

ARTICLE IN PRESS

V.D.G. Sinhorin et al. / Brain Research xx (2005) xxx–xxx

3

148 to 2 ml with type I ultrapure water and heated at 95 °C for 90
 149 min in a water bath using a glass ball as a condenser. After
 150 cooling to room temperature, absorbance was measured in
 151 the supernatant at 532 nm [29].

152 2.5. Protein carbonyl assay ex vivo

153 Immediately after the behavioral evaluation, the animals
 154 were killed by decapitation and had their forebrain removed.
 155 Tissues were homogenized in 10 volumes (w/v) of 10 mM
 156 Tris-HCl buffer, pH 7.4, using a glass homogenizer and its
 157 carbonyl protein content was determined by the method
 158 described by Yan et al. [44], adapted for brain tissue, as
 159 below. Briefly, homogenates were diluted to 750–800 µg/ml
 160 of protein in each sample, and 1 ml of aliquots was mixed
 161 with 0.2 ml of 2,4-dinitrophenylhydrazine (DNPH, 10 mM)
 162 or 0.2 ml HCl (2 M). After incubation at room temperature
 163 for 1 h in a dark ambient, 0.6 ml of denaturing buffer (150
 164 mM sodium phosphate buffer, pH 6.8, containing 3% SDS),
 165 1.8 ml of heptane (99.5%) and 1.8 ml of ethanol (99.8%)
 166 were added sequentially, and mixed with vortex agitation for
 167 40 s and centrifuged for 15 min. Next, the protein isolated
 168 from the interface was washed two times with 1 ml of ethyl
 169 acetate/ethanol 1:1 (v/v) and suspended in 1 ml of
 170 denaturing buffer. Each DNPH sample was read at 370
 171 nm in a Hitachi U-2001 spectrophotometer against the
 172 corresponding HCl sample (blank), and total carbonylation
 173 calculated using a molar extinction coefficient of 22,000
 174 M⁻¹cm⁻¹, as described by Levine et al. [22].

175 2.6. Protein determination

176 Protein content was measured by the method of Bradford
 177 [4] using bovine serum albumin (1 mg/ml) as standard.

178 2.7. Statistical analysis

179 Behavioral and biochemical data were analyzed by one-
 180 way or two-way analysis of variance (ANOVA), depending
 181 on the experimental design. Post hoc analysis was carried
 182 out by the Duncan multiple range test, when appropriate. A
 183 P value less than 0.05 was considered significant.

184 3. Results

185 Fig. 1A shows the effect of increasing doses of succinate
 186 (0.7, 1.0 and 1.7 µmol/5 µl icv) on the content of TBARS in
 187 forebrain ex vivo. Statistical analysis (one-way ANOVA
 188 followed by Duncan multiple range test) revealed that while
 189 the intracerebroventricular administration of 0.7 and 1.0
 190 µmol succinate increased TBARS content compared with
 191 the control group ($F_{(3,23)} = 29.76$; $P < 0.0001$), 1.7 µmol
 192 succinate had no effect on forebrain TBARS content. The
 193 effect of increasing doses of succinate (0.7, 1.0 and 1.7
 194 µmol/5 µl icv) on total protein carbonyl content ex vivo is

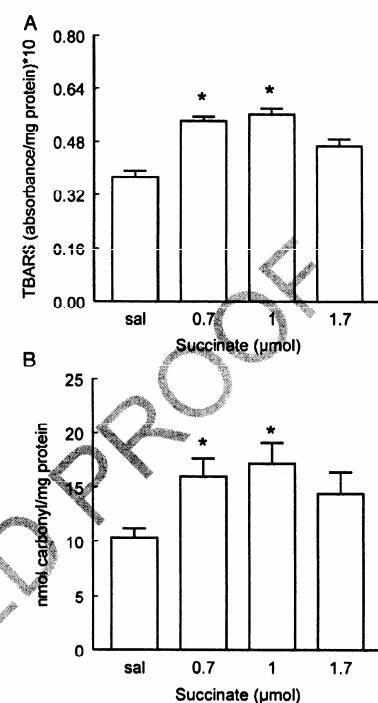


Fig. 1. (A) Effect of succinate administration (icv) on TBARS content ex vivo. * $P < 0.0001$ compared with control by Duncan multiple range test. Data are the means \pm SEM; $n = 6$ –7 animals in each group. (B) Effect of succinate administration (icv) on total protein carbonylation content ex vivo. * $P < 0.05$ compared with control by Duncan multiple range test. Data are the means \pm SEM; $n = 6$ –7 animals in each group.

shown in Fig. 1B. Statistical analysis (one-way ANOVA followed by analysis Duncan multiple range test) revealed that while the intracerebroventricular administration of 0.7 and 1.0 µmol succinate increased total protein carbonyl content compared with the control group ($F_{(3,23)} = 3.26$; $P < 0.05$), 1.7 µmol succinate had no effect on forebrain total protein carbonyl content. The effect of succinate on the locomotor behavior of the animals was also investigated. Statistical analysis (one-way ANOVA followed by Duncan multiple range test) revealed that intracerebroventricular administration of 0.7 and 1.0 µmol succinate decreased the number of areas crossed ($F_{(3,23)} = 7.58$; $P < 0.001$), the number of rearing responses ($F_{(3,23)} = 4.62$; $P < 0.01$) and increased time spent in immobility ($F_{(3,23)} = 13.65$; $P < 0.0001$) compared with the control group (Table 1).

The involvement of NMDA receptors in the increase of total protein carbonyl content induced by succinate was assessed by co-administrating MK-801 (7 nmol/2.5 µl icv), a noncompetitive NMDA receptor antagonist, with succinate (1.0 µmol/2.5 µl icv). Statistical analysis (two-way ANOVA) of total protein carbonyl content data revealed a

ARTICLE IN PRESS

4

V.D.G. Sinhorin et al. / Brain Research xx (2005) xxx–xxx

t1.1	Table 1 Effect of succinate injection (0.7, 1.0 and 1.7 μ mol/5 μ l icv) on exploratory activity of mice			
t1.2	Treatment	Rearing responses	Number of crossing	Immobility time
t1.3	0.9% NaCl	39.42 T 12.00	188.42 T 20.47	124.50 T 19.86
t1.4	0.9% NaCl + succinate (μ mol)	4.71 T 4.06*	31.57 T 22.99*	516.14 T 1.02*
t1.5	0.7	7.50 T 2.76*	81.33 T 29.51*	386.66 T 52.90*
t1.6	1.0	23.00 T 6.44	121.57 T 25.61	171.17 T 77.60
t1.7	1.7			

t1.9 Data are the means T SEM for $n = 6$ –7 in each group.t1.10 * $P < 0.01$.t1.11 * $P < 0.0001$ compared with 0.9% NaCl group (Duncan multiple range test).

216 significant pretreatment (0.9% NaCl or 1.0 μ mol succinate)
 217 by treatment (0.9% NaCl or 7 nmol MK-801) interaction
 218 ($F_{(3,36)} = 9.545$; $P < 0.01$, Fig. 2), suggesting the involve-
 219 ment of NMDA receptors in the protein carbonylation
 220 induced by succinate.

221 Statistical analysis (two-way ANOVA) of the number
 222 of crossing responses and time spent in immobility in the
 223 open field revealed a significant pretreatment (0.9% NaCl
 224 or 1.0 μ mol succinate) by treatment (0.9% NaCl or 7
 225 nmol MK-801) interaction: ($F_{(1,38)} = 4.61$; $P < 0.05$ and
 226 $F_{(1,38)} = 4.99$; $P < 0.05$), respectively, since MK-801
 227 prevented the inhibitory effect of succinate on locomotor
 228 activity (Table 2).

229 4. Discussion

230 In the present study, we showed, for the first time, that
 231 the intracerebroventricular administration of succinate
 232 decreases exploratory activity and increases forebrain
 233 thiobarbituric acid reactive substances (TBARS) and protein
 234 carbonyl content biphasically. Interestingly, succinate doses
 235 (0.7 and 1.0 μ mol), which decreased locomotor activity, also
 236 increased the biochemical markers of oxidative damage.

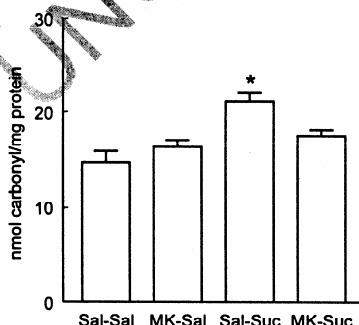


Fig. 2. Effect of MK-801 administration (7 nmol/2.5 μ l icv) on succinate-induced (1.0 μ mol/2.5 μ l) total protein carbonylation content ex vivo. * $P < 0.01$ compared with 0.9% NaCl plus 0.9% NaCl by Duncan multiple range test. Data are the means T SEM; $n = 10$ –11 animals in each group.

t2.1	Table 2 Effect of MK-801 (7 nmol/2.5 μ l icv) on succinate-induced (1 μ mol/2.5 μ l icv) exploratory behavior in mice			
t2.2	Treatment	Rearing responses	Number of crossing	Immobility time
t2.3	0.9% NaCl + 0.9% NaCl	27.27 T 4.80	90.45 T 12.80	164.36 T 32.53
t2.4	0.9% NaCl + succinate	17.30 T 5.38	61.40 T 14.74	249.40 T 40.85
t2.5	MK-801 + 0.9% NaCl	14.72 T 3.21	71.63 T 14.58	267.45 T 37.89
t2.6	MK-801 + succinate	20.90 T 8.91	129.40 T 33.59	188.60 T 35.01

t2.8 Data are the means T SEM for $n = 10$ –11 in each group.

MK-801 prevented the increase of total protein carbonylation and the decrease of locomotor activity induced by 1.0 μ mol succinate.

The currently reported succinate-induced increase in TBARS production ex vivo suggests that this organic acid causes oxidative damage, because TBA adducts are mainly formed from the reaction of lipid-derived MDA with TBA [29]. Accordingly, succinate increased total protein carbonylation, a relatively specific marker of oxidative damage. These findings strongly suggest that 0.7 and 1.0 μ mol succinate induce oxidative damage. However, since we did not test doses of succinate lower than 0.7 μ mol, we cannot rule out that even lower amounts of succinate do not cause oxidative damage. Conversely, a high dose of succinate (1.7 μ mol) returned TBARS and total protein carbonyl content to control levels, characterizing a biphasic effect of succinate on the studied oxidation markers. The molecular mechanisms underlying the currently reported antioxidant effect of 1.7 μ mol succinate (see Figs. 1A and B) are not established, but it is interesting that a similar biphasic effect of succinate on fEPSPs has been reported. While low concentrations (0.3–1.0 mM) of succinate increase neuronal fEPSPs in hippocampal slices by NMDA-mediated mechanisms, high concentrations of succinate (3–10 mM) decrease fEPSPs and reverse the increase of fEPSPs induced by low concentrations of succinate [33]. In that study, the biphasic effect on fEPSPs was attributed to a putative biphasic effect of succinate on the NMDA receptor. Given the marked similar pattern of variation of the currently reported dose-effect curve for succinate and that reported by Roehrs et al. [33], one might propose that they might occur by similar mechanisms, i.e., NMDA receptors. Therefore, we decided to investigate whether NMDA receptors were involved in the succinate-induced oxidative damage. The co-administration of MK-801, an noncompetitive NMDA receptor antagonist, protected against succinate-induced protein carbonylation increase, suggesting the involvement of this subtype of glutamate receptors in the oxidative damage induced by succinate.

