

**UNIVERSIDADE FEDERAL DE SANTA MARIA
CENTRO DE CIÊNCIAS NATURAIS E EXATAS
PROGRAMA DE PÓS-GRADUAÇÃO EM CIÊNCIAS BIOLÓGICAS:
BIOQUÍMICA TOXICOLÓGICA**

**DEXMEDETOMIDINA DIMINUI A RESPOSTA
INFLAMATÓRIA APÓS CIRURGIA MIOCÁRDICA
SOB MINI-CIRCULAÇÃO EXTRACORPÓREA**

TESE DE DOUTORADO

Neusa Maria Heinzmann Bulow

Santa Maria, RS, Brasil

2013

**DEXMEDETOMIDINA DIMINUI A RESPOSTA
INFLAMATÓRIA APÓS CIRURGIA MIOCÁRDICA SOB
MINI-CIRCULAÇÃO EXTRACORPÓREA**

Neusa Maria Heinzmann Bulow

Tese apresentada ao Curso de Doutorado do Programa de Pós-Graduação em Ciências Biológicas: Bioquímica Toxicológica da Universidade Federal de Santa Maria (UFSM, RS), como requisito parcial para obtenção do grau de **Doutora em Bioquímica Toxicológica.**

Orientador: Prof. Dr. João Batista Teixeira da Rocha

Santa Maria, RS, Brasil

2013

Ficha catalográfica elaborada através do Programa de Geração Automática da Biblioteca Central da UFSM, com os dados fornecidos pelo(a) autor(a).

Bulow, Neusa Maria Heinzmann
Dexmedetomidina diminui a resposta inflamatória após Cirurgia Miocárdica sob Mini-Circulação Extracorpórea / Neusa Maria Heinzmann Bulow.-2013.
159 p.; 30cm

Orientador: João Batista Teixeira da Rocha
Tese (doutorado) - Universidade Federal de Santa Maria, Centro de Ciências Naturais e Exatas, Programa de Pós-Graduação em Ciências Biológicas: Bioquímica Toxicológica, RS, 2013

1. Dexmedetomidina 2. Inflamação 3. Estresse Oxidativo
4. Anestesia Intravenosa Total (AIVT) 5. Circulação Extracorpórea (CEC) I. Teixeira da Rocha, João Batista
II. Título.

**Universidade Federal de Santa Maria
Centro de Ciências Naturais E Exatas
Programa de Pós-Graduação em Ciências Biológicas: Bioquímica
Toxicológica**

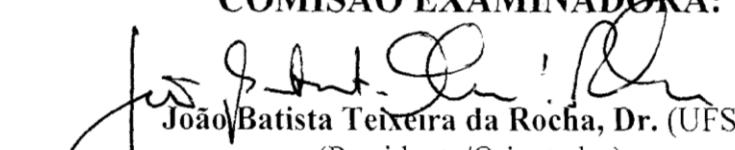
A Comissão Examinadora, abaixo assinada, aprova a Tese de Doutorado

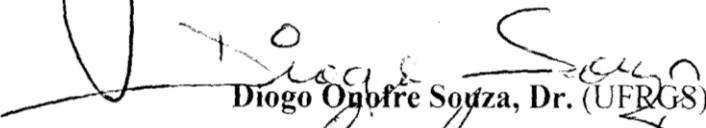
**DEXMEDETOMIDINA DIMINUI A RESPOSTA INFLAMATÓRIA
APÓS CIRÚRGIA MIOCÁRDICA SOB MINI-CIRCULAÇÃO
EXTRACORPÓREA**

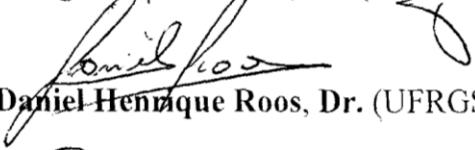
Elaborada por
Neusa Maria Heinzmann Bulow

como requisito parcial para obtenção do grau de
Doutora em Bioquímica Toxicológica

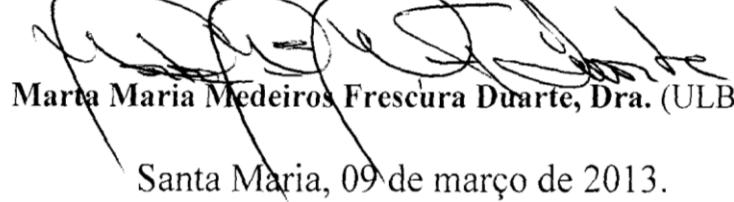
COMISÃO EXAMINADORA:


João Batista Teixeira da Rocha, Dr. (UFSM)
(Presidente/Orientador)


Diogo Onofre Souza, Dr. (UFRGS)


Daniel Henrique Roos, Dr. (UFRGS)


Maria Rosa Chitolina Schetinger, Dra. (UFSM)


Marta Maria Medeiros Frescura Duarte, Dra. (ULBRA)

Santa Maria, 09 de março de 2013.

DEDICATÓRIA

Aos meus filhos Mateus e Arthur, amo vocês!

AGRADECIMENTOS

Agradeço ao Ernani, por seu amor, carinho e companheirismo.

Agradeço aos meus pais, Lúcia e Luiz, que moldaram em mim um caráter forte e a noção clara de que bons resultados advêm do trabalho e da dedicação.

Agradeço aos meus queridos irmãos, Germano, Gilberto, Marcos, Vicente, Joaquim e Daniel, por serem exemplos de pessoas de bem e pelo apoio incondicional.

Gostaria de expressar a minha mais profunda gratidão ao meu orientador Prof. PhD. João Batista Teixeira da Rocha pela sua disponibilidade, paciência, pelos ensinamentos e pelo estímulo à conclusão deste trabalho.

Aos amigos queridos, sempre presentes de alguma maneira. Vocês alegram meus dias.

Um carinho especial àqueles que contribuíram para a realização deste trabalho: Marta, Elisângela, Eduardo, Rochelle, Anelise, Emily, Romaiana, Ana Lima, Ralf, Roberta, Mariane, Andréia, Darlan, Ellen. Sem sua ajuda, a realização deste estudo seria impossível.

A todos os professores do Programa de Pós-Graduação em Ciências Biológicas: Bioquímica Toxicológica, pelo seu estímulo, trazendo aos alunos o gosto pela experimentação.

Ao CNPq e FAPERGS pelo suporte financeiro.

Aos pacientes que se disponibilizaram a participar deste estudo todo o meu respeito, e desejo que outros deles possam se beneficiar do objetivo real das investigações aqui conduzidas.

Agradeço também à Universidade Federal de Santa Maria e ao Programa de Pós-Graduação em Ciências Biológicas: Bioquímica Toxicológica, pela possibilidade de realização deste curso.

“Sabemos de quase nada adequadamente, de poucas coisas *a priori*, e da maioria por meio da experiência”.

Gottfried Wilhelm Leibniz

RESUMO

Tese de Doutorado

Programa de Pós-Graduação em Ciências Biológicas: Bioquímica Toxicológica
Universidade Federal de Santa Maria, RS, Brasil

