costa JC, Gomes I, et al.  
450 Morphofunctional analysis of sciatic nerve and motor performance of rats after  
451 cryotherapy with liquid nitrogen. *Oral Surg Oral Med Oral Pathol Oral Radiol.*  
452 2012;113(3):319–26.
- 453 9. de Medinaceli L, Freed WJ, Wyatt RJ. An index of the functional condition of rat  
454 sciatic nerve based on measurements made from walking tracks. *Exp Neurol.*  
455 1982;77(3):634–43.

- 456 10. Puntel GO, Carvalho NÉLR, Amaral GP, Lobato LD, Silveira SÉRO,  
457 Daubermann MF, et al. Therapeutic cold : An effective kind to modulate the  
458 oxidative damage resulting of a skeletal muscle contusion.  
459 2011;45(February):133–46.
- 460 11. Carvalho N, Puntel G, Correa P, Gubert P, Amaral G, Morais J, et al. Protective  
461 effects of therapeutic cold and heat against the oxidative damage induced by a  
462 muscle strain injury in rats. *J Sports Sci.* 2010;28(9):923–35.
- 463 12. Bennett GJ, Xie YK. A peripheral mononeuropathy in rat that produces disorders  
464 of pain sensation like those seen in man. *Pain.* 1988;33(1):87–107.
- 465 13. Dixon WJ. Efficient Analysis of Experimental Observations. *Annu Rev  
466 Pharmacol Toxicol [Internet].* 1980;20(1):441–62. Available from:  
467 <http://www.annualreviews.org/doi/10.1146/annurev.pa.20.040180.002301>
- 468 14. Bain JR, Mackinnon SE, Hunter DA. Functional Evaluation of Complete Sciatic,  
469 Peroneal, and Posterior Tibial Nerve Lesions in the Rat. *Plast Reconstr Surg  
470 [Internet].* 1989;83(1):129–36. Available from:  
471 <http://content.wkhealth.com/linkback/openurl?sid=WKPTLP:landingpage&an=0006534-198901000-00024>
- 473 15. Myhre O, Andersen JM, Aarnes H, Fonnum F. Evaluation of the probes 2',7'-  
474 dichlorofluorescin diacetate, luminol, and lucigenin as indicators of reactive  
475 species formation. Vol. 65, *Biochemical Pharmacology.* 2003. p. 1575–82.
- 476 16. Tim M, Mosmann T. Rapid colorimetric assay for cellular growth and survival:  
477 application to proliferation and cytotoxicity assays. *J Immunol Methods  
478 [Internet].* 1983;65(1–2):55–63. Available from:  
479 <http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&do>  
480 pt=Citation&list\_uids=6606682%5Cn<http://www.sciencedirect.com/science/article>

- 481 le/pii/0022175983903034
- 482 17. Bradford MM. A rapid and sensitive method for the quantitation of microgram  
483 quantities of protein using the principle of protein dye binding. *Anal Biochem.*  
484 1976;72:248–54.
- 485 18. Rutkove SB. Effects of temperature on neuromuscular electrophysiology. Vol.  
486 24, *Muscle and Nerve*. 2001. p. 867–82.
- 487 19. Rosso G, Young P, Shahin V. Implications of Schwann Cells Biomechanics and  
488 Mechanosensitivity for Peripheral Nervous System Physiology and  
489 Pathophysiology. *Front Mol Neurosci [Internet]*. 2017;10. Available from:  
490 <http://journal.frontiersin.org/article/10.3389/fnmol.2017.00345/full>
- 491 20. Li R, Wu J, Lin Z, Nangle MR, Li Y, Cai P, et al. Single injection of a novel  
492 nerve growth factor coacervate improves structural and functional regeneration  
493 after sciatic nerve injury in adult rats. *Exp Neurol*. 2017;288:1–10.
- 494 21. Guirro R, Abib C, Máximo C. Os Efeitos Fisiológicos da Crioterapia: uma  
495 Revisão [Internet]. Vol. 6, *Fisioterapia e Pesquisa*. 1999. p. 164–70. Available  
496 from: <http://www.revistas.usp.br/fpusp/article/view/79629>
- 497 22. Ashworth NL, Marshall SC, Satkunam LE. The effect of temperature on nerve  
498 conduction parameters in carpal tunnel syndrome. *Muscle Nerve [Internet]*.  
499 1998;21(8):1089–91. Available from:  
500 <http://www.ncbi.nlm.nih.gov/pubmed/9655132>
- 501 23. Kim D, You B, Jo E-K, Han S-K, Simon MI, Lee SJ. NADPH oxidase 2-derived  
502 reactive oxygen species in spinal cord microglia contribute to peripheral nerve  
503 injury-induced neuropathic pain. *Proc Natl Acad Sci [Internet]*.  
504 2010;107(33):14851–6. Available from:  
505 <http://www.pnas.org/cgi/doi/10.1073/pnas.1009926107>