One remarkable finding in this study is that succinate-induced oxidative damage was accompanied by significant behavioral alterations, whose pattern of variation along succinate doses coincided with the markers of oxidative damage. More specifically, succinate caused forebrain oxidative damage markers increase, which coincided with

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

457

ARTICLE IN PRESS

V.D.G. Sinhorin et al. / Brain Research xx (2005) xxx–xxx

5

282 a decrease in locomotor activity. Accordingly, increasing
 283 succinate doses returned locomotor activity scores and
 284 oxidative damage markers to control levels. These inter-
 285 esting results reveal a parallelism between these events and
 286 tempt us to propose that some relationship may exist
 287 between them. Additional evidences that oxidative damage
 288 increase and locomotor activity decrease are somehow
 289 related come from the experiment that showed that MK-
 290 801 prevents succinate effects on both a biochemical and a
 291 behavioral parameter (Fig. 2). Since pharmacological
 292 treatment affected both behavioral and biochemical param-
 293 eters in the same direction, to a similar extent, and did not
 294 dissociate them, it is rather possible that the currently
 295 reported biochemical and behavioral effects of succinate
 296 are related. However, if there is a cause–effect relationship
 297 between them cannot be established at the moment, and a
 298 more detailed analysis of the variation of cerebral
 299 oxidative stress markers levels and convulsive activity
 300 along time has to be performed, in order to determine
 301 whether a cause–effect relationship between these events
 302 does exist. In addition, we did not assess if succinate
 303 increases oxidative damage markers other than TBARS
 304 and protein carbonyl. Therefore, it remains to be deter-
 305 mined if other important cellular targets for oxidative
 306 damage, such as DNA, are affected by succinate, and in
 307 what extent our results comparatively apply to the human
 308 condition, because species-specific susceptibility is possi-
 309 ble and succinate concentrations in the white matter are not
 310 known.

311 Regardless if the currently reported behavioral and
 312 neurochemical effects of succinate are causally related to
 313 each other or not, this study shows, for the first time, that
 314 succinate causes significant oxidative damage and behav-
 315 ioral effects at subconvulsant doses by NMDA receptor-
 316 mediated mechanisms. This is an important finding for
 317 different areas, particularly those concerned with the study
 318 of the succinate-accumulating conditions, such as expo-
 319 sure to exogenous [1,7,17,19,32,40] and endogenous
 320 [10,9,12–14,18,24,34,38] SDH inhibitor toxins and
 321 inherited deficiencies of SDH [5,6,14,30,31], because it
 322 shows that at least some of the deleterious effects of these
 323 conditions may arise from secondary succinate accumu-
 324 lation, a possibility that has been overlooked in the
 325 literature.

326 Acknowledgments

327 Research supported by CNPq (grant: 500120/2003-0 and
 328 500096/2003-1) and CAPES. Carlos Fernando de Mello and
 329 Maribel Antonello Rubin are the recipients of CNPq
 330 fellowships (grant: 500120/2003-0 and 500096/2003-1).
 331 Valéria Dornelles Gindri Sinhorin and Juliana Saibt Martins
 332 Pasin are the recipients of CAPES fellowships. We thank
 333 Florindo Gonçalves Duarte and Dinah Barbara Pathek for
 334 capable technical assistance.

References

- | | |
|---|--|
| <p>[1] T. Alexi, P.E. Hughes, R.L.M. Faull, C.E. Williams, 3-Nitropropionic acid's lethal triplet: cooperative pathways of neurodegeneration, <i>NeuroReport</i> 9 (1998) 59–64.</p> <p>[2] M.F. Beal, Mitochondria, free radicals, and neurodegeneration, <i>Curr. Opin. Neurobiol.</i> 6 (1996) 661–666.</p> <p>[3] M.F. Beal, Energetics in the pathogenesis of neurodegenerative diseases, <i>Trends Neurosci.</i> 23 (2000) 298–304.</p> <p>[4] M.M. Bradford, A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding, <i>Anal. Biochem.</i> 72 (1976) 248–254.</p> <p>[5] J. Brismar, P.T. Ozand, CT and MR of the brain in the diagnosis of organic acidemias: experiences from 107 patients, <i>Brain Dev.</i> 16 (1994) 104–124.</p> <p>[6] K. Brockmann, A. Bjørnstad, P. Dechent, C.G. Korenke, J. Smeitink, F. Trijbels, S. Athanasiopoulos, R. Villagran, O.H. Skjeldal, E. Wilichowski, J. Frahm, F. Hanefeld, Succinate in dystrophic white matter: a proton magnetic resonance spectroscopy finding characteristic for complex II deficiency, <i>Ann. Neurol.</i> 52 (2002) 38–46.</p> <p>[7] P. Calabresi, P. Gibellini, B. Picconi, D. Centonze, A. Pisani, P. Bonsi, P. Greengard, R.A. Hipskind, E. Borrelli, G. Bernardi, Inhibition of mitochondrial complex II induces a long-term potentiation of NMDA-mediated synaptic excitation in the striatum requiring endogenous dopamine, <i>J. Neurosci.</i> 21 (2001) 5110–5120.</p> <p>[8] W.G. Clark, C.A. Vivona, C.F. Baxter, Accurate freehand injection into a lateral brain ventricle of the conscious mouse, <i>J. Appl. Physiol.</i> 25 (1968) 319–321.</p> <p>[9] C. Davolio, J.T. Greenamyre, Selective vulnerability of the CA1 region of hippocampus to the indirect excitotoxic effects of malonic acid, <i>Neurosci. Lett.</i> 192 (1995) 29–32.</p> <p>[10] C.F. De Mello, J. Begnini, R.E. Jiménez-Bernal, M.A. Rubin, J. de Bastiani, E.J.M. da Costa, M. Wajner, Intrastriatal methylmalonic administration induces rotational behavior and convulsions through glutamatergic mechanisms, <i>Brain Res.</i> 721 (1996) 120–125.</p> <p>[11] J.C. Dutra, C.S. Dutra-Filho, S.E.C. Cardoso, C.M.D. Wannmacher, J.J.F. Sarkis, M. Wajner, Inhibition of succinate dehydrogenase and β-hydroxybutyrate dehydrogenase activities by methylmalonate in brain and liver of developing rats, <i>J. Inher. Metab. Dis.</i> 16 (1993) 147–153.</p> <p>[12] M.R. Fighera, C.M. Queiroz, M.P. Stracke, M.C. Brauer, L.L. Gonzalez-Rodriguez, R. Frussa-Filho, M. Wajner, C.F. de Mello, Ascorbic acid and alpha-tocopherol attenuate methylmalonic acid-induced convulsions, <i>NeuroReport</i> 10 (1999) 2039–2043.</p> <p>[13] M.R. Fighera, J.S. Bonini, T.G. de Oliveira, R. Frussa-Filho, C.S. Dutra-Filho, M.E. Hagen, M.A. Rubin, C.F. Mello, GM1 ganglioside attenuates convulsions and thiobarbituric acid reactive substances production induced by the intrastriatal injection of methylmalonic acid, <i>Int. J. Biochem. Cell Biol.</i> 35 (2003) 465–473.</p> <p>[14] J. Fleck, M.C.P. Ribeiro, C.M. Schneidler, V.D.G. Sinhorin, M.A. Rubin, C.F. Mello, Intrastriatal malonate administration induces convulsive behaviour in rats, <i>J. Inher. Metab. Dis.</i> 24 (2004) 211–219.</p> <p>[15] M.E. Götz, G. Küning, P. Riederer, M.B.H. Youdim, Oxidative stress: free radical production in neural degeneration, <i>Pharmacol. Ther.</i> 63 (1994) 37–122.</p> <p>[16] G.G. Greene, J.T. Greenamyre, Characterization of the excitotoxic potential of the reversible succinate dehydrogenase inhibitor malonate, <i>J. Neurochem.</i> 64 (1995) 430–436.</p> <p>[17] G.G. Greene, J.T. Greenamyre, Manipulation of membrane potential modulates malonate-induced striatal excitotoxicity in vivo, <i>J. Neurochem.</i> 66 (1996) 637–643.</p> <p>[18] J.G. Greene, R.H.P. Porter, R.V. Eller, J.T. Greenamyre, Inhibition of succinate dehydrogenase by malonic acid produces an excitotoxic lesion in rat striatum, <i>J. Neurochem.</i> 61 (1993) 1151–1154.</p> <p>[19] O.B. Hassel, U. Sonnewald, Selective inhibition of the tricarboxylic</p> | <p>335
336</p> <p>337
338</p> <p>339</p> <p>340</p> <p>341</p> <p>342</p> <p>343</p> <p>344</p> <p>345</p> <p>346</p> <p>347</p> <p>348</p> <p>349</p> <p>350</p> <p>351</p> <p>352</p> <p>353</p> <p>354</p> <p>355</p> <p>356</p> <p>357</p> <p>358</p> <p>359</p> <p>360</p> <p>361</p> <p>362</p> <p>363</p> <p>364</p> <p>365</p> <p>366</p> <p>367</p> <p>368</p> <p>369</p> <p>370</p> <p>371</p> <p>372</p> <p>373</p> <p>374</p> <p>375</p> <p>376</p> <p>377</p> <p>378</p> <p>379</p> <p>380</p> <p>381</p> <p>382</p> <p>383</p> <p>384</p> <p>385</p> <p>386</p> <p>387</p> <p>388</p> <p>389</p> <p>390</p> <p>391</p> <p>392</p> <p>393</p> <p>394</p> <p>395</p> <p>396</p> <p>397</p> <p>398</p> <p>399</p> <p>400</p> <p>401</p> |
|---|--|