DEXMEDETOMIDINA DIMINUI A RESPOSTA INFLAMATÓRIA APÓS CIRURGIA MIOCÁRDICA SOB MINI-CIRCULAÇÃO EXTRACORPÓREA

AUTORA: Neusa Maria Heinzmann Bulow

ORIENTADOR: João Batista Teixeira da Rocha

LOCAL E DATA DA DEFESA: Santa Maria, 09 de Março de 2013

Apesar dos grandes avanços tecnológicos nas cirurgias de revascularização miocárdica (CRM), ocorre uma grande incidência de disfunção cardíaca e déficit neurocognitivo no período pós-operatório. As medidas preventivas são essenciais para a redução destas situações adversas, responsáveis pelo comprometimento da qualidade de vida dos pacientes. A cirurgia e a circulação extra-corpórea (CEC) produzem alterações importantes no sistema imunológico, diretamente envolvidas na incidência das complicações e acredita-se que a escolha anestésica possa modificá-las. Em estudo prospectivo e randomizado, pretendemos demonstrar a influência da dexmedetomidina (grupo AIVT-DEX), um anestésico (α)-2-agonista, associado à anestesia intravenosa total (AIVT) no comportamento da resposta inflamatória em pacientes submetidos à CRM, sob mini-circulação extracorpórea (mini-CEC). O grupo AIVT-DEX recebeu infusão contínua de dexmedetomidina associado à técnica de AIVT convencional e o outro grupo foi submetido à AIVT convencional (infusão contínua de propofol e sufentanil). Os grupos foram comparados pela dosagem plasmática trans-operatória de citocinas, como a interleucina-1(IL-1), a interleucina-6 (IL-6), a interleucina-10 (IL-10), o interferon gama (INF- γ) e o fator de necrose tumoral alfa (TNF- α), bem como a proteína C reativa (PCR), creatinofosfoquinase (CPK), creatinofosfoquinase miocárdio específica (CPK-MB), troponina I (cTnI), cortisol e glicose. A peroxidação lipídica foi avaliada pelo estudo das substâncias reativas ao ácido tiobarbitúrico (TBARS) e a presença de estresse oxidativo pela atividade da enzima delta-aminolevulinato desidratase (δ -ALA-D). O uso da dexmedetomidina induziu redução significativa de IL-1, IL-6, TNF- α e INF- γ se comparado ao grupo sem dexmedetomidina. Houve redução progressiva dos níveis de IL-10 ao longo do tempo, de forma semelhante entre os grupos. Não houve diferença entre os grupos para a atividade da enzima δ -ALA-D e os níveis de TBARS foram maiores no grupo AIVT-DEX. Concluímos que a dexmedetomidina associada à AIVT convencional foi capaz de reduzir os níveis plasmáticos das citocinas pró-inflamatórias IL-1, IL-6, TNF- α e INF- γ em pacientes submetidos à CRM sob mini-CEC, se comparados aos pacientes que receberam apenas a AIVT convencional. Estes resultados reforçam os dados da literatura quanto à potencialidade da dexmedetomidina como agente modulador da resposta inflamatória no período trans-operatório.

Palavras Chave: Dexmedetomidina. Inflamação. Estresse Oxidativo. Anestesia Intravenosa Total (AIVT). Circulação Extracorpórea (CEC).

ABSTRACT

Thesis of PhD's Degree
 Graduate Course in Biological Sciences: Toxicological Biochemistry
 Federal University of Santa Maria, RS, Brazil

DEXMEDETOMIDINE DECREASE THE INFLAMMATORY RESPONSE TO MYOCARDIAL SURGERY UNDER MINI CARDIOPULMONARY BYPASS

AUTHOR: NEUSA MARIA HEINZMANN BULOW

ADVISER: João Batista Teixeira da Rocha

PLACE AND DATE OF THE DEFENSE: Santa Maria, 09 March of 2013

Despite great technological advances in coronary artery bypass grafting (CABG) surgery, there is a high incidence of cardiac dysfunction and neurocognitive deficits in the postoperative period. Preventive measures are essential to reducing these adverse situations that are responsible for significant morbidity and impairment on life quality of these patients. Surgery and cardiopulmonary bypass (CPB) produces important changes in the immune system, directly involved in the incidence of these complications and is credible that anesthesia choice can it modified. We hypothesized that dexmedetomidine, an (α)-2-agonist, could the inflammatory response to CABG and CPB modified. In a prospective and randomized study, we intend to demonstrate the influence of dexmedetomidine (TIVA-DEX group), as a component of a conventional total intravenous anesthesia (TIVA-propofol+sufentanil) in patients undergoing CABG, with mini-CPB, on the behavior of this inflammatory response. The TIVA-DEX group received a continuous infusion of dexmedetomidine associated to a conventional venous anesthesia (continuous infusion of propofol+sufentanil). Intraoperative dosage of cytokines, such as interleukin-1 (IL-1), interleukin-6 (IL-6), interleukin-10 (IL-10), gamma interferon (INF- γ) and tumor necrosis factor (TNF- α) were performed, and also C-reactive protein (CRP), creatine phosphokinase (CPK), creatine phosphokinase-MB (CPK-MB), I troponin (cTnI), cortisol and glucose. The occurrence of lipid peroxidation, by the study of thiobarbituric acid reactive substances (TBARS) and the activity of delta-aminolevulinate dehydratase (δ -ALA-D) to oxidative stress verify were also evaluated. Dexmedetomidine induce a significative reduction of IL-1, IL-6, TNF- α and INF- γ , as compared to group that not receive dexmedetomidine. The levels of IL-10 were decreased in both groups along the time, at a similar pattern. Differences between groups on δ -ALA-D activity do not occur and TBARS was higher in TIVA-DEX group. We concluded that dexmedetomidine associated to TIVA was able to reduce plasma levels of proinflammatory cytokines IL-1, IL-6, TNF- α and INF- γ in patients submitted to CABG surgery under mini-CPB, as compared to a conventional TIVA. These results reinforce literature data about dexmedetomidine potentiality as an anti inflammatory agent.

Keywords: Dexmedetomidine. Inflammation. Oxidative Stress. Total Intravenous Anesthesia (TIVA). Cardiopulmonary Bypass (CPB).

LISTA DE ILUSTRAÇÕES

INTRODUÇÃO

Figura 1- Sistema convencional de circulação extracorpórea.....	22
Figura 2 - Comparação entre os sistemas de MECC (Mini-Extracorporeal Circulation) e SECC (Standard Extracorporeal Circulation).....	23
Figura 3 - Fase de estímulo dos neutrófilos e células endoteliais pelos fatores inflamatórios circulantes.....	28
Figura 4 - Efeito da dexmedetomidina sobre os receptores (α)-2-adrenérgicos pré e pós-sinápticos.....	34
Figura 5 - Efeitos clínicos induzidos pelo uso da dexmedetomidina e os receptores específicos envolvidos.....	36
Esquema 1 - Resposta inflamatória gerada pela circulação extracorpórea que se assemelha à síndrome da resposta inflamatória sistêmica (SIRS).....	20
Esquema 2 - Complexa cascata inflamatória relacionada à isquemia/reperfusão.....	25
Esquema 3 - A resposta inflamatória decorrente da Circulação Extra Corpórea (CPB-Cardiopulmonary Bypass) está dividida em duas fases: fase precoce (early phase) e fase tardia (late phase).....	26

MANUSCRITO 1

Figure 1 - Standard extracorporeal circulation system.....	92
Figure 2 - Cardiopulmonary bypass neutrophil activation.....	93
Figure 3 - Mini-extracorporeal circulation system compared to standard extracorporeal circulation system.....	94
Figure 4 - Dexmedetomidine and clonidine structural formulae.....	94
Figure 5 - Dexmedetomidine clinical effects mediated via activation of (α)-2-adrenergic and imidazoline receptors.....	95
Figure 6 - Dexmedetomidine can exert its effects via activation of three (α)-2-adrenoceptor subtypes.....	96
Figure 7 - Putative intracellular mechanisms involved in the (α)-2-adrenoreceptors activation.....	97
Figure 8 - Neuroprotective mechanism(s) triggered by (α)-2-adrenoreceptors agonists.....	98
Scheme 1 - Cardiopulmonary bypass and the extracorporeal circulation responses with the pathophysiologic changes resembling the systemic inflammatory response syndrome (SIRS).....	99

Scheme 2 - The inflammatory response to cardiopulmonary bypass is divided into 2 phases: “early” and “late” phases.....	100
Scheme 3 - Complex cascade of pathophysiologic phenomena associated with ischemia/reperfusion in CABG.	101

MANUSCRITO 2

Figure 1 - Sampling protocol.....	122
Figure 2 - Mean arterial pressure (MAP) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX)	123
Figure 3 - Statistical analysis heart rate (HR) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).....	123
Figure 4 - Hematocrit (HT) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	124
Figure 5 - Hemoglobin (HB) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	124
Figure 6 - Plasma interleukin-1 (IL-1) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	125
Figure 7 - Plasma interleukin-6 (IL-6) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	125
Figure 8 - Plasma interleukin-10 (IL-10) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	126
Figure 9 - Plasma gamma interferon (INF- γ) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	126
Figure 10 - Plasma alpha-tumoral necrosis factor (TNF- α) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).....	127
Figure 11- Erithrocytic thiobarbituric acid reactive substances (TBARS) (TBA) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	127
Figure 12 - Plasma C-reactive protein (PCR) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).....	128

Figure 13 - Plasma creatine phosphokinase (CPK) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).....	128
Figure 14 - Plasma MB-creatine phosphokinase (MB-CPK) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).....	129
Figure 15 - Plasma troponin (cTn-I) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX)	129
Figure 16 - Plasmatic cortisol of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	130
Figure 17 - Plasmatic glucose of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).	130
Figure 18 - Mini mental state examination (MMSE) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX).....	131

LISTA DE TABELAS**MANUSCRITO 2**

Table 1 - Patients demographic characteristics and surgery related parameters	132
Table 2 - Haemodynamics perioperative parameters.	133
Table 3 - Hemodilution of patients in collected times.....	133
Table 4 - TBARS and δ - ALA-D activity in collected times.	134