- 506 24. Komirishetty P, Areti A, Gogoi R, Sistla R, Kumar A. Combination strategy of  
507 PARP inhibitor with antioxidant prevent bioenergetic deficits and inflammatory  
508 changes in CCI-induced neuropathy. *Neuropharmacology*. 2017;113:137–47.
- 509 25. Lanza C, Raimondo S, Vergani L, Catena N, Sénès F, Tos P, et al. Expression of  
510 antioxidant molecules after peripheral nerve injury and regeneration. *J Neurosci  
511 Res*. 2012;90(4):842–8.
- 512 26. Qiu T, Yin Y, Li B, Xie L, Yan Q, Dai H, et al. PDLLA/PRGD/??-TCP conduits  
513 build the neurotrophin-rich microenvironment suppressing the oxidative stress  
514 and promoting the sciatic nerve regeneration. *J Biomed Mater Res - Part A*.  
515 2014;102(10):3734–43.
- 516 27. Valko M, Leibfritz D, Moncol J, Cronin MTD, Mazur M, Telser J. Free radicals  
517 and antioxidants in normal physiological functions and human disease. *Int J  
518 Biochem Cell Biol* [Internet]. 2007;39(1):44–84. Available from:  
519 <http://linkinghub.elsevier.com/retrieve/pii/S1357272506002196>
- 520 28. Siqueira AF, Vieira A, Ramos GV, De R, Marqueti C, Salvini TDF, et al.  
521 Communications in Free Radical Research Multiple cryotherapy applications  
522 attenuate oxidative stress following skeletal muscle injury. *Redox Rep* [Internet].  
523 2016;0(0):1–7. Available from:  
524 <http://dx.doi.org/10.1080/13510002.2016.1239880>
- 525 29. Zheng Y Bin, Wang Z, Chen BY, Wang XC. Multiple effects of chemical reagent  
526 on enzyme: o-phthalaldehyde-induced inactivation, dissociation and partial  
527 unfolding of lactate dehydrogenase from pig heart. *Int J Biol Macromol*.  
528 2003;32(3–5):191–7.
- 529 30. Echtay KS. Mitochondrial uncoupling proteins — What is their physiological  
530 role ? 2007;43:1351–71.

- 531 31. Siqueira AF, Vieira A, Ramos GV, Marqueti R de C, Salvini T de F, Puntel GO,  
532 et al. Multiple cryotherapy applications attenuate oxidative stress following  
533 skeletal muscle injury. Redox Rep. 2017;22(6):323–9.
- 534 32. Skrzep-poloczek B, Romuk E, Wi B, Owczarek AJ, Chor P, Siero A, et al. Effect  
535 of Whole-Body Cryotherapy on Antioxidant Systems in Experimental Rat  
536 Model. 2017;2017.
- 537

### **3. CONCLUSÃO**

Ao planejar este estudo, objetivou-se preencher lacunas deixadas por pesquisas previamente realizadas. A maioria dos estudos que envolvem a crioterapia interrompem o tratamento durante a fase aguda após a lesão, e não temos clareza do que acontece funcionalmente e fisiologicamente após esse período. Os resultados deste estudo indicam que o modelo experimental de compressão nervosa em ratos determinou um comprometimento significativo do comportamento motor associado a mudanças significativas nos marcadores bioquímicos do estresse oxidativo. Notamos que a crioterapia durante um limiar de tempo maior pode ser um recurso de tratamento potencial para o tratamento da dor. Por essa razão, procuramos avançar o período de tratamento para 15 dias, a fim de estudar e aprofundar os efeitos que a crioterapia pode ter sobre o sistema nervoso periférico e muscular e compreender seu mecanismo de ação no tratamento da dor, aspectos funcionais e repercussões nos mecanismos bioquímicos.