ARTICLE IN PRESS

- 402 acid cycle of GABAergic neurons with 3-nitropionic acid in vivo,
403 J. Neurochem. 65 (1995) 1184–1191.
- [20] J. Kedziora, G. Bartosz, Down's syndrome: a pathology involving the
405 lack of balance of reactive oxygen species, Free Radical Biol. Med. 4
406 (1988) 317–330.
- [21] M.A. La Fontaine, J.W. Geddes, A. Banks, D.A. Butterfield, 3-
407 Nitropionic acid induced *in vivo* protein oxidation in striatal and
408 cortical synaptosomes: insights into Huntington's disease, Brain Res.
409 858 (2000) 356–362.
- [22] R.L. Levine, D. Garland, C.N. Oliver, A. Mici, I. Climent, A.G. Lenz,
410 B.W. Ahn, S. Shaltiel, E.R. Stadtman, Determination of carbonyl
411 content in oxidatively modified proteins, Method Enzymol. 186
412 (1990) 464–478.
- [23] C.R.M. Malfatti, L.F.F. Royes, L. Francescato, E.R. Sanabria, M.A.
413 Rubin, E.A. Cavalheiro, C.F. Mello, Intrastriatal methylmalonic acid
414 administration induces convulsions and TBARS production, and alters
415 Na^+ , K^+ -ATPase activity in the rat striatum and cerebral cortex,
416 Epilepsia 44 (2003) 761–767.
- [24] P.C. Marisco, M.C.P. Ribeiro, J.S. Bonini, T.T.F. Lima, K.C. Mann,
417 G.M. Brenner, C.S. Dutra-Filho, C.F. Mello, Ammonia potentiates
418 methylmalonic acid-induced convulsions and TBARS production,
419 Exp. Neurol. 182 (2003) 455–460.
- [25] R.T. Matthews, L. Yang, M.F. Beal, S-methylthiocitrulline, a neuronal
420 nitric oxide synthase inhibitor, protects against malonate and MPTP
421 neurotoxicity, Exp. Neurol. 143 (1997) 282–286.
- [26] C.A. Messan, J.G. Greene, J.T. Greenamyre, M.B. Robinson,
422 Intrastriatal injections of the succinate dehydrogenase inhibitor,
423 malonate, cause a rise in extracellular amino acids, Brain Res. 684
424 (1995) 221–224.
- [27] M.L. Monje, J. Chatten-Brown, S.E. Hye, K.M. Raley-Susman, Free
425 radicals are involved in the damage to protein synthesis after
426 anoxia/aglycemia and NMDA exposure, Brain Res. 857 (2000)
427 172–182.
- [28] H. Monyer, R. Sprang, R. Schoepfer, A. Herb, M. Ilguchi, H.
428 Lomeli, N. Burnashev, B. Sakmann, P.H. Seeburg, Heteromeric
429 NMDA receptors: molecular and functional distinction of subtypes,
430 Science 256 (1992) 1217–1221.
- [29] H. Ohkawa, N. Ohishi, K. Yagi, Assay for peroxides in animal tissues
431 by thiobarbituric acid reaction, Anal. Biochem. 95 (1979) 351–358.
- [30] J.G. Okun, F. Hörster, L.M. Farkas, P. Fehn, A. Hinz, S. Sauer, G.F.
432 Hoffmann, K. Unsicker, E. Mayatepek, S. Kölker, Neurodegeneration
433 in methylmalonic aciduria involves inhibition of complex II and the
434 tricarboxylic acid cycle, and synergistically acting excitotoxicity,
435 J. Biol. Chem. 277 (2002) 14674–14680.
- [31] P.T. Ozand, W.L. Nyhan, A. Al Aqel, J. Christodoulou, Malonic
436 aciduria, Brain Dev. 16 (1994) 7–11.
- [32] M. Riepe, N. Hor, V.C. Ludolph, D.O. Carpenter, P.S. Spencer, C.N.
437 Allen, Inhibition of energy metabolism by 3-nitropionic acid
438 activates ATP-sensitive potassium channels, Brain Res. 586 (1992)
439 61–66.
- [33] C. Rochrs, E.R. Garrido-Sanabria, A.C. Da Silva, L.C. Faria, V.D.G.
440 Sinhorin, R.H. Marques, M.R. Priel, M.A. Rubin, E.A. Cavalheiro,
441 C.F. Mello, Succinate increases neuronal post-synaptic excitatory
442 potentials *in vitro* and induces convulsive behaviour through *N*-
443 methyl-D-aspartate-mediated mechanisms, Neuroscience 125 (2004)
444 965–971.
- [34] L.F.F. Royes, M.R. Fighera, A.F. Furian, M.S. Oliveira, L.G.M. da
445 Silva, C.R.M. Malfatti, P.H. Schneider, A.L. Braga, M. Wajner, C.F.
446 Mello, Creatine protects against the convulsive behavior and lactate
447 production elicited by the intrastriatal injection of methylmalonate,
448 Neuroscience 118 (2003) 1079–1090.
- [35] A.H.V. Schapira, Mitochondria involvement in Parkinson's disease,
449 Huntington's disease, hereditary spastic paraparesis and Friedreich's
450 ataxia, Biochim. Biophys. Acta 1410 (1999) 159–170.
- [36] N.A. Simonian, J.T. Coyle, Oxidative stress in neurodegenerative
451 diseases, Annu. Rev. Pharmacol. Toxicol. 36 (1996) 83–106.
- [37] T.S. Smith, J.P. Bennett, Jr., Mitochondrial toxins in models of
452 neurodegenerative diseases, In Vivo Brain Hydroxyl Radical Production
453 During Systemic MPTP Treatment or Following Microdialysis
454 Infusion of Methylpyridinium or Azide Ions, Brain Res., vol. 76,
455 1997, pp. 183–188.
- [38] S. Toyoshima, F. Watanabe, H. Saido, K. Miyatake, Y. Nakano,
456 Methylmalonic acid inhibits respiration in rat liver mitochondria,
457 J. Nutr. 125 (1995) 2846–2850.
- [39] Y. Ueda, H. Yokoyama, R. Niwa, R. Konaka, H. Ohya-Nishiguchi, H.
458 Kamada, Generation of lipid radicals in the hippocampal extracellular
459 space during kainic acid-induced seizures in rats, Epilepsy Res. 26
460 (1997) 329–333.
- [40] E.M. Urbanska, P. Blaszcak, T. Saran, Z. Kleinrok, W.A. Turski,
461 Mitochondrial toxin 3-nitropionic acid evokes seizures in mice,
462 Eur. J. Pharmacol. 359 (1998) 55–58.
- [41] M. Wajner, J.C. Coelho, Neurological dysfunction in methylmalonic
463 aciduria is probably related to the inhibitory effect of methylmalonate
464 on brains energy production, J. Inherit. Metab. Dis. 20 (1997)
465 761–768.
- [42] M. Wajner, J.C. Dutra, S.E. Cardoso, C.M.D. Wannmacher, E.R.
466 Motta, Effect of methylmalonate on *in vitro* lactate release and carbon
467 dioxide production by brain of suckling rats, J. Inherit. Metab. Dis. 64
468 (1992) 455–458.
- [43] L.J. Willmore, M. Hiramatsu, H. Kochi, A. Mori, Formation of
469 superoxide radicals after FeCl_3 injection into rat isocortex, Brain Res.
470 277 (1983) 393–396.
- [44] L.J. Yan, M.G. Traber, L. Packer, Spectrophotometric method for
471 determination of carbonyls in oxidatively modified apolipoprotein B
472 of human low-density lipoproteins, Anal. Biochem. 228 (1995)
473 349–351.

4 - DISCUSSÃO

4. DISCUSSÃO

A ação do L-glutamato, um aminoácido de cadeia acíclica e estrutura flexível, nos vários subtipos de receptores ionotrópicos e metabotrópicos tem um importante papel tanto na comunicação neuronal como em doenças que afetam o SNC (Cotman et al., 1995). Em função disso, muitos pesquisadores tem investido na síntese de novos análogos do L-glutamato, tanto agonistas quanto antagonistas seletivos daqueles receptores. Estes novos agonistas são valiosos para caracterizar a farmacologia dos receptores (Willis et al., 1997).

No primeiro artigo, foi demonstrado por meio de experimentos com glutamato rádio-marcado, que o D,L-*cis*-2,3-PDC, um análogo do L-glutamato de cadeia cíclica e estrutura rígida, inibiu a ligação do [³H]-L-glutamato Na⁺-independente em preparações de membranas plasmáticas de córtex de ratos, sugerindo uma interação com receptores glutamatérgicos. Da mesma forma, o seu diastereoisômero, o L-*trans*-2,3-PDC também inibe a ligação do [³H]-L-glutamato, preferentemente em receptores NMDA (Willis et al., 1996). Por outro lado, o D,L-*cis*-2,3-PDC não interagiu com sítios de captação de L-glutamato, pois ele não inibiu a ligação do [³H]-L-glutamato Na⁺-dependente em preparações de membranas plasmáticas de córtex de ratos. Estes dados contrastam com a capacidade do seu diastereoisômero inibir, embora fracamente, a captação de [³H]-D-aspartato em sinaptossomas de cérebro de ratos ($K_i = 33\mu M$) (Willis et al., 1996). Já o derivado pirrolidínico L-*trans*-2,4-PDC, cujos grupos carboxilas distam um do outro três átomos carbonos, é um potente inibidor seletivo do transportador de glutamato Na⁺-dependente ($K_i = 1\mu M$) (Bridges et al., 1991). Embora, o D,L-*cis*-2,3-PDC não tenha apresentado efeito sobre a ligação do [³H]-L-glutamato Na⁺-dependente, isso não quer dizer que tal composto não altere a captação ou liberação de L-glutamato, já que os estudos de ligação não são diretamente usados para avaliar a atividade funcional e sim, identificar sítios específicos de ação. O fato de que o derivado *cis*-dicarboxilato de pirrolidina não ter alterado a ligação do [³H]-L-glutamato Na⁺-dependente enquanto o isômero *trans* altera, sugere que a configuração *cis* nos grupamentos carboxilas do anel pirrolidínico lhe atribui alguma seletividade de ligação em sítios para o L-glutamato não-transportáveis. Por isso, foi investigado se o D,L-*cis*-2,3-PDC causava alguma alteração comportamental em camundongos, e qual o possível mecanismo de ação envolvido nessas alterações.

A injeção intracerebroventricular de D,L-*cis*-2,3-PDC causou convulsões tônico-clônicas generalizadas nos camundongos de uma maneira dose-dependente. Entretanto, estas convulsões foram prevenidas pela co-administração de MK-801, um antagonista não-competitivo dos receptores NMDA. Por outro lado, a co-administração de DNQX, um antagonista dos receptores AMPA e KA, causou apenas uma ligeira proteção das convulsões induzidas pelo D,L-*cis*-2,3-PDC, sugerindo que o D,L-*cis*-2,3-PDC causa convulsões por ativar os receptores NMDA, possivelmente por se ligar no mesmo sítio que o L-glutamato, e que o envolvimento dos receptores não-NMDA é de menor relevância para o seu efeito convulsivante. Da mesma forma, o diasteroisômero L-*trans*-2,3-PDC causa neurotoxicidade quando injetado no hipocampo de ratos *in vivo* e também em culturas corticais de cérebro de ratos, os quais são revertidos pela co-administração de MK-801 (Willis et al., 1996). Um fato interessante é que o L-*trans*-2,3-PDC também apresenta alguma reatividade com os receptores AMPA e KA (Willis et al., 1996) *in vitro*. Esses resultados sugerem que embora haja uma restrição conformacional imposta pelo anel pirrolidínico este pode sofrer uma interconversão do tipo envelope-envelope o que permite a esses compostos assumir conformações nos três tipos de receptores ionotrópicos (Chamberlin & Bridges, 1993). Da mesma forma, uma relação similar entre estrutura/atividade foi observada em outros análogos do L-glutamato com conformação restrita, tal como carboxiciclopropil-glicinas (Ishida et al., 1991; Kawai et al., 1992).

O succinato é um metabólito endógeno do ciclo de Krebs e também pode ser produzido por meio de uma rota alternativa a partir do GABA (Sanders et al., 1969). Este composto está largamente distribuído no SNC, como na matéria branca e córtex cerebrais e no cerebelo (Petroff et al., 1988).

No segundo artigo, foi demonstrado que a administração ICV de succinato diminuiu a atividade exploratória, aumentou as Substâncias que reagem ao Ácido Tiobarbitúrico (TBARS) e carbonilação protéica bifasicamente. O succinato nas doses de 0,7 µmol e 1,0 µmol diminuiu a atividade locomotora e aumentou os marcadores bioquímicos do dano oxidativo. MK-801 previu o aumento da carbonilação proteica total e a diminuição da atividade locomotora induzida por 1,0 µmol de succinato.

O aumento na produção de TBARS induzido por succinato sugere que este ácido orgânico causa dano oxidativo já que a substância formada durante a

degradação de lipídios por EAO, o MDA, reage com TBA formando um complexo MDA-TBA de coloração rósea (Kohn & Liversedge, 1944; Patton & Kurtz, 1951; Dahle et al., 1962; Ohkawa et al., 1979). Da mesma forma, succinato aumentou a carbonilação protéica, um marcador específico do dano oxidativo. Estes resultados fortemente sugerem que as doses de 0,7 μ mol e 1,0 μ mol induzem dano oxidativo. Por outro lado, uma alta dose de succinato (1,7 μ mol) retornou TBARS e carbonilação protéica aos valores do controle, caracterizando um efeito bifásico do succinato sobre os marcadores do dano oxidativo estudados. O mecanismo molecular pelo qual a dose de 1,7 μ mol apresenta efeito antioxidant não está estabelecido, mas é interessante que um efeito bifásico similar de succinato sobre os potenciais pós-sinápticos excitatórios de campo (PEPSs) tem sido relatado. Enquanto baixas concentrações (0,3-1,0 mM) de succinato aumenta os PEPSs em fatias de hipocampo de ratos através da ativação dos receptores NMDA, altas concentrações de succinato (3-10 mM) diminuem os PEPSs e revertem o aumento destes causados por baixas concentrações de succinato (Roehrs et al., 2004). Naquele estudo, o efeito bifásico sobre os PEPSs foi atribuído a um efeito bifásico do succinato sobre o receptor NMDA. Dado o marcado padrão similar de variação da curva dose-efeito para o succinato o qual foi relatado por Roehrs et al. (2004), pode-se propor que eles podem ocorrer por mecanismos similares, isto é, receptores NMDA. Chen et al. (2003) demonstraram que o succinato exibe uma curva de dose-resposta na forma de U invertido, atuando como ansiolítico no teste de plus maze, corroborando com os efeitos bifásicos do succinato já descritos anteriormente.

Portanto, decidiu-se investigar se os receptores NMDA estavam envolvidos no dano oxidativo e, não no dano lipídico, induzido por succinato utilizando a técnica de carbonilação protéica. Embora a medida de malondialdeído (MDA) tenha sido usada extensivamente nas últimas quatro décadas como um marcador da LPO (Janero, 1990) este método possui algumas peculiaridades e limitações que podem levar a uma interpretação errônea dos resultados. Conseqüentemente, foi avaliado o envolvimento dos receptores NMDA somente sobre a carbonilação protéica induzida por succinato.

A co-administração de MK-801 protegeu contra o aumento da carbonilação protéica induzida por succinato, sugerindo o envolvimento deste subtipo de receptor glutamatérgico no dano oxidativo induzido por este ácido dicarboxílico.