LISTA DE ABREVIATURAS E SIGLAS

- δ-ALA-D - δ-aminolevulinato desidratase
ADP - difosfato de adenosina
AIVT- anestesia intravenosa total
ATP - trifosfato de adenosina
AVC - acidente vascular cerebral
BIS - índice biespectral
CEC - circulação extracorpórea
CPK - creatino fosfoquinase
CPK-MB - creatino fosfoquinase miocárdio específica
CRM - cirurgia de revascularização miocárdica
cTnI - troponina
DNA - ácido desoxirribonucleico
EROs - espécies reativas de oxigênio
H₂O₂ - peróxido de hidrogênio
ICAM - intercellular adhesion molecule
IgE - imunoglobulina E
IL-1 - interleucina-1
IL-6 - interleucina-6
IL-8 - interleucina-8
IL-10 - interleucina-10
INF-γ - interferon gama
NOS - enzima óxido nítrico sintase
I/R - isquemia/reperfusão
MECC - mini extracorporeal circulation
MEEM - mini exame do estado mental
NADPH - nicotinamida adenina dinucleotide fosfato
NADPH-oxidase - nicotinamida adenina dinucleotide fosfato oxidase
NO - óxido nítrico
OONO - peroxinitrito
O₂ - radical superóxido
PaCO₂ - pressão arterial de gás carbônico

PAF - fator de ativação plaquetária
PCR - proteína C reativa
PECAM - platelet/ intercellular adhesion molecule
ROS: reactive oxygen species
SECC- standard extracorporeal circulation
SNS - sistema nervoso simpático
SOD - superóxido dismutase
SIRS: systemic inflammatory response syndrome
SRIS - síndrome da resposta inflamatória sistêmica
TNF- α - fator de necrose tumoral alfa
TBARS - substâncias reativas ao ácido tiobarbitúrico
UFSM - Universidade Federal de Santa Maria
UTI- unidade de tratamento intensivo

SUMÁRIO

1 INTRODUÇÃO	19
1.1 CIRCULAÇÃO EXTRA-CORPÓREA CONVENCIONAL (CEC-SECC) E MINI-CIRCULAÇÃO EXTRA-CORPÓREA (MINI-CEC-MECC)	21
1.2 RESPOSTA INFLAMATÓRIA EM CIRURGIA CARDÍACA.....	24
1.3 MÉTODOS PARA REDUZIR A SRIS PÓS-CEC.....	30
1.4 IMUNOMODULAÇÃO TRANS-OPERATÓRIA PELOS ANESTÉSICOS.....	32
1.5 JUSTIFICATIVA	37
1.6 OBJETIVOS	38
1.6.1 Objetivo Geral	38
1.6.2 Objetivos Específicos	38
2 MANUSCRITOS.....	39
2.1 MANUSCRITO 1 - RESPOSTA INFLAMATÓRIA EM PACIENTES SUBMETIDOS À CIRURGIA DE REVASCULARIZAÇÃO MIOCÁRDICA (CRM) E IMPLICAÇÕES CLÍNICAS: UMA REVISÃO DA RELEVÂNCIA DO USO DE DEXMEDETOMIDINA	40
2.1.1 Summary	42
2.1.2 Introduction.....	42
2.1.2.1 Inflammatory response and ischemia/reperfusion in CABG surgery	43
2.1.3 Cardiopulmonary bypass (CPB)	44
2.1.3.1 Mini-extracorporeal circulation (MECC)	46
2.1.3.2 Oxidative stress and inflammation associated with coronary artery bypass grafting surgery (CABG)	47
2.1.3.3 Neuroinflammation associated with coronary artery bypass grafting surgery (CABG).....	49
2.1.3.4 S-100B as a marker and modulator of neuroinflammation	51
2.1.4 Alpha(α)-2-adrenergic receptor agonists	52
2.1.4.1 Clonidine.....	53
2.1.4.2 Dexmedetomidine	54
2.1.4.3 Dexmedetomidine pharmacokinetics.....	54
2.1.4.4 Dexmedetomidine analgesic and sedative effects	55
2.1.4.5 Antiinflammatory effects of dexmedetomidine	57
2.1.4.6 Neuroprotective effects of dexmedetomidine.....	60
2.1.4.7 Dexmedetomidine as protective agent against ischemia	64
2.1.4.8 Dexmedetomidine hemodynamic and myocardial protective effects.....	67
2.1.4.9 Dexmedetomidine other potential effects	69
2.1.5 Conclusions.....	70

2.1.6 References.....	70
2.2 MANUSCRITO 2 - DEXMEDETOMIDINA REDUZ A RESPOSTA INFLAMATÓRIA APÓS CIRURGIA MIOCÁRDICA SOB MINI-CIRCULAÇÃO EXTRACORPÓREA.....	102
2.2.1 Abstract.....	104
2.2.2 Introduction.....	105
2.2.3 Materials and Methods	106
2.2.4 Results	108
2.2.5 Discussion.....	111
2.2.6 Conclusions.....	115
2.2.7 References.....	116
3 DISCUSSÃO	135
4 CONCLUSÕES	140
REFERÊNCIAS	141

APRESENTAÇÃO

No item **INTRODUÇÃO**, está descrita uma breve apresentação sobre os temas trabalhados nesta tese.

Uma revisão sobre a resposta inflamatória induzida pela circulação extracorpórea e a discussão sobre as potencialidades do anestésico dexmedetomidina como agente modulador desta resposta é apresentada como **MANUSCRITO 1**.

Os resultados que fazem parte desta tese estão apresentados sob a forma de artigo, o qual se encontra no item **MANUSCRITO 2**. As seções Materiais e Métodos, Resultados, Discussão dos Resultados e Referências Bibliográficas, encontram-se nos próprios manuscritos e representam os resultados finais deste estudo.

Os itens, **DISCUSSÃO E CONCLUSÕES**, encontram-se no final desta tese, apresentam interpretações e comentários gerais sobre os artigos científicos contidos neste trabalho.

As **REFERÊNCIAS BIBLIOGRÁFICAS** referem-se somente às citações que aparecem nos itens **INTRODUÇÃO, DISCUSSÃO e CONCLUSÕES** desta tese.

1 INTRODUÇÃO

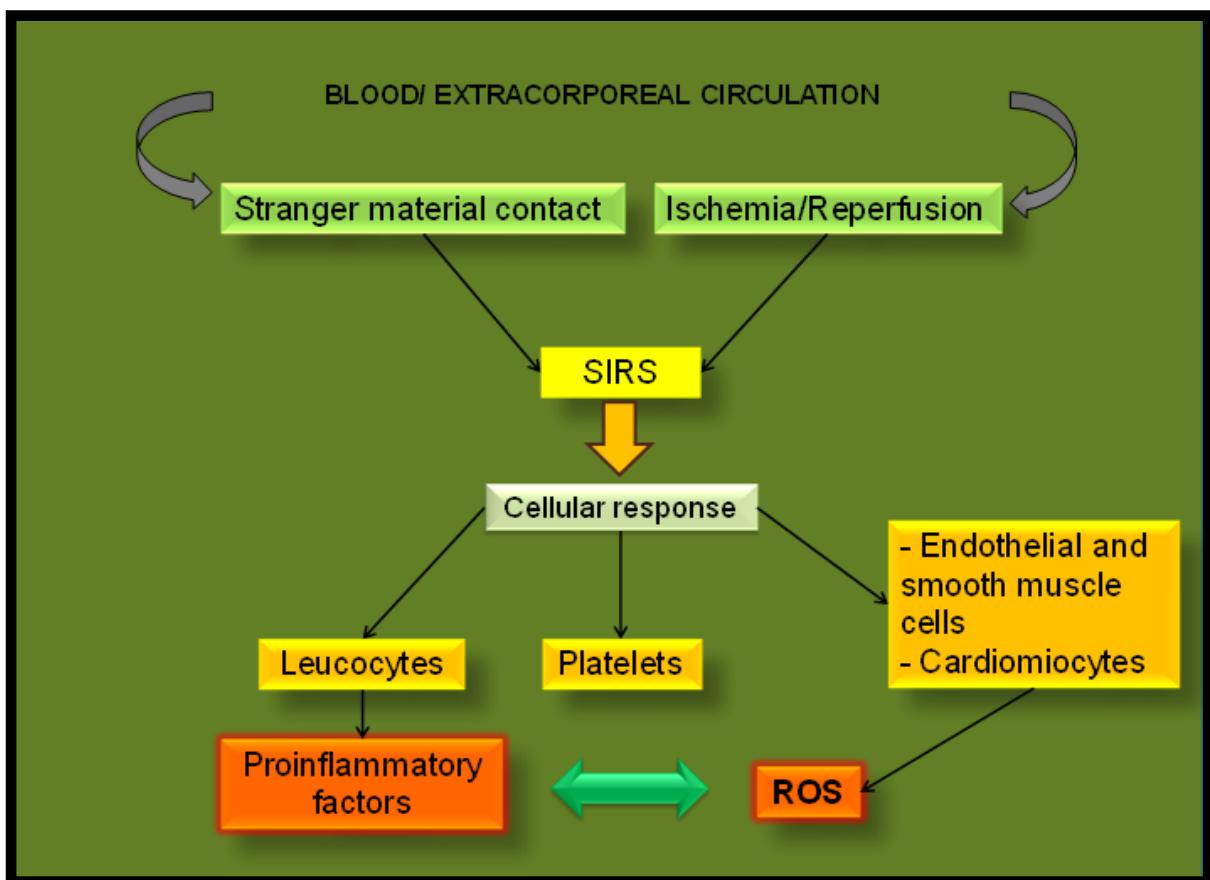
Durante os procedimentos cirúrgicos cardiológicos, devido à agressão tecidual, como uma resposta fisiológica, ocorre o aumento agudo de citocinas pró-inflamatórias, a redução dos níveis de citocinas anti-inflamatórias, o aumento de metabólitos do ácido araquidônico, de espécies reativas de oxigênio (EROs) e de outros mediadores (STEINBERG e cols., 1993; CASEY e cols., 1993; FRANGOGIANNIS e cols., 1998; SAVARIS e cols., 2001; SANDER e cols., 2006; WARREN e cols., 2009; PERRY e cols., 2010). Vários métodos são utilizados para minimizar esta resposta, com o intuito de melhorar as condições pós-operatórias dos pacientes e a técnica anestésica utilizada pode interferir nos mecanismos envolvidos (CROZIER e cols., 1994), sendo objeto interessante para estudo e pesquisa.