## REFÉRENCIAS BIBLIOGRÁFICAS:

BENNETT, Gary J.; XIE, Y. K. A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. **Pain**, [s. l.], v. 33, n. 1, p. 87–107, 1988.

BENNETT, Michael I. et al. Can pain can be more or less neuropathic? Comparison of symptom assessment tools with ratings of certainty by clinicians. **Pain**, [s. l.], v. 122, n. 3, p. 289–294, 2006.

BISWAS, Subrata Kumar. **Does the Interdependence between Oxidative Stress and Inflammation Explain the Antioxidant Paradox?Oxidative Medicine and Cellular Longevity**, 2016.

BRANCACCIO, Nelson et al. **Fisioterapia em Movimento**. [s.l.] : PUCPR, 2005. v. 18DÂNGELO, J. G.; FANTTINI, C. A. **Anatomia Humana Sistêmica e Segmentar**. São Paulo: Editora Ateneu, p. 52-61, 2000.

DETLOFF, Megan Ryan et al. Acute exercise prevents the development of neuropathic pain and the sprouting of non-peptidergic (GDNF- and artemin-responsive) c-fibers after spinal cord injury. **Experimental neurology**, [s. l.], v. 255, p. 38–48, 2014.

DUBOVÝ, Petr. Wallerian degeneration and peripheral nerve conditions for both axonal regeneration and neuropathic pain induction. **Annals of Anatomy**, [s. l.], v. 193, n. 4, p. 267–275, 2011.

FARIAS, Rafaela Soares et al. O Uso da Tens, Crioterapia e Criotens na Resolução da Dor The Use of Tens, cryotherapy and Criotens in the Resolution of Pain. **Revista Brasileira de Ciências da Saúde**, [s. l.], v. 14, n. 1, p. 27–36, 2010.

FERRIGNO, I. S. V.; FREITAS, P. P.; FREITAS, A. D. Lesões dos Nervos Periféricos. In: FREITAS, P. P. **Reabilitação da mão**. São Paulo: Ed Ateneu, p. 211-230, 2005.

GIARDINI, Aline Carolina et al. Neural Mobilization Treatment Decreases Glial Cells and Brain-Derived Neurotrophic Factor Expression in the Central Nervous System in Rats with Neuropathic Pain Induced by CCI in Rats. **Pain Research and Management**, [s. l.], v. 2017, 2017.

GIRRO, Rinaldo; ABIB, Carla; MÁXIMO, Carla. **Os Efeitos Fisiológicos da Crioterapia: uma RevisãoFisioterapia e Pesquisa**, 1999.

GUTIÉRREZ ESPINOZA, H. J.; LAVADO BUSTAMANTE, I. P.; MÉNDEZ PÉREZ, S. J. **Revisión sistemática sobre el efecto analgésico de la crioterapia en el manejo del dolor de origen músculo esqueléticoRevista de la Sociedad Espanola del Dolor**, 2010.

HOOSHMAND, H.; HASHMI, M.; EM, Phillips. Cryotherapy can cause permanent nerve damage: a case report. **American Journal of Pain Management**, [s. l.], v. 14, n. 2, p. 63–70, 2004. Disponível em:

HSU, Michael; STEVENSON, Fang F. Wallerian degeneration and recovery of motor nerves after multiple focused cold therapies. **Muscle and Nerve**, [s. l.], v. 51, n. 2, p. 268–275, 2015.

IDE, C. Peripheral nerve regeneration. **Neuroscience research**, [s. l.], v. 25, n. 2, p. 101–21, 1996.

JU, Hui et al. The potential role of nerve growth factor in cryoneurolysis-induced neuropathic pain in rats. **Cryobiology**, [s. l.], v. 65, n. 2, p. 132–138, 2012.