Um importante resultado deste estudo é que o dano oxidativo foi acompanhado por significantes alterações comportamentais, cujo padrão de variação ao longo das doses de succinato coincidiu com os marcadores do dano oxidativo. Mais especificamente, succinato causou um aumento dos marcadores do dano oxidativo no cérebro, o qual coincidiu com a diminuição na atividade locomotora. Além disso, o aumento nas doses de succinato retornou os escores da atividade locomotora e marcadores bioquímicos ao nível do controle. Esses resultados revelam um paralelismo entre esses eventos, sendo tentador propor que alguma relação pode existir entre eles. Evidência adicional de que o aumento no dano oxidativo e a diminuição na atividade locomotora estão, de alguma forma, relacionados, é proveniente dos experimentos que mostraram que o MK-801 previne os efeitos do succinato sobre ambos os parâmetros bioquímicos e comportamental. Uma vez que o tratamento farmacológico afetou ambos os parâmetros (bioquímico e comportamental) na mesma direção, de forma similar, não dissociável, é possível que esses efeitos estejam relacionados. De qualquer maneira, se existe uma relação causa-efeito entre eles ainda não pode ser estabelecido e, uma análise mais detalhada da variação dos níveis de marcadores do estresse oxidativo cerebral e atividade convulsiva ao longo do tempo é necessária para determinar se existe uma relação causa-efeito entre esses efeitos. Além disso, não foi investigado se o succinato altera outros importantes marcadores do dano oxidativo, tais como o DNA, e em que extensão nossos resultados se comparam às condições humanas.

Independentemente se os efeitos neuroquímicos e neurocomportamentais do succinato aqui descritos estão casualmente relacionados, este estudo mostra que o succinato causa dano oxidativo e efeitos comportamentais em doses subconvulsivantes, por mecanismos mediados por receptores NMDA. Estes resultados são particularmente importantes para o estudo de condições nas quais acumulam succinato, tais como a exposição a toxinas exógenas inibidoras da SDH (Riepe et al., 1992; Hassel & Sonnewald, 1995; Greene & Greenamyre, 1996; Urbanska et al., 1998; Alexi et al., 1998; Calabresi et al., 2001) e endógenas (Greene et al., 1993; Davolio & Greenamyre, 1995; Toyoshima et al., 1995; De Mello et al., 1996; Fighera et al., 1999, 2003; Marisco et al., 2003; Royes et al., 2003; Fleck et al., 2004) bem como deficiências da SDH herdadas por erros inatos do metabolismo (Brismar & Ozand, 1994; Ozand et al., 1994; Brockman et al., 2002; Okun et al., 2002; Fleck et al., 2004), pois revela que pelo menos alguns dos efeitos

deletérios dessas condições podem ser decorrentes do acúmulo secundário de succinato, uma possibilidade que tem sido negligenciada.

Nesse contexto, pode-se concluir do presente estudo que o succinato, um ácido dicarboxílico de cadeia acíclica e estrutura flexível, provavelmente exerce seus efeitos neurotóxicos por ativar os receptores NMDA e que a formação das EAO podem estar envolvidas nos episódios convulsivos causados por este ácido. Além disso, não se pode descartar que o acúmulo de succinato seja de grande importância nas acidemias orgânicas nas quais acumulam malonato e metilmalonato, inibidores reversíveis da SDH. E, que o D,L-*cis*-2,3-PDC, um análogo do L-glutamato de cadeia cíclica e estrutura rígida, diasteroisômero do L-*trans*-2,3-PDC, interage com sítios de ligação para o L-glutamato não-transportáveis e causa convulsões em camundongos via ativação dos receptores NMDA. Outros estudos são ainda necessários para determinar se esta nova neurotoxina afeta outras funções do L-glutamato, bem como sua utilidade como uma ferramenta farmacológica.

Esses resultados demonstram que mesmo havendo mudança na cadeia ou estrutura química, os compostos dicarboxílicos parecem modular funcionalmente os receptores glutamatérgicos.

5 - CONCLUSÃO

- a) O D,L-*cis*-2,3-PDC inibi a ligação do [³H]-L-glutamato Na⁺-independente em receptores de membranas plasmáticas de córtex de ratos e não apresenta efeito sobre a ligação de [³H]-L-glutamato Na⁺-dependente.
- b) A administração intracerebroventricular de D,L-*cis*-2,3-PDC em camundongos induz convulsões tônico-clônicas generalizadas de uma maneira dose-dependente.
- c) A co-administração de MK-801, um bloqueador do canal NMDA, com D,L-*cis*-2,3-PDC totalmente protege os animais das convulsões induzidas por D,L-*cis*-2,3-PDC. A co-administração de DNQX, um antagonista dos receptores AMPA e KA, aumenta a latência para as convulsões induzidas por D,L-*cis*-2,3-PDC, mas não altera a percentagem de animais que apresentam convulsões. Estes resultados sugerem que o D,L-*cis*-2,3-PDC induz seus efeitos preferencialmente por meio da ativação dos receptores NMDA.
- d) A administração intracerebroventricular de succinato diminui bifasicamente a atividade locomotora dos camundongos.
- e) A administração intracerebroventricular de succinato aumenta bifasicamente a produção de Substâncias que reagem ao Ácido Tiobarbitúrico (TBARS) em cérebros de camundongos *ex vivo*.
- f) A administração intracerebroventricular de succinato aumenta bifasicamente a carbonilação de proteínas em cérebros de camundongos *ex vivo*.
- g) A co-administração de MK-801 previne a diminuição da atividade locomotora *in vivo* e o aumento da carbonilação de proteínas em cérebros de camundongos *ex vivo*. Estes resultados sugerem o envolvimento dos receptores NMDA no dano oxidativo induzido por succinato.

6 – REFERÊNCIAS BIBLIOGRÁFICAS

- ALLAN, R.D., HANRAHAN, J.R., HAMBLEY, T.W., JOHNSTON, G.A.R., MEWETT, K.N., MITROVIC, A.D. Synthesis and activity of a potent N-methyl-D-aspartic acid agonist, trans-1-aminocyclobutane-1, 3-dicarboxilic acid, and related phosphonic and carboxylic acids. **Journal of Medical Chemistry**, **33**: 2905-2915, 1990.
- ALEXI, T., HUGHES, P.E., FAULL, R.L.M., WILLIAMS, C.E. 3-nitropropionic acid's lethal triplet: cooperative pathways of neurodegeneration. **NeuroReport**, **9**: 59-64, 1998.
- ARMSTRONG, D.L., SOHAL, R.S., CUTLER, R.G. Eds. Free radicals in autoxidation and in aging. In: **Free Radicals in Molecular Biology, Aging, and Disease**. New York, Raven Press, 13-42, 1984.
- ARRIZA, J.L., FAIRMAN, W.A., WADICHE, J.I., MURDOCH, G.H., KAVANAUGH, M.P., AMARA, S.G. Functional comparisons of three glutamate transporter subtypes cloned from human motor cortex. **Journal of Neuroscience**, **14**: 5559-5569, 1994.
- BEAL, M.F. Mitochondria, free radicals, and neurodegeneration. **Current Opinion in Neurobiology**, **6(5)**: 661-666 1996.
- BEAL, M. F. Energetics in the pathogenesis of neurodegenerative diseases. **Trends in Neurosciences**, **23**: 298-304, 2000.
- BEAR, M.F., CONNORS, B.W., PARADISO, M.A. **Neurociências: desvendando o sistema nervoso**. Editora Artmed, 2^a edição, 150-153, 2002.
- BECKMAN, J.S., BECKMAN, T.W., CHEN, J. MARSHALL, P.A., FREEMAN, B.A. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. **Proceedings of the National Academy of Sciences USA**, **87**: 1620-1624, 1990.
- BEHRENS M.I., CHOI D.W., CANZIONERO L.M.T., SENSI, S.L., CSERNANSKY, C.A., CHOI, D.W. 3-Nitropropionic acid induces apoptosis in cultured striatal and cortical neurons. **NeuroReport**, **6**: 545-548, 1995.
- BERGENDI, L., BENES, L., DURACKOVA, Z., FERENCIK, M. Chemistry, physiology and pathology of free radicals. **Life Sciences**, **65**: 1865-1874, 1999.
- BETTLER, B., EGEBJERG, J., SHARMA, G., PECHT, G., HERMANS-BORGMEYER, I., MOLL, C., STEVENS, C.F., HEINEMANN, S. Cloning of a putative glutamate receptor: a low affinity kainate-binding subunit. **Neuron**, **8**: 257-265, 1992.
- BLISS, T.V.P. & COLLINGRIDGE, G.L. A synaptic model of memory: long-term potentiation in the hippocampus. **Nature**, **361**: 31-39, 1993.
- BOVERIS, A. & CHANCE, B. The mitochondrial generation of hydrogen peroxide: general properties and effect of hyperbaric oxygen. **Journal of Biochemistry**, **134**: 707-716, 1973.

- BOVERIS, A. Biochemistry of free radicals: from electrons to tissues. Buenos Aires, **Medicina**, **58**: 350-356, 1998.
- BRABET, I., PARMENTIER, M. L., DE COLLE, C., BOCKAERT, J., ACHER, F., PIN, J.P. Comparative effect of L-CCG-I, DCG-IV and gamma-carboxy-l-glutamate on all cloned metabotropic glutamate receptor subtypes. **Neuropharmacology**, **37**: 1043-1051, 1998.
- BRANN, D.W. Glutamate: A major excitatory transmitter in neuroendocrine regulation. **Neuroendocrinology**, **61**: 213-225, 1995.
- BRÄUNER-OSBORNE, H., EGEBJERG, J., NIELSEN, E.Ø., MADSEN, U., KROGSGAARD-LARSEN, P. Ligands for glutamate receptors: design and therapeutics prospects. **Journal of Medical Chemistry**, **43**(14): 2609-2645, 2000.
- BRÄUNER-OSBORNE, H., NIELSEN, E.Ø., STENSBØL, T.B., JOHANSEN, T.N., SKJÆRBÆK, N., KROGSGAARD-LARSEN, P. Molecular pharmacology of 4-substituted glutamic acid analogues at ionotropic and metabotropic excitatory amino acid receptors. **European Journal of Pharmacology**, **335**: R1-R3, 1997.
- BRIDGES, R.J., STANLEY, M.S., ANDERSON, M.W., COTMAN, C.W., CHAMBERLIN, A.R. Conformationally defined neurotransmitter analogues. Selective inhibition of glutamate uptake by one pyrrolidine-2,4-dicarboxylate diastereoisomer. **Journal of Medical Chemistry**, **34**: 717-725, 1991.
- BRISMAR, J. & OZAND, P.T. CT and MR of the brain in the diagnosis of organic acidemias: Experiences from 107 patients. **Brain & Development**, **16**: 104-124, 1994.
- BROCKMANN, K., BJORNSTAD, A., DECENT, P., KORENKE, C.G., SMEITINK, J., TRIJBELS, F., ATHANASSOPOULOS, S., VILLAGRAN, R., SKJELDAL, O.H., WILICHOWSKI, E., FRAHM, J., HANEFELD, F. Succinate in dystrophic white matter: a proton magnetic resonance spectroscopy finding characteristic for complex II deficiency. **Annals of Neurology**, **52**: 38-46, 2002.
- BRORSON, J.R., BLEAKMAN, D., CHARD, P.S., MILLER, R.J. Calcium directly permeates kainate/ α -amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid receptors in cultured cerebellar Purkinje's neurons. **Molecular Pharmacology**, **41**: 603-608, 1992.
- BURNASHEV, N., KHORDORVA, A., JONAS, P., HELM, P.J., WISDEN, W., MONYER, H., SEEBURG, P.H., SAKMANN, B. Calcium-permeable AMPA-kainate receptors in fusiform cerebellar glial cells. **Science**, **256**: 1566-1570, 1992.
- BUTTERFIELD, D.A. & KANSKI, J. Brain protein oxidation in age-related. **Mechanisms of Ageing and Development**, **122**: 945-962, 2001.