Na verdade, em pacientes submetidos à cirurgia de revascularização miocárdica (CRM) sob circulação extra-corpórea (CEC), pelo contato do sangue com superfícies não endoteliais e pela reperfusão de órgãos isquêmicos ocorrem alterações que se assemelham à síndrome da resposta inflamatória sistêmica (SIRS) (FRANGOGIANNIS e cols., 1998; de MOURA e cols., 2001; WARREN e cols., 2009) (Esquema 1). A SIRS caracteriza-se por temperatura corporal $> 38^{\circ}\text{C}$ ou $< 36^{\circ}\text{C}$, freqüência cardíaca >90 batidas por minuto, freqüência respiratória > 20 inspirações por minuto ou pressão arterial de gás carbônico (PaCO_2) $<32\text{mmHg}$, contagem de glóbulos brancos >12.000 ou < 4000 ou $>10\%$ das formas imaturas (De MOURA e cols., 2001).

A liberação de citocinas pró-inflamatórias (interleucina-1(IL-1), interleucina-6 (IL-6), fator de necrose tumoral alfa (TNF- α) e interferon gama (INF- γ)), é responsável por induzir febre, neutrofilia e modular a produção de outras citocinas pelos monócitos e neutrófilos, correlacionando-se com maior mortalidade pós-operatória (SAVARIS e cols., 2001; SANDER e cols., 2006). As citocinas anti-inflamatórias, ao contrário, têm papel regulador importante na redução da liberação das interleucinas pró- inflamatórias. A interleucina-10 (IL-10) inibe a síntese do TNF- α , IL-1, IL-6 e IL-8 em monócitos e macrófagos (SABLOTZKI e cols. 1997). Sander e colaboradores (2006) confirmaram em seus estudos, achados anteriores de que a cirurgia cardíaca com CEC leva ao aumento pós-operatório da IL-10 (SABLOTZKI e cols., 1997; SANDER e cols., 2006) e relataram também aumentos significativos para IL-10 em pacientes que desenvolveram infecção no período pós-operatório. O aumento da IL-10 tem sido descrito como preditivo de evolução desfavorável

após a cirurgia (GALLEY e cols., 2003). Seu aumento parece estar correlacionado à infecção e sépsis após trauma ou cirurgia (LYONS e cols., 1997) e indica aumento da mortalidade (VANDISSEL e cols., 1998) sendo que um estado anti-inflamatório acentuado também parece ser prejudicial à evolução clínica do paciente (GOGOS e cols., 2000).

As espécies reativas de oxigênio (EROs) também possuem papel fundamental no aumento de complicações pós-operatórias que ocorrem em cirurgias de revascularização miocárdica sob CEC (BOLLI e cols., 1988; MAULIK e cols., 1998). Com um ou mais elétrons não pareados, tornam-se altamente reativas e exercem seus efeitos nocivos sobre as células, obtendo elétrons de outras moléculas que poderão ser de lipídios, de proteínas e mesmo do ácido desoxirribonucleico (DNA) (MARCZIN e cols., 2003; ELAHI e cols., 2008).



Esquema 1 - Resposta inflamatória gerada pela circulação extracorpórea que se assemelha à síndrome da resposta inflamatória sistêmica (SRIS). O contato do sangue com os materiais estranhos do circuito de circulação extracorpórea, a isquemia/reperfusão e a indução pela hiperoxigenação, geram uma resposta semelhante à SRIS. Esta resposta está associada com a ativação patológica de leucócitos, plaquetas (que contribuem para o aumento da coagulação), células endoteliais e cardiomiócitos. A secreção de fatores pró-inflamatórios pelos leucócitos e o aumento da tensão sanguínea de oxigênio estimulam a produção anormal de espécies reativas de oxigênio (EROs), o que leva a um ciclo inflamatório vicioso. SIRS: systemic inflammatory response syndrome; ROS: reactive oxygen species.

1.1 Circulação Extra-Corpórea Convencional (CEC-SECC) e Mini-Circulação Extra-Corpórea (Mini-CEC-MECC)

A circulação extracorpórea (CEC) (Figura 1) é um procedimento que surgiu na década de cinquenta, com o objetivo de simular, mecanicamente, as funções do coração e manter a oxigenação do sangue, permitindo aos cirurgiões, examinar e corrigir as lesões cardíacas com maior especificidade. John e Mary Gibbon (GIBBON, 1970; GIBBON, 1971), depois de várias pesquisas e experiências, montaram um efetivo sistema de respiração e circulação artificiais. Com o passar dos anos, os aparelhos foram sendo modificados, com a utilização de materiais de maior biocompatibilidade e a introdução de outros mecanismos que possibilitaram uma dinâmica mais adequada do sistema circulatório (GALLETTI e BRECHER, 1962; GOMES e CONCEIÇÃO, 1985). Hoje, a CEC é de grande utilização, nas mais diversas cirurgias, permitindo intervenções em recém-nascidos, em crianças, em portadores de lesões múltiplas e/ou graves, em idosos com doenças sistêmicas associadas e em cirurgias para transplantes cardíacos.

Apesar de a CEC ter solucionado os obstáculos que impediam o acesso às cavidades do coração, apresenta um conjunto de complicações advindas da resposta do organismo às agressões impostas por seu mecanismo pouco fisiológico. Dentre os problemas, os principais são a importante resposta inflamatória apresentada pelos pacientes, a hemodiluição excessiva e/ou hemólise, a necessidade de uso de sangue homólogo para o preenchimento do circuito de CEC e a lesão da microcirculação, causada pelo tipo de fluxo induzido pelas bombas do aparelho. A hemodiluição apresenta efeitos indesejados, tais como a redução da pressão osmótica sangüínea e da capacidade de carrear oxigênio, causando acidose, hipóxia, edema e alterações na coagulação (GIBBON, 1954).

Durante a CEC, a circulação é totalmente modificada pela indução de um fluxo não pulsátil do lado arterial, passando o fluxo capilar a ser contínuo, o que aumenta a pressão do lado venoso. Este fluxo contínuo leva à adaptação celular que resulta no desenvolvimento de uma resposta inflamatória semelhante à síndrome da resposta inflamatória sistêmica (SRIS) (De SOUZA e ELIAS, 2006). Ocorrem formações de microbolhas, que causam obstrução dos capilares, promovendo isquemia, inflamação, ativação de complemento e da agregação plaquetária (BARAK e KATZ, 2005).