KARVAT, Jhenifer et al. Evaluation of nociception and edema in experimental sciatic nerve compression model in Wistar rats treated with cryotherapy. **Revista Dor**, [s. l.], v. 17, 2016.

KOMIRISHETTY, Prashanth et al. Morin Mitigates Chronic Constriction Injury (CCI)-Induced Peripheral Neuropathy by Inhibiting Oxidative Stress Induced

PARP Over-Activation and Neuroinflammation. **Neurochemical research**, [s. l.], v. 41, n. 8, p. 2029–42, 2016.

KRAYCHETE, Durval Campos; GOZZANI, Judymara Lauzi; KRAYCHETE, Angiolina Campos. Dor neuropática: aspectos neuroquímicos. **Revista Brasileira de Anestesiologia**, [s. l.], v. 58, n. 5, p. 492–505, 2008.

LANZA, Cristina et al. Expression of antioxidant molecules after peripheral nerve injury and regeneration. **Journal of Neuroscience Research**, [s. l.], v. 90, n. 4, p. 842–848, 2012.

LEE, Yoon-Mi; SONG, Byeng Chun; YEUM, Kyung-Jin. Impact of Volatile Anesthetics on Oxidative Stress and Inflammation. **BioMed Research International**, [s. l.], v. 2015, p. 1–8, 2015.

MEDZHITOV, Ruslan. **Origin and physiological roles of inflammation**. Nature, 2008.

MERSKEY, H.; BOGDUK (EDS), N. Classification of Chronic Pain. Descriptions of Chronic Pain Syndromes and Definitions of Pain Terms. **IASP Press**, [s. l.], p. i–xvi, 1994.

QIU, Tong et al. PDLLA/PRGD/??-TCP conduits build the neurotrophin-rich microenvironment suppressing the oxidative stress and promoting the sciatic nerve regeneration. **Journal of Biomedical Materials Research - Part A**, [s. l.], v. 102, n. 10, p. 3734–3743, 2014.

ROCHA, Anita Perpétua Carvalho et al. Dor: aspectos atuais da sensibilização periférica e central. **Revista Brasileira de Anestesiologia**, [s. l.], v. 57, n. 1, p. 94–105, 2007.

RODRÍGUEZ, F. J.; VALERO-CABRÉ, A.; NAVARRO, X. Regeneration and functional recovery following peripheral nerve injury. **Drug Discovery Today: Disease Models**, [s. l.], v. 1, n. 2, p. 177–185, 2004.

ROSSO, Gonzalo; YOUNG, Peter; SHAHIN, Victor. Implications of Schwann Cells Biomechanics and Mechanosensitivity for Peripheral Nervous System Physiology and Pathophysiology. **Frontiers in Molecular Neuroscience**, [s. l.], v. 10, 2017.

RUTKOVE, Seward B. **Effects of temperature on neuromuscular electrophysiology**. Muscle and Nerve, 2001.

SCHESTATSKY, Pedro. Definição, Diagnóstico e Tratamento da Dor Neuropática. **Revista HCPA**, [s. l.], v. 28, n. 3, 2009.

TEIXEIRA, M.J. **Fisiopatologia da Dor Neuropática**. In: Teixeira M.J..Dor: Contextos Interdisciplinar. Curitiba: Ed.Maio, 2003, p. 155-69.

UCHÔA, S. M. M.; FREITAS, P. P.; Modalidades Terapêuticas na Reabilitação da Mão. In: FREITAS , P. P. **Reabilitação da Mão**, São Paulo: Ed Ateneu, p. 211-230, 2005.

VALKO, Marian et al. Free radicals and antioxidants in normal physiological functions and human disease. **The International Journal of Biochemistry & Cell Biology**, [s. l.], v. 39, n. 1, p. 44–84, 2007.

ZHANG, Xiu Xiu et al. Short-term observations of the regenerative potential of injured proximal sensory nerves crossed with distal motor nerves. **Neural Regeneration Research**, [s. l.], v. 12, n. 7, p. 1172–1176, 2017.