- CALABRESI, P., GUBELLINI, P., PICCONI, B., CENTONZE, D., PISANI, A., BONSI, P., GREENGARD, P., HIPSKIND, R.A., BORRELLI, E., BERNARDI, G. Inhibition of mitochondrial complex II induces a long-term potentiation of NMDA-mediated synaptic excitation in the striatum requiring endogenous dopamine. **Journal of Neuroscience**, **21**: 5110-5120, 2001.
- CARPES, M.J.S. **Reações de Heck da N-Boc-3-pirrolina com sais de diazônio. Aplicações na síntese de arilcainatos, arilaspartatos, lactamas ariladas e do baclofeno e análogos.** 2001. Tese (Doutorado em Química - Pós-Graduação de Química, Unicamp, Campinas, SP).
- CHAMBERLIN, A.R. & BRIDGES, R.J. Conformationally constrained amino acids as probes of glutamate receptors and transporters. In: **Drug Design for Neuroscience** (A.P. Kozikowski, Ed.), Raven Press, New York, 231-259, 1993.
- CHANCE, B., SIES, H., BOVERIS, A. Hydroperoxide metabolism in mammalian organs. **Physiological Reviews**, **59**: 527-605, 1979.
- CHEN, S.W., XIN, Q., KONG, W.X., MIN, L., LI, J.F. Anxiolytic-like effect of succinic acid in mice. **Life Sciences**, **73**: 3257-3264, 2003.
- CHITTAJALLU, R., VIGNES, M., DEV, K.K., BARNES, J.M., COLLINGRIDGE, G.L., HENLEY, J.M. Regulation of glutamate release by presynaptic kainate receptors in the hippocampus. **Nature**, **379**: 78-81, 1996.
- CHITTAJALLU, R., BRAITHWAITE, S.P., CLARKE, V.R.J., HENLEY, J.M. Kainate receptors: subunits, synaptic localization and function. **Trends in Pharmacological Sciences**, **20**: 26-35, 1999.
- CHOI, D. W. Excitotoxic cell death. **Journal of Neurobiology**, **23**: 1261-1276, 1992.
- CONN, P.J. & PATEL, J. Eds. **The metabotropic glutamate receptors**; Humana Press: Totowa, New Jersey, 1994.
- COTMAN, C.W., KAHLE, J.S., MILLER, S.E., ULAS, J. BRIDGES, R.J. Excitatory amino acid neurotransmission. In **Psychopharmacology: The Forth Generation of Progress**. Floyd, E., Bloom and Kupfer, D.J. eds. Raven Press, Ltd., New York, 75-85, 1995.
- COYLE, J.T. & SCHWARCZ, R. Lesion of estriatal neurones with kainic acid provides a model for Huntington's chorea. **Nature**, **263**: 244-246, 1976.
- COYLE, J. T. & PUTTFARCKEN, P. Oxidative stress, glutamate, and neurodegenerative disorders. **Science**, **262**: 689-695, 1993.
- CRAWFORD, N., LANG, T.K., KERR, D.S., de VRIES, D. High affinity [³H]-kainic acid binding to brain membranes: a re-evaluation of ligand potency and selectivity. **Journal of Pharmacological and Toxicological Methods**, **42**: 121-125, 2000.

- CROUCHER, M.J., COTTERELL, K.L., BRADFORD, H.F. Amygdaloid kindling by repeated focal N-methyl-D-aspartate administration: comparison with electrical kindling. **European Journal of Pharmacology**, 265-271, 1995.
- CURTIS, D.R., PHILLIS, J.W., WATKINS, J.C. Chemical excitation of spinal neurons. **Nature**, 183: 611, 1959.
- CURTIS, D.R., PHILLIS, J.W., WATKINS, J.C. The chemical excitation of spinal neurons by certain acidic amino acids. **Journal of Physiology**, 150: 656-682, 1960.
- DAHLE, L.K., HILL, E.G., HOLMN, R.T. Thiobarbituric acid reaction and autoxidations of polyunsaturated fatty acid methyl esters. **Archives of Biochemistry and Biophysics**, 98: 252-260, 1962.
- DAVOLIO, C. & GREENAMYRE, J.T. Selective vulnerability of the CA1 region of hippocampus to the indirect excitotoxic effects of malonic acid. **Neuroscience Letters**, 192: 29-32, 1995.
- DEBERNARDI, R., MAGISTRETTI, P.J., PELLERIN, L. Trans-inhibition of glutamate transport prevents excitatory amino acid-induced glycolysis in astrocytes. **Brain Research**, 850: 39-46, 1999.
- DINGLEDINE, R. & MCBAIN, C.J. Excitatory amino acid transmitters. In: **Basic Neurochemistry: molecular, cellular and medical aspects**, eds: Siegel, G.J., Agranoff, B.W., Albers, R.W. & Molinoff, P.B. Raven Press, New York, 367-387, 1994.
- DANBOLT, N.C. Glutamate uptake. **Progress in Neurobiology**, 65: 1-105, 2001.
- DAWSON, V.L. & DAWSON, T.M. Free radicals and neuronal cell death. **Cell Death and Differentiation**, 3: 71-78, 1996.
- DEL MAESTRO, R. F. An approach and to free radicals in medicine and biology. **Acta Physiologica Scandinavica Supplementum**, 492: 153-167, 1980.
- DE MELLO, C.F., BEGNINI, J., JIMÉNEZ-BERNAL, R.E., RUBIN, M.A., DE BASTIANI, J., DA COSTA, E.J., WAJNER, M. Intrastriatal methylmalonic administration induces rotational behavior and convulsions through glutamatergic mechanisms. **Brain Research**, 721: 120-125, 1996.
- DESAI, M.A., & CONN, P.J. Selective activation of phosphoinositide hydrolysis by a rigid analogue of glutamate. **Neuroscience Letters**, 109: 157-162, 1990.
- DINGLEDINE, R., BOLAND, R.M., CHAMBERLIN, N.L., KAWASAKI, K., KLECKENER, N.W., TRAYNELIS, S.F., VERDOON, T.A. Amino acid receptor and uptake systems in the mammalian central nervous system. **Critical Reviews in Neurobiology**, 4: 1-96, 1988.

- DONEVAN, S.D., BEG, A., GUNTHER, J.M., TWYNAN, R.E. The methylglutamate, SYM 2081, is a potent and highly selective agonist at kainate receptors. **Journal of Pharmacology and Experimental Therapeutics**, **285**: 539-545, 1998.
- DUGAN, L.L., SENSI, S.L., CANZONIERO, L.M.T., HANDRAN, S.D., ROTHMAN, S.M., LIN, T.S., GOLDBERG, M.P., CHOI, D.W. Mitochondrial production of reactive oxygen species in cortical neurons following exposure to N-methyl-D-aspartate. **Journal of Neuroscience**, **15**: 6377-6388, 1995.
- EREKINSKA, M. & NELSON, D. Effects of 3-nitropropionic acid on synaptosomal energy and transmitter metabolism: relevance to neurodegenerative brain diseases. **Journal of Neurochemistry**, **63**: 1033-1041, 1994.
- ESCRIBANO, A., EZQUERRA, J., PEDREGAL, C., RUBIO, A., YRURETAGOYENA, B., BAKER, S.R., WRIGHT, R.A., JOHNSON, B.G., SCHOEPP, D.D. (2S,4S)-Amino-4-(2,2-diphenylethyl)pentanedioic acid selective group 2 metabotropic glutamate receptor antagonist. **Bioorganic and Medicinal Chemistry Letters**, **8**: 765-770, 1998.
- ESTERBAUER, H., KOLLER, E., SLEE, R.G., KOSTER, J.F. Possible involvement of the lipid-peroxidation product 4-hydroxynonenal in the formation of fluorescent chromolipids. **Biochemical Journal**, **239**: 405-409, 1986.
- EULER, G.V. & LIU, Y. Glutamate and glycine decrease the affinity of [³H]MK-801 binding in the presence of Mg²⁺. **European Journal of Pharmacology**, **24**: 233-239, 1993.
- FENG, B., TSE, H.W., SKIFTER, D.A., MORLEY, R., JANE, D.E., MONAGHAN, D.T. Structure-activity analysis of a novel NR2C/NR2D-preferring NMDA receptor antagonist: 1-(phenanthrene-2-carbonyl) piperazine-2,3-dicarboxylic acid. **British Journal of Pharmacology**, **141**: 508-516, 2004.
- FIGHERA, M.R., QUEIROZ, C.M., STRACKE, M.P., NIN BAUER, M.C., GONZÁLEZ-RODRIGUEZ, L.L., FRUSSA-FILHO, R., WAJNER, M., DE MELLO, C.F. Ascorbic acid and α-tocopherol attenuate methylmalonic acid-induced convulsions. **NeuroReport**, **10**: 2039-2043, 1999.
- FIGHERA, M.R., BONINI, J.S., DE OLIVEIRA, T.G., FRUSSA-FILHO, R., DUTRA-FILHO, C.S., HAGEN, M.E., RUBIN, M.A., MELLO, C.F. GM1 ganglioside attenuates convulsions and thiobarbituric acid reactive substances production induced by the intrastriatal injection of methylmalonic acid. **International Journal of Biochemistry & Cell Biology**, **35**: 465-473, 2003.
- FISCHER, A.B. Intracellular production of oxygen-derived free radicals. **Proceedings of a Brook Lodge Symposium**, Augusta, Apr. 27-29, 99-104, 1987.
- FLECK, J., RIBEIRO, M.C.P., SCHNEIDER, C.M., SINHORIN, V.D.G., RUBIN, M.A., MELLO, C.F. Intrastriatal malonate administration induces convulsive behavior in rats. **Journal of Inherited Metabolic Disease**, **24**: 211-219, 2004.

- FLETCHER, E.J., MEWETT, K.N., DREW, C.A., ALLAN, R.D., JOHNSTON, G.A.R. Inhibition of high affinity L-glutamic acid uptake into rat cortical synaptosomes by the conformationally restricted analogue of glutamic acid, cis-1-aminocyclobutane-1, 3-dicarboxylic acid. **Neuroscience Letters**, **121**(1-2): 133-135, 1991.
- FLOYD, R.A. Neuroinflammatory processes are important in neurodegenerative diseases: an hypothesis to explain the increase formation of reactive oxygen and nitrogen species as major factors involved in neurodegenerative disease development. **Free Radical Biology and Medicine**, **26**: 1346-1355, 1999.
- FONNUN, F. Glutamate: a neurotransmitter in mammalian brain. **Journal of Neurochemistry**, **42**: 1-11, 1984.
- FOSTER, A.C. & WONG, E.H.F. The novel anticonvulsant MK-801 binds to the activated state on the N-methyl-D-aspartate receptor in the rat brain. **British Journal of Pharmacology**, **91**: 403-409, 1987.
- FRIDOVICH, I. The biology of oxygen radicals. **Science**, **201**: 875-880, 1978.
- FYKSE, E.M., IVERSEN, E.G., FONNUM. Inhibition of L-glutamate uptake into synaptic vesicles. **Neuroscience Letters**, **135**: 125-128, 1992.
- FYKSE, E.M. & FONNUN, F. Amino acid neurotransmission: dynamics of vesicular uptake. **Neurochemical Research**, **21**: 1053-1060, 1996.
- GERSCHEMAN, R., GILBERT, D.L., NYE, S.W., DWYER, P., FENN, W.O. Oxygen poisoning in common. **Science**, **119**: 623-626, 1954.
- GREENAMYRE, J.T. & PORTER, R.H.P. Anatomy and physiology of glutamate in the CNS. **Neurology**, **44**(8): S7-S13, 1994.
- GREENE, J.G., PORTER, R.H.P., ELLER, R.V., GREENAMYRE, J.T. Inhibition of succinate dehydrogenase by malonic acid produces an excitotoxic lesion in rat striatum. **Journal of Neurochemistry**, **61**: 1151-1154, 1993.
- GREENE, J. G. & GREENAMYRE, J. T. Characterization of the excitotoxic potential if the reversible succinate dehydrogenase inhibitor malonate. **Journal of Neurochemistry**, **64**: 430-436, 1995.
- GREENE, G.G. & GREENAMYRE, J.T. Manipulation of membrane potential modulates malonate-induced striatal excitotoxicity *in vivo*. **Journal of Neurochemistry**, **66**: 637-643, 1996.
- GU, Z.Q., HESSON, D.P., PELLETIER, J.C., MACCECCHINI, M.L., ZHOU, L.M., SKOLNICK, P. Synthesis, resolution, and biological evaluation of the four stereoisomers of 4-methylglutamic acid: selective probes of kainate receptor. **Journal of Medical Chemistry**, **38**: 2518-2520, 1995.