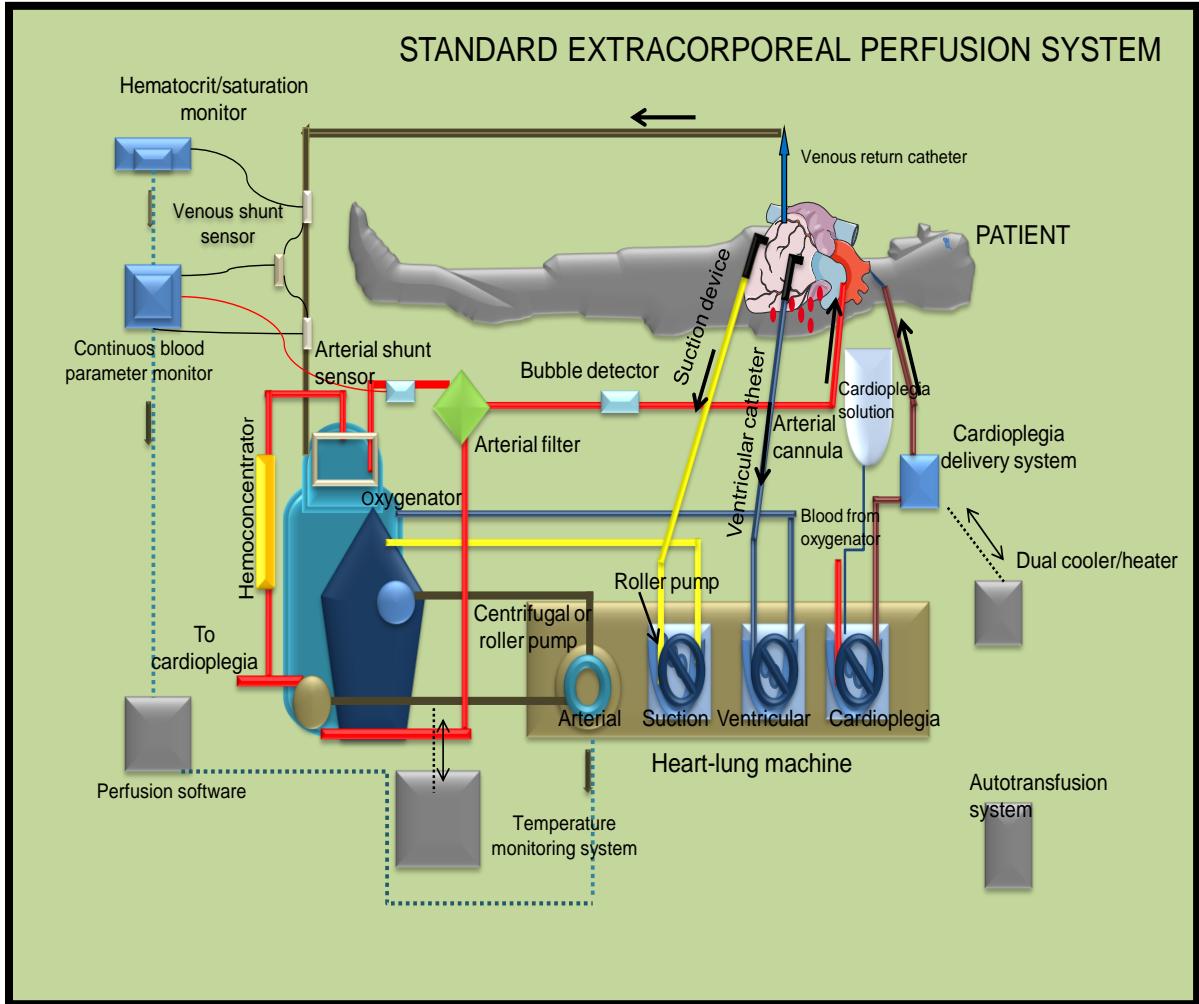


Figura 1 - Sistema Convencional de Circulação Extracorpórea. Em cirurgias com parada circulatória e uso de circulação extracorpórea, o sangue é desviado do coração e pulmões, passando por um oxigenador externo e por amplo sistema de tubulações e bombas que permitem a oxigenação dos tecidos, durante a cirurgia.

A SRIS não é complicação exclusiva da CEC em cirurgias cardíacas, mas a CEC continua sendo o maior fator envolvido em seu aparecimento (BUTLER e cols., 1993; NIEMAN e cols., 1999). Na tentativa de reduzir esta resposta, surgiram os circuitos menores, fechados, que parecem reduzir a hemodiluição e as superfícies de contato do sangue com material estranho. A evolução destes sistemas de CEC levou ao desenvolvimento do sistema de mini-CEC, ou sistema de MECC (Mini-Extracorporeal Circulation). Este circuito pode oferecer algumas vantagens, tais como uma menor exposição do sangue a componentes estranhos pela sua menor extensão, por ser fechado sem contato com o ar, o que também diminui as alterações celulares, por ter a necessidade de volume inicial menor para preencher

o sistema (*priming*), levando a menor hemodiluição e pelo uso de bombas centrífugas, o que diminui a lesão celular e a resposta inflamatória (Figura 2).

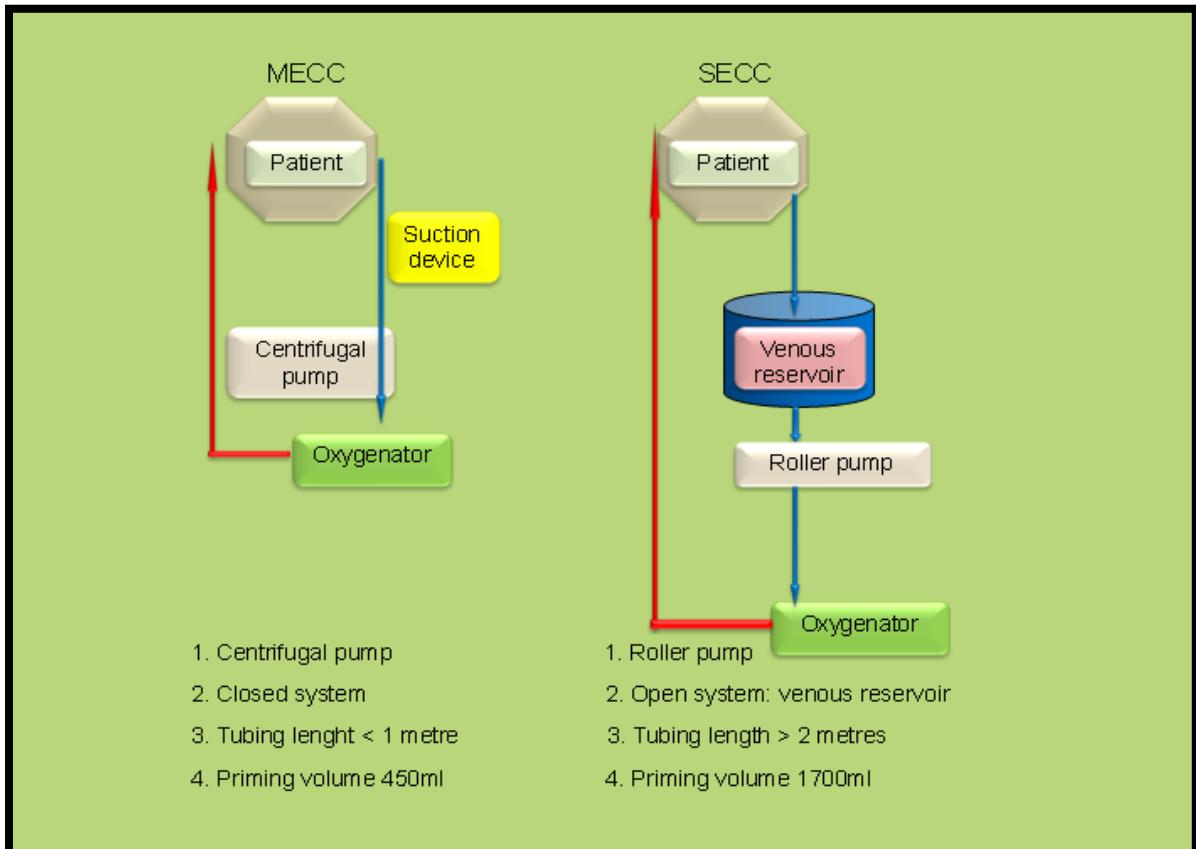


Figura 2 - Comparaçao entre os sistemas de MECC (Mini-Extracorporeal Circulation) e SECC (Standard Extracorporeal Circulation). No sistema de MECC, existem vantagens por ser um circuito com menor extensão, usar bomba centrífuga, usar um sistema fechado onde o sangue não entra em contato com o ar e ter uma necessidade de *priming* menor que o sistema convencional (SECC).