- GUIDETTI, P. & SCHWARCZ, R. Determination of α -amino adipic acid in brain, peripheral tissues, and body fluids using GC/MS with negative chemical ionization. **Molecular Brain Research**, **118**: 132-139, 2003.
- GULDBRANDT, M., JOHANSEN, T.N STENSBØL,T.B., NIELSEN, B., KARLA, R., SANTI, F., KROGSGAARD-LARSEN, P., MADSEN, U. Glutamate receptor ligands: Synthesis stereochemistry, and enantioselectivity of methylated 2-amino adipic acid analogs. **Chirality**, **14(4)**: 351-363, 2002.
- GUNTER, T.E., GUNTER, K.K., SHEU, S.S., GAVIN, C.F. Mitochondrial calcium transport. **American Journal of Physiology**, **267**: 313-339, 1994.
- HALL, J.G., McLENNAN, H., WHEAL, H.V. The actions of certain amino acids as neuronal excitants. **Journal of Physiology**, **272**: 52, 1977.
- HALLIWELL, B. Free radicals and antioxidants: a personal view. **Nutrition Reviews**, **52**: 253-280, 1994.
- HALLIWELL, B. & GUTTERIDGE, J.M.C. In: **Free Radical Biology and Medicine**, 3^a ed., New York, Oxford University Press, 1999.
- HAMBERGER, A. & NYSTRÖM, B. Extra- and intracellular amino acids in the hippocampus during development of hepatic encephalopathy. **Neurochemical Research**, **9**: 1181-1192, 1984.
- HAMBERGER, A., BERTHOLD, C.H., KARLSSON, B., LEHMANN, A., NYSTRÖM, B. Extracellular GABA, glutamate and glutamine in vivo- perfusion-dialysis of rabbit hippocampus. **Neurological Neurobiology**, **7**: 473-492, 1983.
- HANSEN, J.J. & KROGSGAARD-LARSEN, P. Structural, conformational, stereochemical requirements of central excitatory amino acid receptors. **Medicinal Research Reviews**, **10**: 55-94, 1990.
- HARRIS, M.E. Regulation of antioxidant enzymes. **FASEB Journal**, **6**: 2675-2683, 1992.
- HASSEL, B. & SONNEWALD, U. Selective inhibition of the tricarboxylic acid cycle of GABAergic neurons with 3-nitropropionic acid *in vivo*. **Journal of Neurochemistry**, **65**: 1184-1191, 1995.
- HAYASHI, Y., TANABE, Y., ARAMORI, I., MASU, M., SHIMAMOTO, K., OHFUNE, Y., NAKANISHI, S. Agonist analysis of 2-(carboxycyclopropyl) glycine isomers for cloned metabotropic glutamate receptor subtypes expressed in Chinese hamster ovary cells. **Brazilian Journal of Pharmacology**, **107**: 539-543, 1992.
- HAYASHI, T. Effects of sodium glutamate on the nervous system. Keio. **Journal of Medicine**, **3**: 183-192, 1954.
- HAWKINS, L.M., BEAVER, K.M., JANE, D.E., TAYLOR, P.M., SUNTER, D.C., ROBERTS, P.J. Characterization of the pharmacology and regional distribution

- of (S)-[3H]-5-fluorowillardiine binding in rat brain. **Brazilian Journal of Pharmacology**, **116**: 2033-2039, 1995.
- HEFFNER, J. E. & REPINE, J. E. Pulmonary strategies of antioxidant defense. **American Reviews Respiratory Disease**, **140**: 531-554, 1989.
- HIMI, T., IKEDA, M., YASUHARA, T., MUROTA, S. Oxidative neuronal death caused by glutamate uptake inhibition in cultured hippocampal neurons. **Journal of Neuroscience Research**, **71(5)**: 679-688, 2002.
- HOLLMANN, M. & HEINEMANN, S. Cloned glutamate receptors. **Annual Review of Neuroscience**, **17**: 31-108, 1994.
- HASHIMOTO, A. & OKA, T. Free D-aspartate and D-serine in the mammalian brain and periphery. **Progress in Neurobiology**, **52**: 325-353, 1997.
- HUMPHREY, J.M., BRIDGES, R.J., CHAMBERLIN, A.R. 2,3-pyrrolidine dicarboxylates as neurotransmitter conformer mimics: enantioselective synthesis via chelation-controlled enolate alkylation. **Journal of Organic Chemistry**, **59**: 2467-2472, 1994.
- ISHIDA, M., OHFUNE, Y., SHIMADA, Y., SHIMAMOTO, K., SHINOZAKI, H. Changes in a preference for receptor subtypes of configurational variants of a glutamate analog: conversion from the NMDA-type to the non-NMDA type. **Brain Research**, **550**: 152-156, 1991.
- ISHIDA, M., & SHINOZAKI, H. Acromelic acid is a much more potent excitant than kainic acid or domoic acid in the rat spinal cord. **Brain Research**, **474**: 386-389, 1988.
- IZQUIERDO, I. & MEDINA, J.H. Correlation between the pharmacology of Long-term potentiation and the pharmacology of memory. **Neurobiology of Learning and Memory**, **63**: 19-32, 1995.
- JANERO, D.R. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. **Free Radical Biology and Medicine**, **9**: 515-540, 1990.
- JOHNSTON, G.A.R. Glutamate uptake and its possible role in neurotransmitter inactivation. In: Roberts, P.J., Storm-Mathisen, J., Johnston, G.A.R. (Eds.), **Glutamate: Transmitter in the Central Nervous System**. Wiley, Chichester, UK, 77-87, 1981.
- JONES, K.A., WILDING, T.J., HUETTNER, J.E., COSTA, A.M. Desensitization of kainate receptor by kainate, glutamate and diastereomers of 4-methylglutamate. **Neuropharmacology**, **36**: 853-863, 1997.
- JUNQUEIRA, D., BRUSQUE, A.M., PORCIÚNCULA, L.O., ROTTA, L.N., FRIZZO, M.E.S., WYSE, A.T.S., WANNMACHER, C.M.D., SOUZA, D.O., WAJNER, M. In vitro effects of D-2-hydroxyglutaric acid on glutamate binding, uptake and

- release in cerebral cortex of rats. **Journal of the Neurological Sciences**, **217**: 189-194, 2004.
- KAWAI, M., HORIKAWA, Y., ISHIHARA, T., SHIMAMOTO, K., OHFUNE, Y. 2-(Carboxycyclopropyl)glycines: binding, neurotoxicity and induction of intracellular free Ca^{2+} increase. **European Journal of Pharmacology**, **211**: 195-202, 1992.
- KIM, H., JHOO, W., BING, G. Phenidone prevents kainate-induced neurotoxicity via antioxidant mechanisms. **Brain Research**, **874**: 15-23, 2000.
- KOCH, H.P., CHAMBERLIN, A.R., BRIDGES, R.J. Nontransportable inhibitors attenuate reversal of glutamate uptake in synaptosomes following a metabolic insult. **Molecular Pharmacology**, **55**: 1044-1048, 1999.
- KOHN, H.I. & LIVERSEDGE, M. On a new aerobic metabolite whose production by brain is inhibited by apomorphine, emetine, ergotamine, epinephrine, and menadione. **Journal of Pharmacology and Experimental Therapeutics**, **82**: 292-300, 1944.
- KÖLKER, S., OKUN, J.G., AHLEMEYER, B., WYSE, A.T.S., HÖRSTER, F., WAJNER, M., KOHLMÜLLER, D., MAYATEPEK, E., KRIEGLSTEIN, J., HOFFMANN, G.F. Chronic treatment with glutaric acid induces partial tolerance to excitotoxicity in neuronal cultures from chick embryo telencephalons. **Journal of Neuroscience Research**, **68**(4): 424-431, 2002.
- KOZIKOWSKI, A.P., ARALDI, G.L., TÜCKMANTEL, W., PSHENICHKIN, S., SURINA, E., WROBLEWSKI, J.T. 1-Amino-APDC, a partial agonist of group II metabotropic glutamate receptors with neuroprotective properties. **Bioorganic and Medicinal Chemistry Letters**, **9**: 1721-1726, 1999.
- KOZIKOWSKI, A.P., STEENSMA, D., ARALDI, G.L., TÜCKMANTEL, W., WANG, S., PSHENICHKIN, S., SURINA, E., WROBLEWSKI, J.T. Synthesis and biology of the conformationally restricted ACPD analogue, 2-aminobicyclo[2.1.1]hexane-2, 5-dicarboxylic acid-I, a potent mGluR agonist. **Journal of Medical Chemistry**, **41**: 1641-1650, 1998.
- KREBS, H.A. Metabolism of amino acids. IV. Synthesis of glutamine from glutamic acid and ammonia, and the enzymatic hydrolysis of glutamine in animal tissue. **Biochemical Journal**, **29**: 1951-1969, 1935.
- KRIEGLSTEIN, J. Excitotoxicity and neuroprotection. **European Journal of Pharmaceutical Sciences**, **5**: 181-187, 1997.
- KUNZ, W., GOUSSAKOV, I.V., BECK, H., ELGER, C.E. Altered mitochondrial oxidative phosphorylation in hippocampal slices of kainate-treated rats. **Brain Research**, **826**: 236-242, 1999.
- LARM, J.A., BEART, P.M., CHEUNG, N.S. Neurotoxin domoic acid produces cytotoxicity via kainate and AMPA-sensitive receptors in cultured cortical neurones. **Neurochemistry International**, **31**: 677-682, 1997.

- LEE, S.H., HAN, S.H., LEE, K.W. Kainic acid-induced seizures cause neuronal death in infant rats pretreated with lipopolysaccharide. **NeuroReport**, **11**(3): 507-510, 2000.
- LEHMANN, A., ISACSSON, H., HAMBERGER, A. Effects of in vivo administration of kainic acid on the extracellular amino acid pool in the rabbit hippocampus. **Journal of Neurochemistry**, **40**: 1314-1320, 1983.
- LEVINE, R.L. Carbonyl modified proteins in cellular regulation, aging, and disease. **Free Radical Biology and Medicine**, **32**: 790-796, 2002.
- LIMA, T.T.F., BEGNINI, J., DE BASTIANI, J., JURACH, A., RIBEIRO, M.C.P., WAJNER, M., MELLO, C.F. Pharmacological evidence for GABAergic and glutamatergic involvement in the convulsant and behavioral effects of glutaric acid. **Brain Research**, **802**: 55-60, 1998.
- LIPTON, S.A. & ROSENBERG, P.A. Mechanisms of disease. Excitatory amino acids as a final common pathway for neurological disorders. **New England Journal of Medicine**, **330**: 613-622, 1994.
- LIU, Q., RAINA, A.K., SMITH, M.A., SAYRE, L.M., PERRY, G. Hydroxynonenal, toxic carbonyls, and Alzheimer disease. **Molecular Aspects of Medicine**, **24**: 305-313, 2003.
- LOMELLI, H., WISDEN, W., KÖHLER, M., KEINÄNEN, K., SOMMER, B., SEEURG, P.H. High-affinity Kainate and domoate receptors in rat brain. **Federation of European Biochemical Societies**, **307**: 139-143, 1992.
- MACHLIN, L.J. & BENDICH, A. Free radical tissue damage: protective role of antioxidant nutrients. **FASEB Journal**, **1**: 441-445, 1987.
- MARISCO, P.C., RIBEIRO, M.C.P., BONINI, J.S., LIMA, T.T.F., MANN, K.C., BRENNER, G.M., DUTRA-FILHO, C.S., MELLO, C.F. Ammonia potentiates methylmalonic acid-induced convulsions and TBARS production. **Experimental Neurology**, **182**: 455-460, 2003.
- MARTIN, D.C., PLAGENHOEF, M., ABRAHAM, J., DENNISON, R.L., ARONSTAM, R.S. Volatile anesthetics and glutamate activation of N-methyl-D-aspartate receptors. **Biochemical Pharmacology**, **49**: 809-817, 1995.
- MARKLUND, S.L. Role of toxic effects of oxygen in reperfusion damage. **Journal Molecular Cellular Cardiology**, **20**(2): 23-30, 1988.
- MAYER, M.L., BENEVISTE, J.M., PATNEAU, D.K., VYKLICKY Jr., L. Pharmacology properties of NMDA receptors. **Annals of the New York Academy of Sciences**, **648**: 194-204, 1992.
- MCBEAN, G.J. Inhibition of the glutamate transporter and glial enzymes in rat striatum by the gliotoxin, alpha amino adipate. **Brazilian Journal of Pharmacology**, **113**: 536-540, 1994.

- McDONALD, J.W. & SCHOEPP, D.D. Aminooxyacetic acid produces excitotoxic brain injury in neonatal rats. **Brain Research**, **624**: 239-244, 1993.
- MCENTEE, W.J. & CROOK, T.H. Glutamate: its role in learning, memory, and the aging brain. **Psychopharmacology**, **111**: 391-401, 1993.
- MCGREGOR, D.G., HIGGINS, M.J., JONES, P.A., MAXWELL, W.L., WATSON, M.W., GRAHAM, D.I., STONE, T.W. Ascorbate attenuates the systemic kainate-induced neurotoxicity in the rat hippocampus. **Brain Research**, **727**: 133-144, 1996.
- MCINTOSH, L.J., TRUSH, M.A., TRONCOSO, J.C. Increased susceptibility of Alzheimer's disease temporal cortex to oxygen free radical-mediated processes. **Free Radical Biology and Medicine**, **23**: 183-190, 1997.
- MELLO, C.F., DE BEGNINI, J., JIMÉNEZ-BERNAL, R. E., RUBIN, M.A., BASTIANI, J., COSTA, E.J.M., WAJNER, M. Unstriatal methylmalonic acid administration induces rotational behavior and convulsions through glutamatergic mechanisms. **Brain Research**, **721**: 120-125, 1996.
- MELLO, C.F., DE KÖLKER, S., AHLEMAYER, B., DE SOUZA, F.R., FIGHERA, M.R., MAYATEPEK, E., KRIEGLSTEIN, J., HOFFMANN, G.F., WAJNER, M. Intrastratal administration of 3-hydroxyglutaric acid induces convulsions and striatal lesions in rats. **Brain Research**, **916**: 70-75, 2001.
- MENEGHINI, R. A toxicidade do oxigênio. **Ciência Hoje**, **5**: 57-62, 1987.
- MONN, J.A., VALLI, M.J., JOHNSON, B.G., SALHOFF, C.R., WRIGHT, R.A., HOWE, T., BOND, A., LODGE, D., SPANGLE, L.A., PASCHAL, J.W., CAMPBELL, J.B., GRIFFEY, K., TIZZANO, J.P., SCHOEPP, D.D. Synthesis of the four isomers of 4-aminopyrrolidine-2,4-dicarboxylate: Identification of a potent, highly selective, and systemically active agonist for metabotropic glutamate receptors negatively coupled to adenylate cyclase. **Journal of Medical Chemistry**, **39**: 2990-3000, 1996.
- MONN, J.A., VALLI, M.J., MASSEY, S.M., WRIGHT, R.A., SALHOFF, C.R., JOHNSON, B.G., HOWE, T., ALT, C.A., RHODES, G.A., ROBEY, R.L., GRIFFEY, K.R., TIZZANO, J.P., KALLMAN, M.J., HELTON, D.R., SCHOEPP, D.D. Design, synthesis and pharmacological characterization of (+)-2-aminobicyclo[3.1.0] hexane-2, 6-dicarboxylic acid (LY354740): a potent, selective and orally active group 2 metabotropic glutamate receptor agonist possessing anticonvulsant and anxiolytic properties. **Journal of Medical Chemistry**, **40**: 528-537, 1997.
- MONN, J.A., VALLI, M.J., MASSEY, S.M., HANSEN, M.M., KRESS, T.J., WEPSIEC, J.P., HARKNESS, A.R., GRUTSCH, J.L.JR., WRIGHT, R.A., JOHNSON, B.G., ANDIS, S.L., KINGSTON, A., TOMLINSON, R., LEWIS, R., GRIFFEY, K.R., TIZZANO, J.P., SCHOEPP, D.D. Synthesis, pharmacological characterization, and molecular modeling of heterobicyclic amino acids related to (+)-2-

- aminobicyclo[3.1.0]-hexane-2, 6-dicarboxylic acid (LY354740): identification of two new potent, selective, and systemically active agonist for group II metabotropic glutamate receptors. **Journal of Medical Chemistry**, **42**: 1027-1040, 1999.
- MORIYAMA, Y., HAYASHI, M., YAMADA, H., YATSUSHIRO, S., ISHIO, S., YAMAMOTO, A. Synaptic-like microvesicles, synaptic vesicle counterparts in endocrine cells, are involved in a novel regulatory mechanism for the synthesis and secretion of hormones. **Journal of Experimental Biology**, **203**: 117-125, 2000.
- MORONI, F., NICOLETTI, F., PELLEGRINI-GIAMPIETRO, D.E. Eds. **The metabotropic glutamate receptors and brain function**; Portland Press: London, 1998.
- MURPHY, M.E. & SIES, H. Reversible conversion of nitroxyl anion to nitric oxide by superoxide dismutase. **Proceedings of the National Academy of Sciences USA**, **88**: 10860-10864, 1991.
- NAKANISHI, S. & MASU, M. Molecular diversity of glutamate receptors and functions of glutamate receptors. **Annual Review of Biophysics and Biomolecular Structure**, **23**: 319-348, 1994.
- NAKANISHI, S. Molecular diversity of glutamate receptors and implications for brain function. **Science**, **258**: 597-603, 1992.
- NICOLETTI, F., BRUNO, V., COPANI, A., CASABONA, G., KNOPFEL, F. Metabotropic glutamate receptors: a new target for the therapy of neurodegenerative disorders? **Trends in Neurosciences**, **19**: 267-272, 1996.
- NICHOLLS, D.G. Ion channels and the regulation of neurotransmitter glutamate release. **Biochemical Society Transactions**, **21**: 53-58, 1993.
- NORDBERG, J. & ARNÉR, S.J. Reactive oxygen species, antioxidants, and the mammalian thioredoxin system. **Free Radical Biology and Medicine**, **31**: 1287-1312, 2001.
- OHKAWA, H., OHISHI, N., YAGI, K. Assay for peroxides in animal tissues by thiobarbituric acid reaction. **Analytical Biochemistry**, **95**: 351-358, 1979.
- OKUN, J.G., HÖRSTER, F., FARKAS, L.M., FEYH, P., HINZ, A., SAUER, S., HOFFMANN, G.F., UNSICKER, K., MAYATEPEK, E., KÖLKER, S. Neurodegeneration in methylmalonic aciduria involves inhibition of complex II and the tricarboxylic acid cycle, and synergistically acting excitotoxicity. **Journal of Biological Chemistry**, **277**: 14674-14680, 2002.
- OLNEY, J.W. Neurotoxicity of amino acids. In: **Kainic acid as a tool in neurobiology**, eds: McGeer, E.G., Olney, J.W. & McGeer, P.L. Raven Press, New York, 95-121, 1978.

- OTTERSEN, O.P., LAAKE, J.H., REICHELT, W., HAUG, F.M., TORP, R. Ischemic disruption of glutamate homeostasis in brain: quantitative immunocytochemical analysis. **Journal of Chemical Neuroanatomy**, **12**: 1-14, 1996.
- OTTERSEN, O.P., ZHANG, N., WALBERG, F. Metabolic compartmentation of glutamate and glutamine: morphological evidence obtained by quantitative immunocytochemistry in rat cerebellum. **Neuroscience**, **46**: 519-534, 1992.
- ORTWINE, D.F., MALONE, T.C., BIGGE, C.F., DRUMMOND, J.T., HUMBLET, C., JOHNSON, G., PINTER, G.W. Generation of N-methyl-D-aspartate agonist and competitive antagonist pharmacophore models. Design and synthesis of phosphonoalkyl-substituted tetrahydroisoquinolines as novel antagonists. **Journal of Medical Chemistry**, **35**: 1345-1370, 1992.
- OZAND, P.T., NYHAN, W.L., AQEEL, A.A., CHRISTONOMIDOULOU, J. Malonic aciduria. **Brain & Development**, **16**: 7-11, 1994.
- OZAWA, S., KAMIYA, H., TZUSUKI, K. Glutamate receptors in the mammalian central nervous system. **Progress in Neurobiology**, **54**: 581-618, 1998.
- PACKER, L.; MIDORI, H.; TOSHIKAZU, Y. In: **Free radicals in brain physiology and disorders**. Academic Press, New York, 1996.
- PALMER, E., MONAGHAN, D.T., COTMAN, C.W., Trans-ACPD, a selective agonist of the phosphoinositide-coupled excitatory amino acid receptor. **European Journal of Pharmacology**, **166**: 585-587, 1989.
- PAN, J.C., PEI, Y.Q., AN, L., LAI, L., D'HOOGE, R.D., DEYN, P.P. Epileptiform activity and hippocampal damage produced by intrahippocampal injection of guanidinosuccinic acid in rat. **Neuroscience Letters**, **209**: 121-124, 1996.
- PANNICKE, T., STABEL, J., HEINEMANN, U., REICHELT, W. Alpha-amino-adipic acid blocks the Na^+ -dependent glutamate transport into acutely isolated Muller glial cells from guinea pig retina. **Pflüger's Archives**, **429**: 140-142, 1994.
- PATTON, S. & KURTZ, G.W. **Journal of Dairy Science**, **34**: 669-674, 1951.
- PAVLAKOVIC, G., EYER C.L., ISOM G.E. Neuroprotective effects of PKC inhibition against chemical hypoxia. **Brain Research**, **676**: 205-211, 1995.
- PENNYPACKER, K.R., HONG, J.S., MCMILLIAN, M. Implication of prolonged expression of Fos-related antigens. **Trends in Pharmacological Sciences**, **16**: 317-321, 1995.
- PERRY, G., NUNOMURA, A., HIRAI, K., ZHU, X., PEREZ, M., AVILA, J., CASTELLANI, R., ATWOOD, C.S., ALIEV, G., SAYRE, L.M., TAKEDA, A., SMITH, M.A. Is oxidative damage the fundamental pathogenic mechanism of Alzheimer's and other neurodegenerative diseases? **Free Radical Biology and Medicine**, **33**: 1475-1479, 2002.

- PETROFF, O.A.C., OGINO, T., ALGER, J.R. High-resolution proton magnetic resonance spectroscopy of rabbit brain: regional metabolite levels and postmortem changes. **Journal of Neurochemistry**, **51**: 163-171, 1988.
- PIGGOTT, M.A., PERRY, E.K., SAGHAL, A., PERRY, R.H. Examination of parameters influencing [³H]MK-801 binding in postmortem human cortex. **Journal of Neurochemistry**, **58**: 1001-1008, 1992.
- PIN, J.P. & DUVOISIN, R. The metabotropic glutamate receptors structure and functions. **Neuropharmacology**, **34**: 1-26, 1995.
- POLI, G., ALBANO, E., DIANZANI, M.U. eds. Free radicals and antioxidants in muscular neurological diseases ans disorders. In: **Free Radicals: from basic Science to Medicine**. Basel Birkhäuser Verlag, 425-437, 1993.
- PORCIÚNCULA, L.O., EMANUELLI, T., TAVARES, R.G., SCHWARZBOLD, C., FRIZZO, M.E.S., SOUZA, D.O., WAJNER, M. Glutaric acid stimulates glutamate binding and astrocytic uptake and inhibits vesicular glutamate uptake in forebrain from young rats. **Neurochemistry International**, **45**: 1075-1086, 2004.
- POW, D.V. Visualising the activity of the cystine-glutamate antiporter in glial cells using antibodies to aminoacidic acid, a selectively transported substrate. **Glia**, **34**: 27-38, 2001.
- PRATICÒ, D. & DELANTY, N. Oxidative injury in diseases of the central nervous system: focus on Alzheimer's disease. **American Journal of Medicine**, **109**: 577-585, 2000.
- PRICE, D.L. New order from neurological disorders. **Nature**, **399**: A3-A5, 1999.
- PRYBYLOWSKI, K. & WENTHOLD, R.J. N-methyl-D-aspartate receptors: subunit assembly and trafficking to the synapse. **Journal of Biological Chemistry**, **279**: 9673-9676, 2004.
- PUNCHARD, N.A. & KELLY, F.J. **Free Radicals: a practical approach**. Oxford University Press Inc., New York, 1996.
- RANSOM, R.W. & STEC, N.L. Cooperative modulation of [³H]MK-801 binding to the N-methyl-D-aspartate receptor-ion channel complex by L-glutamate, glycine and poliamines. **Journal of Neurochemistry**, **51**: 830-836, 1988.
- REBEC, G.V., BARTON, S.J., MARSEILLES, A.M., COLLINS, K. Ascorbate treatment attenuates the Huntington behavioral phenotype in mice. **NeuroReport**, **14**: 1263-1265, 2003.
- RIEDEL, G., WETZEL, W., REYMANN, K.G. Metabotropic glutamate receptor in spatial and nonspatial learning in rats studied by means of agonist and antagonist application. **Learning and Memory**, **2**: 243-265, 2003.