Demonstrou-se inicialmente que o sistema de mini-CEC pode reduzir a SRIS se comparado com os circuitos convencionais (FROMES e cols., 2002; REMADI e cols., 2006). Contudo, apesar de alguns resultados promissores, ainda não parece existir consenso sobre as vantagens destes circuitos miniaturizados sobre a resposta inflamatória. Em estudo de Bical e colaboradores (BICAL e cols., 2006), os níveis de proteína C reativa (PCR) e interleucina-6 (IL-6) aumentaram sem haver diferenças entre o grupo submetido aos circuitos convencionais de CEC e o grupo submetido à mini-CEC, contudo, ocorreu menor liberação de interleucina-10 (IL-10) com o uso de mini-CEC. Outros autores mostram inclusive não haver diferença para estes marcadores (PCR e IL-6) entre grupos de revascularização miocárdica com CEC se

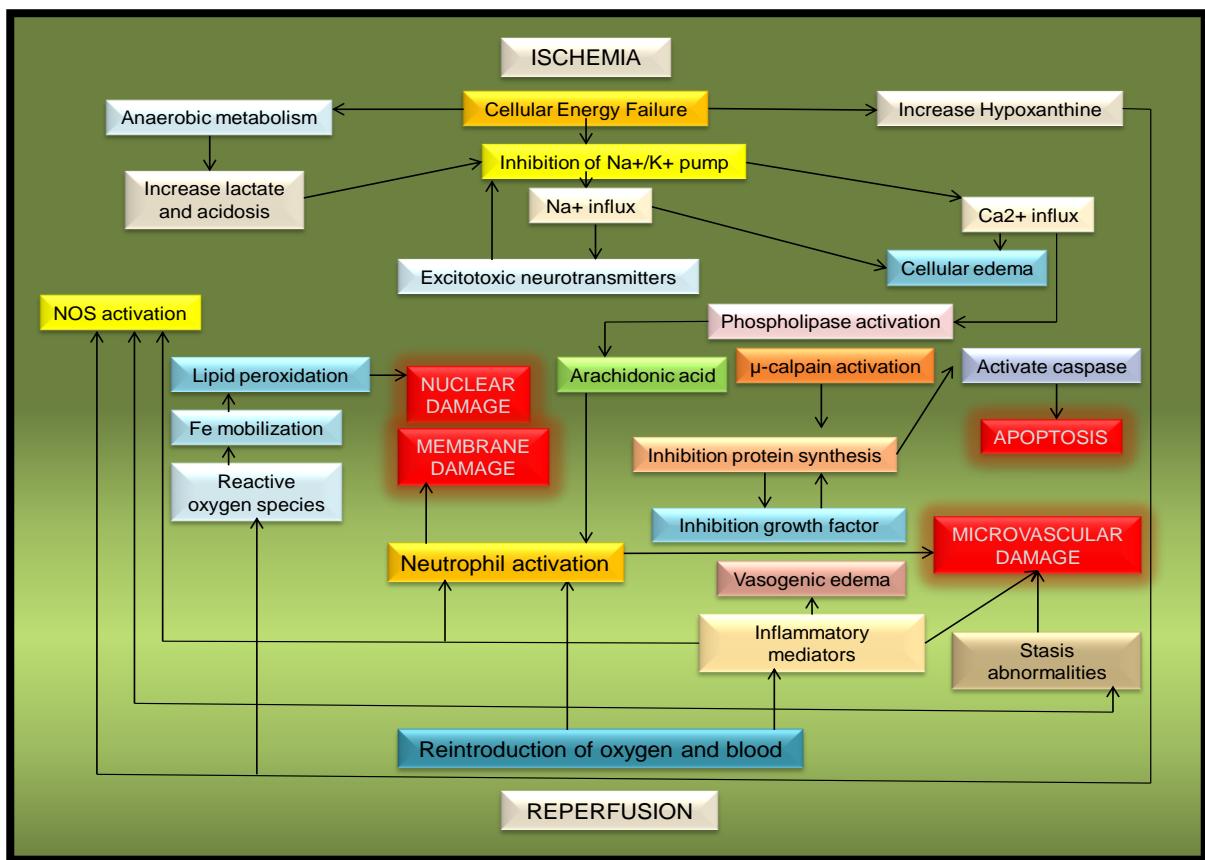
comparados ao grupo sem CEC (ASCIONE e cols., 2000; FRANKE e cols., 2005), sugerindo que a PCR e IL-6 possam ser, na verdade, marcadores ativados principalmente pelo trauma cirúrgico e não propriamente pela CEC. O fator de necrose tumoral- α (TNF- α) e a elastase parecem ser mais sensíveis em avaliar a SRIS pós-CEC (TORRE-AMIONE e cols., 1995; EPPINGER e cols., 1996; TANG e cols., 2004). Foi relatada uma menor liberação de TNF- α e elastase em pacientes submetidos ao sistema de mini-CEC se comparado a circuitos convencionais (FROMES e cols., 2002).

A disfunção orgânica após a CEC, tendo como base a SRIS, é um dos grandes problemas envolvendo a cirurgia cardiovascular (KAPOOR e cols., 2004) e a presença de endotoxemia piora a resposta clínica dos pacientes, principalmente pela indução de disfunção respiratória (LI e cols., 2005; MADDEN e cols., 2007). Como a SRIS tem fisiologia multifatorial, uma única intervenção não seria capaz de minimizá-la. O uso do mini- circuito de circulação extracorpórea é uma solução parcial para o problema em questão. Porém, como os resultados continuam contraditórios em demonstrar diferenças na evolução clínica de pacientes, se comparadas cirurgias sob CEC e mini-CEC (REMADI e cols., 2004; PERTHEL e cols., 2005) ou mesmo entre grupos sem uso de CEC (SHROYER e cols., 2009), há motivação para se buscar outros meios para redução da SRIS relacionada a cirurgias cardíacas.

1.2 Resposta Inflamatória em Cirurgia Cardíaca

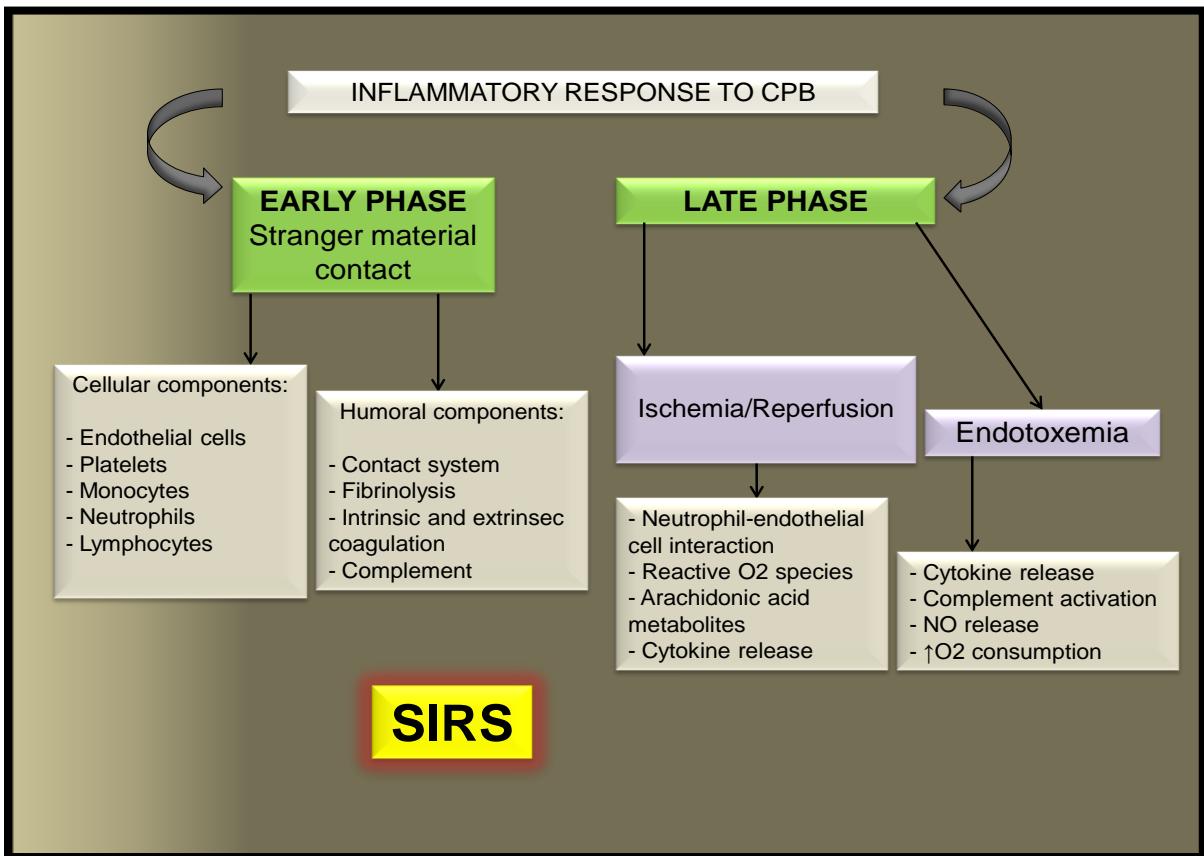
Maqsood e colaboradores (MAQSOOD e cols., 2008), fizeram ampla revisão sobre a resposta inflamatória relacionada à cirurgia cardíaca, com suas alterações humorais e celulares, e as possibilidades de interferência sobre as mesmas. Além da exposição do sangue a estruturas não fisiológicas do circuito externo de circulação, o trauma cirúrgico, a anestesia, a hipotermia com posterior reaquecimento, a liberação de endotoxinas intestinais e a lesão por isquemia/reperfusão (I/R) são importantes mediadores destas alterações (Elahi e cols., 2006) (Esquema 1). A isquemia/reperfusão, principalmente, leva a uma complexa reação imunológica, intermediada pelos leucócitos, com a liberação de EROs, metabólitos do ácido araquidônico, fatores plaquetários ativados, endotelinas, citocinas pró-inflamatórias e moléculas de adesão plaquetária e endotelial (MATATA e cols., 2000; MATATA e GALINANES, 2000) (Esquema 2).