- RIEPE, M., HORI, N., LUDOLPH, A.C., CARPENTER, D.O., SPENCER, P.S., ALLEN, C.N. Inhibition of energy metabolism by 3-nitropropionic acid activates ATP-sensitive potassium channels. **Brain Research**, **586**: 61-66, 1992.
- ROBERTS, P.J., STORM-MATHISEN, J., JOHNSTON, G.A.R. **Glutamate: Transmitter in the Central Nervous System**. Wiley, New York, 1981.
- ROBINSON, M.B. & DOWD, L.A. Heterogeneity and functional properties of subtypes of sodium-dependent glutamate transporters in the mammalian central nervous system. **Advanced Pharmacology**, **37**: 69-115, 1996.
- ROEHRHS, C., GARRIDO-SANABRIA, E.R., DA SILVA, A.C., FARIA, L.C., SINHORIN, V.D.G., MARQUES, R.H., PRIEL, M.R., RUBIN, M.A., CAVALHEIRO, E.A., MELLO, C.F. Succinate increases neuronal post-synaptic excitatory potentials *in vitro* and induces convulsive behavior through *N*-methyl-D-aspartate-mediated mechanisms. **Neuroscience**, **125**: 965-971, 2004.
- ROSA, R.B., SCHWARZBOLD, C., DALCIN, K.B., GHISLENI, G.C., RIBEIRO, C.A.J., MORETTO, M.B., FRIZZO, M.E.S., HOFFMANN, G.F., SOUZA, D.O., WAJNER, M. Evidence that 3-hydroxyglutaric acid interacts with NMDA receptors in synaptic plasma membranes from cerebral cortex of Young rats. **Neurochemistry International**, **45**: 1087-1094, 2004.
- ROYES, L.F.F., FIGHERA, M.R., FURIAN, A.F., OLIVEIRA, M.S., DA SILVA, L.G.M., MALFATTI, C.R.M., SCHNEIDER, P.H., BRAGA, A.L., WAJNER, M., MELLO, C.F. Creatine protects against the convulsive behavior and lactate production elicited by the intrastriatal injection of methylmalonate. **Neuroscience**, **118**: 1079-1090, 2003.
- SANDERS, A.P., CURRIE, W.D., WOODHALL, B. Protection of brain metabolism with glutathione, glutamate, γ -aminobutyrate and succinate. **Proceedings of the Society for Experimental Biology and Medicine**, **130**: 1021-1027, 1969.
- SAUGSTAD, J.A., KINZIE, J.M., MULVIHILL, E.R., SEGERSON, T.P., WESTBROOK, G.L. Cloning and expression of a new member of the L-2-amino-4-phosphonobutyric acid-class of metabotropic glutamate receptors. **Molecular Pharmacology**, **45**: 367-372, 1994.
- SCATTON, B. The NMDA receptor complex. **Fundamental & Clinical Pharmacology**, **7**: 389-400, 1993.
- SCATTON, B., FROST, J., GEORGE, P., CARTER, C., BENAVIDES, J. Present developments in NMDA receptor antagonists against cerebral ischaemia. **Current Opinion in Therapeutics Pathology**, **4**: 523-545, 1991.
- SCHOEPP, D.D. & CONN, P.J. Metabotropic glutamate receptors in brain function and pathology. **Trends in Pharmacology**, **14**: 13-20, 1993.

- SCHOEPP, D.D., JANE, D.E., MONN, J.A. Pharmacological agents acting at subtypes of metabotropic glutamate receptors. **Neuropharmacology**, **38**: 1431-1477, 1999.
- SCHOEPP, D.D., JOHNSON, B.G., WRIGHT, R.A., SALHOFF, C.R., MAYNE, N.G., WU, S., COCKERHAM, S.L., BURNETT, J.P., BELEGAJE, R., BLEAKMAN, D., MONN, J.A. LY354740 is a potent and highly selective group II metabotropic glutamate receptor agonist in cells expressing human glutamate receptors. **Neuropharmacology**, **36**: 1-11, 1997.
- SCHOUSBOE, A. Transport and metabolism of glutamate and GABA in neurons and glial cells. **International Review of Neurobiology**, **22**: 1-45, 1981.
- SHEARDOWN, M.J., NIELSEN, E. φ., HANSEN, A.J., JACOBSEN, P., HONORÉ, T. 2,3-Dihydroxy-6-nitro-7-sulfamoyl-benzo(F)quinoxaline. A neuroprotectant for cerebral ischemia. **Science**, **247**: 571-574, 1990.
- SHIMAMOTO, K., LEBRUN, B., YASUDA-KAMATANI, Y., SAKAITANI, M., SHIGERI, Y., YUMOTO, N., NAKAJIMA, T. DL-threo-beta-benzylxyspartate, a potent blocker of excitatory amino acid transporters. **Molecular Pharmacology**, **53**: 195-201, 1998.
- SHINOZAKI, H., ISHIDA, M., SHIMAMOTO, K., OHFUNE, Y., Potent NMDA-like actions and potentiation of glutamate responses by conformational variants of a glutamate analogue in the rat spinal cord. **Brazilian Journal of Pharmacology**, **98**: 1213-1224, 1989.
- SINGH, L., OLES, R., WOODRUFF, G. In vivo interaction of a polyamine with NMDA receptor. **European Journal of Pharmacology**, **180**: 391-392, 1990.
- SMALL, B., THOMAS, J., KEMP, M., HOO, K., BALLYK, B., DEVERILL, M., OGDEN, A.M., RUBIO, A., PEDREGAL, C., BLEAKMAN, D. LY339434, a GluR5 kainate receptor agonist. **Neuropharmacology**, **37**: 1261-1267, 1998.
- SOUTHORN, P.A. & POWIS, G. Free radicals in medicine I. Nature and biologic reactions. **Mayo Clinic Proceedings**, **63**: 381-389, 1988.
- STENR, J.R., EGGLESTON, L.V., HEMS, R., KREBS, H.A. Accumulation of glutamic acid in isolated brain tissue. **Journal of Biochemistry**, **44**: 410-418, 1949.
- STORM-MATHISEN, J., ZHANG, N., OTTERSEN, O.P. Electron microscopic localization of glutamate, glutamine and GABA at putative glutamatergic and GABA-ergic synapses. **Molecular Neuropharmacology**, **2**: 7-13, 1992.
- SUCHER, N.J., AWOBULUYI, A., CHOI, Y.B., LIPTON, S.A. NMDA receptors: from genes to channels. **Trends in Pharmacological Sciences**, **17**: 348-355, 1996.
- SUTCLIFFE, M.J., WO, Z.G., OSWALD, R.E. Three-dimensional models of non-NMDA glutamate receptors. **Biophysical Journal**, **70**: 1575-1589, 1996.

- TANABE, S., MASU, M., ISHII, T., SHIGEMOTO, R., NAKANISHI, S. A family of metabotropic glutamate receptors. **Neuron**, **8**: 169-172, 1992.
- TEITELBAUM, J.S., ZATORRE, R.J., CARPENTER, S., GENDRON, D., EVANS, A.C., GJEDDE, A., CASHMAN, N.R. Neurologic sequelae of domoic acid intoxication due to the ingestion of contaminated mussels. **New England Journal of Medicine**, **322**: 1781-1787, 1990.
- TOYOSHIMA, S., WATANABE, F., SAIDO, H., MIYATAKE, K., NAKANO, Y. Methylmalonic acid inhibits respiration in rat liver mitochondria. **Journal of Nutrition**, **25**: 2846-2850, 1995.
- TSAI, M.J., CHANG, Y.-F., SCHWARCZ, R., BROOKES, N. Characterization of L- α -amino adipic acid transport in cultured rat astrocytes. **Brain Research**, **741**: 166-173, 1996.
- TÜCKMANTEL, W., KOZIKOWSKI, A.P., WANG, S., PSHENICHKIN, S., WROBLEWSKI, J.T. Synthesis, molecular modelling, and biology of the 1-benzyl derivative of APDC – an apparent mGluR6 selective ligand. **Bioorganic and Medicinal Chemistry Letters**, **7**: 601-606, 1997.
- URBANSKA, E.M., BLASZCZAK, P., SARAN, T., KLEINROK, Z., TURSKI, W.A. Mitochondrial toxin 3-nitropropionic acid evokes seizures in mice. **European Journal of Pharmacology**, **359**: 55-58, 1998.
- VIGNES, M., CLARKE, V.R., PARRY, M.J., BLEAKMAN, D., LODGE, D., ORNSTEIN, P.L., COLLINGRIDGE, G.L. The GluR5 subtype of kainate receptor regulates excitatory synaptic transmission in areas CA1 and CA3 of the rat hippocampus. **Neuropharmacology**, **37**: 1269-1277, 1998.
- XIA, X.G., SCHMIDT, N., TEISMANN, P., FERGER, B., SCHULZ, J.B. Dopamine mediates striatal malonate toxicity via dopamine transporter-dependent generation of reactive oxygen species and D2 but D1 receptor activation. **Journal of Neurochemistry**, **79**: 63-70, 2001.
- WAJNER, M., DUTRA, J.C., CARDOSO, S.E., WANNMACHER, C.M.D., MOTTA, E.R. Effect of methylmalonate on in vitro lactate release and carbon dioxide production by brain of suckling rats. **Journal of Inherited Metabolic Disease**, **15**: 92-96, 1992.
- WAJNER, M. & COELHO, J.C. Neurological dysfunction in methylmalonic aciduria is probably related to the inhibitory effect of methylmalonate on brains energy production. **Journal of Inherited Metabolic Disease**, **20**: 761-768, 1997.
- WILLIS, C.L., DAUENHAUER, D.L., HUMPHREY, J.M., CHAMBERLIN, A.R., BULLER, A.L., MONAGHAN, D.T., BRIDGES, R.J. Methylation of the NMDA receptor agonist L-trans-2,3-pyrrolidine-dicarboxylate: enhanced excitotoxic potency and selectivity. **Toxicology and Applied Pharmacology**, **144**: 45-55, 1997.

- WILLIS, C.L., HUMPHREY, J.M., KOCH, H.P., BLAKELY, T., RALSTON, L., BAKER, C.A., SHIM, S., KADRI, M. CHAMBERLIN, A.R., BRIDGES, R.J. L-Trans-2,3-Pyrrolidine Dicarboxylate: Characterization of a Novel Excitotoxin. **Neuropharmacology**, **35(5)**: 531-539, 1996.
- WILLIANS, K., ROMANO, C., DICHTER, M.A., MOLINOFF, P.B. Modulation of the NMDA receptor by poliamines. **Life Sciences**, **48**: 469-498, 1991.
- WATKINS, J.C. & EVANS, R.H. Excitatory amino acid transmitters. **Annual Review of Pharmacology and Toxicology**, **21**: 165-204, 1981.
- WATKINS, J.C., KROGSGAARD-LARSEN P., HONORE, T. Structure-activity relationships in the development of excitatory amino acid receptor agonists and competitive antagonists. **Trends in Pharmacological Science**, **11**: 25-33, 1990.
- YAMAKURA, T. & SHIMOJI, K. Subunit- and site-specific pharmacology of the NMDA receptor channel. **Progress in Neurobiology**, **56**: 279-298, 1999.
- YAMAMOTO, N., KABUTO, H., MATSUMOTO, S., OGAWA, N., YOKOI, I. α-Tocopheryl-L-ascorbate-2-O-phosphate diester, a hydroxyl radical scavenger, prevents the occurrence of epileptic foci in a rat model of post-traumatic epilepsy. **Pathophysiology**, **00**: 1-10, 2002.
- WERMUTH, C.G., MANN, A., SCHOENFELDER, A., WRIGHT, R.A., JOHNSON, B.G., BURNETT, J.P., MAYNE, N.G., SCHOEPP, D.D. (2S,4S)-2-amino-4-(4,4-diphenylbut-1-yl)pentane-1,5-dioic acid: A potent and selective antagonist for metabotropic glutamate receptors negatively linked to adenylate cyclase. **Journal of Medical Chemistry**, **39**: 814-816, 1996.
- ZEEVALK, G.D., DERR-YELLIN, E., NICKLAS, W.J. NMDA receptor involvement in toxicity to dopamine neurons in vitro caused by the succinate dehydrogenase inhibitor 3-nitropropionic acid. **Journal of Neurochemistry**, **64**: 455-458, 1995.
- ZIGMOND, M., BLOOM, F.E., LANDIS, S.C., ROBERTS, J.L., SQUIRE, L.R. **Fundamental Neuroscience**. Editora Academic Press, London, 1^a edição, 1999.