Manifestações clínicas, tais como internações prolongadas, maior incidência de infecção na ferida operatória, falência respiratória, disfunção miocárdica, alterações metabólicas hepáticas, coagulopatias, insuficiência renal, distúrbios neurológicos e maior mortalidade podem decorrer de tais alterações (PLOMONDON e cols., 2001).



Esquema 2 - Complexa cascata inflamatória relacionada à isquemia/reperfusão. O metabolismo anaeróbico leva a um aumento do lactato e redução do pH com falência das bombas trans-membrana, levando a um acúmulo de Ca^{2+} e Na^+ , gerando edema celular. O aumento intracelular de Ca^{2+} ativa a fosfolipase A2 e a calpaína, levando à degradação do ácido araquidônico e inibição da síntese proteica, com ativação dos neutrófilos e ativação das caspases, gerando apoptose celular. A ativação dos neutrófilos leva a lesões de membrana e liberação de mediadores inflamatórios, inclusive óxido nítrico pela ativação da óxido nítrico sintase (NOS), com alteração da microcirculação lesão endotelial, formando-se um círculo vicioso inflamatório.

Franke, Warren e colaboradores (FRANKE e cols., 2002; WARREN e cols., 2009) demonstraram que a resposta inflamatória pós-CEC ocorre em duas fases (Esquema 3).



Esquema 3 - A resposta inflamatória decorrente da circulação extra corpórea (CPB-Cardiopulmonary Bypass) está dividida em duas fases: fase precoce (early phase) e fase tardia (late phase). A primeira fase é induzida pelo contato com material estranho do circuito extracorpóreo, enquanto a fase tardia se deve mais à reperfusão de oxigênio após a isquemia e à endotoxemia. SIRS: systemic inflammatory response syndrome; CPB: cardiopulmonary bypass.

A fase precoce decorre da exposição do sangue a superfícies não endoteliais do circuito de CEC, favorecendo a formação de coágulos, pelo desequilíbrio entre substâncias pró-coagulantes e anti-coagulantes, normalmente produzidas pelo tecido endotelial. As proteínas plasmáticas aderidas no circuito sofrem alterações conformacionais, levando à ativação de outros grupos de proteínas plasmáticas e grupos celulares que, numa complexa interação, iniciam uma resposta inflamatória difusa. Conforme a CEC avança, diminui esta resposta humoral e celular inicial, possivelmente pela aderência das proteínas plasmáticas ao circuito interno, tornando-o mais biocompatível. Começa a ser verificada então, a segunda fase da resposta inflamatória, ou fase tardia, relacionada à isquemia/reperfusão dos órgãos e tecidos e à endotoxemia (Esquema 2, Esquema 3).

O clampeamento aórtico realizado durante a CEC retira o aporte de sangue ao coração e aos pulmões (estes recebendo ainda algum suprimento das artérias brônquicas). A liberação do clampeamento, reperfunde amplamente estas áreas anteriormente isquêmicas, gerando uma

resposta inflamatória com alteração da perfusão capilar, acúmulo de fluido intersticial, leucocitose e coagulopatia. A ativação dos neutrófilos leva a uma extensa lesão endotelial. Em reação independente da ativação leucocitária, ocorre também a produção de EROs. Estas levam à liberação de metabólitos do ácido araquidônico, liberação de TNF- α e citocinas pelas células isquêmicas e ativação do sistema do complemento e da coagulação. A endotoxemia que ocorre pela liberação de toxinas (lipopolissacarídeos da parede celular das bactérias gram-negativas intestinais) na corrente circulatória parece ser um dos maiores estímulos para o desenvolvimento da SRIS (OPAL e cols., 2007).

Em nível celular, o metabolismo anaeróbico durante a isquemia leva a um aumento do lactato e fosfato inorgânico e redução do pH, que por sua vez leva à falência das bombas transmembrana, com acúmulo de sódio e cálcio, gerando edema intracelular (Esquema 2). O cálcio intracelular ativa a fosfolipase A2 e a calpaína entre outras proteases, e a falência das bombas de hidrogênio lisossomais e a queda do pH ativam enzimas lisossômicas que lesam as organelas celulares (De MOURA e cols., 2001).

A fosfolipase A2 degrada o ácido araquidônico, originando mediadores da inflamação, como leucotrienos, prostaglandinas e tromboxanos, substâncias que levam à adesão e ativação neutrofílica, vasoconstrição, lesão tecidual, agregação plaquetária e quimiotaxia na área isquêmica (De MOURA e cols., 2001). As EROs e os produtos da reação inflamatória, em um círculo vicioso, atraem e ativam os leucócitos, os quais liberam várias enzimas proteolíticas, como elastases, hidrolases, mieloperoxidases e proteases, causando destruição tecidual e amplificando a resposta inflamatória e a quimiotaxia (JUNQUEIRA e CARNEIRO, 2004; FRANCISCHETTI e cols., 2010).

Além dos leucócitos, as células endoteliais também são ativadas pelas substâncias inflamatórias circulantes. Inicia-se, então, a fase de rolamento dos leucócitos (principalmente os neutrófilos) (Figura 3) que por meio da exposição das L-selectinas (seus receptores) e a interação destas com os receptores P-selectinas (glicoproteínas intracelulares) das células endoteliais ativadas, desenvolvem uma fase de adesão frouxa ao endotélio vascular (MITCHELL e BEVILACQUA, 2006). Essa ligação é normalmente induzida pelo TNF- α e IL-1.

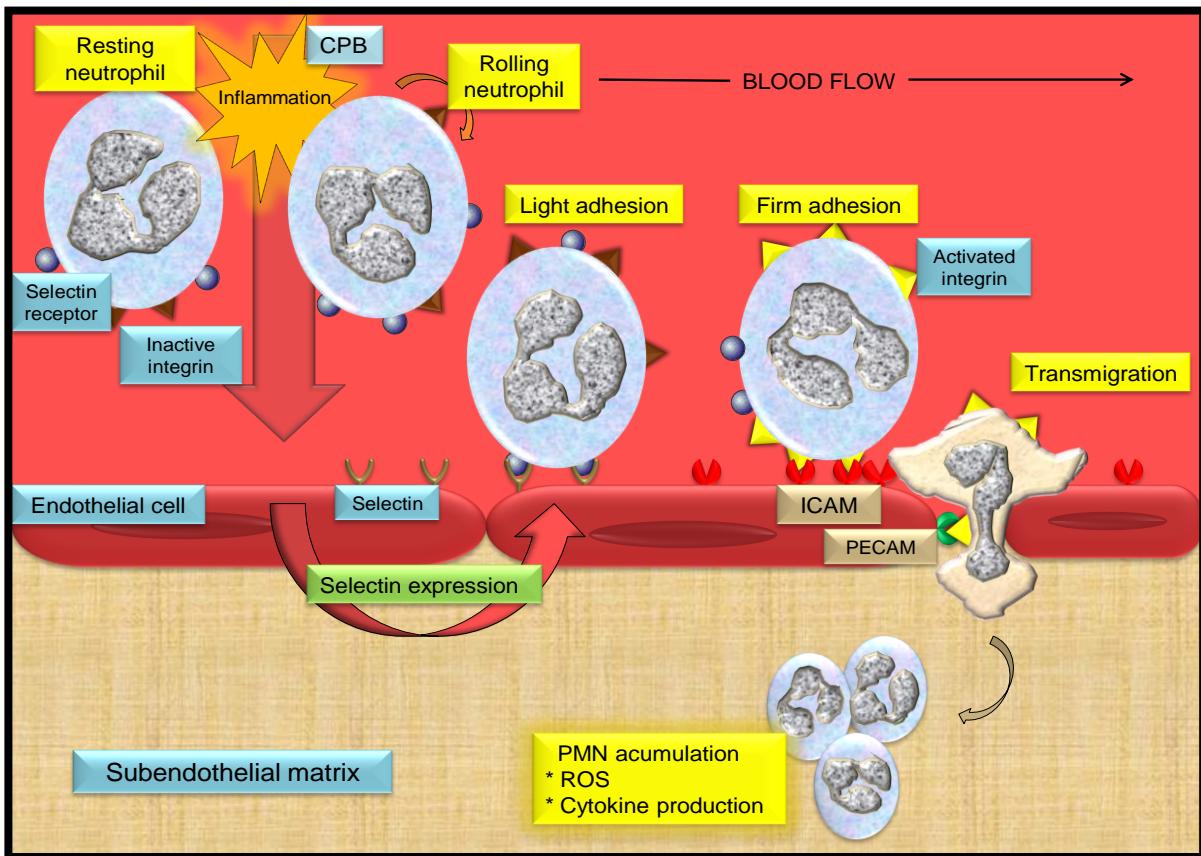


Figura 3 - Fase de estímulo dos neutrófilos e células endoteliais pelos fatores inflamatórios circulantes. Ocorre o rolamamento dos neutrófilos, adesão ao endotélio e posterior transmigração aos sítios inflamatórios, gerando acúmulo de polimorfonucleares e lesão tecidual. CPB: Cardiopulmonary Bypass; PMN: polymorphonuclears; ROS: Reactive Oxygen Species; ICAM: Intercellular adhesion molecule PECAM: Platelet/endothelial cell adhesion molecule.

A adesão intensa ocorre logo a seguir, por meio do contato das integrinas leucocitárias (complexos CD11/CD18) com as imunoglobulinas endoteliais vasculares (intercellular adhesion molecule- ICAM) (MITCHELL e BEVILACQUA, 2006), com migração dos leucócitos por diapedese aos sítios de inflamação. Estes são guiados por vários fatores quimiotáticos (MITCHELL e BEVILACQUA, 2006) (as aminas vasoativas, como a histamina e a serotonina, os metabólitos do ácido araquidônico (prostaglandinas, leucotrienos), as proteínas plasmáticas (sistemas do complemento, das cininas e da coagulação), o fator de ativação plaquetária (PAF), as citocinas (TNF- α e IL-1), o óxido nítrico (NO), os componentes lisossômicos dos leucócitos e as EROs) (FRANCISCHETTI e cols., 2010). A IL-8 parece ter papel importante na indução da migração dos neutrófilos pela parede endotelial (HUBER e cols., 1991). Há evidências de que a aderência induzida pelo CD11/CD18 induz à degranulação dos neutrófilos (RICHTER e cols., 1990), gerando EROs e

enzimas proteolíticas, que resultam em lesão importante às células endoteliais e consequentemente aos tecidos (SHAPPELL e cols., 1990).

A histamina e a serotonina, encontrados nos mastócitos, nos basófilos e nas plaquetas, são os primeiros mediadores liberados durante a inflamação. A imunoglobulina E (IgE), fragmentos do complemento C3a e C5a, citocinas e fatores liberadores de histamina derivados de leucócitos participam da liberação nos mastócitos. A liberação de histamina das plaquetas é estimulada após contato com colágeno, trombina, difosfato de adenosina (ADP), complexos antígeno- anticorpo e o fator ativador plaquetário (PAF). Os fragmentos C3a e C5a são importantes iniciadores da ativação neutrofílica e da produção de IL-8 (De MOURA e cols., 2001) nas lesões de isquemia/reperfusão (WALSH e cols., 2005). Desde estudo inicial de Weisman e colegas (WEISMAN e cols., 1990), que demonstrou depósitos de complemento em miocárdio reperfundido, outros estudos também comprovaram sua participação nos mecanismos de isquemia/reperfusão em outros órgãos (PEMBERTON e cols., 1993; WADA e cols., 2001; ZHAO e cols., 2002). Uma vez ativado, o complemento libera potentes substâncias inflamatórias, incluindo anafilatoxinas e complexos citolíticos, interagindo com as EROs (STAHL e cols., 2003).

As citocinas pró-inflamatórias são produzidas por linfócitos e macrófagos, mas também por células endoteliais e as principais são o TNF- α e as interleucinas (IL-1, IL-6 e IL-8). As citocinas levam à exposição de cargas negativas na superfície das células endoteliais (fase de rolamento e adesão), ativando a pré-calicreína que, então, converte-se em calicreína e ativa o fator XII, o qual, ativado, ativa os neutrófilos, levando à destruição da arquitetura endotelial vascular por meio da liberação de enzimas proteolíticas (FRANCISCHETTI e cols., 2010). Ocorre aumento progressivo de elastase sérica, de IL-6, de IL-8 e de PCR (CHRISTEN e cols., 2005; ELAHI e MATATA, 2006), tornando-se um mecanismo mantenedor da liberação de substâncias inflamatórias.

O óxido nítrico (NO) é outro mediador da resposta inflamatória que modifica o tônus e a permeabilidade vascular além de ser um agente quimiotático. É produzido pela enzima indutora óxido nítrico sintetase (iNOS) do endotélio, ativada pelo aumento do cálcio intracelular ou pelos macrófagos após indução por determinadas citocinas, como o INF- γ . Pode-se combinar com as EROs levando à formação de metabólitos como o peroxinitrito (OONO-), altamente reativo (CERQUEIRA e YOSHIDA, 2002). Estudos envolvendo a reação inflamatória pós-CEC, sugeriram que a iNOS esteja relacionada à indução de citocinas pró-inflamatórias, tais como o TNF- α (MATATA e GALINANES, 2002; ELAHI e cols.,

2006) e o NO induzido na resposta inflamatória relaciona-se à disfunção miocárdica (SATO e cols., 1997; OYAMA e cols., 1998).

As EROs, além de mediadores químicos da resposta inflamatória, ativam diferentes vias, tanto de adaptação celular como de apoptose. Enquanto níveis baixos são controlados pelos mecanismos antioxidantes endógenos, níveis elevados podem lesar o DNA das células, assim como proteínas e lipídios, levando à apoptose (MITCHELL e BEVILACQUA, 2006). Nos tecidos pós-isquêmicos (na reperfusão), há o acúmulo da xantina oxidase que, em lugar da nicotinamida adenina dinucleotídeo fosfato (NADPH), utiliza o O₂ como acceptor final de elétrons. Na reação hipoxantina-xantina, os elétrons são transferidos para o O₂, gerando o radical superóxido (O₂ •-), o qual sofre dismutação em peróxido de hidrogênio (H₂O₂). A auto-oxidação de catecolaminas e a enzima NADPH-oxidase dos neutrófilos ativados (JOHNSON e cols., 1994) são outras vias na produção de EROs.

1.3 Métodos para reduzir a SRIS pós-CEC

Desde a observação de que a liberação de substâncias inflamatórias e EROs durante a CEC e isquemia/reperfusão leva a eventos indesejáveis como dano microvascular e disfunção em diversos órgãos (ELAHI e MATATA, 2006; GONENC e cols., 2006), pesquisadores buscam a comprovação de estratégias para proteção celular com resultados discrepantes. Além do uso dos mini- circuitos já descritos (TAKAY e cols., 2005), o uso de antioxidantes exógenos foi um dos mais estudados. Porém, como estes possuem uma ação predominantemente extracelular, são limitados na sua capacidade para a proteção celular (PRYOR, 1984). Estudos não comprovaram o benefício de antioxidantes exógenos em pacientes submetidos à CEC (ELAHI e cols., 2005; ELAHI e MATATA, 2006; GONENC e cols., 2006) mesmo que estes possam reduzir de alguma maneira a formação de EROs (KAWAHITO e cols., 2000).

A administração de corticosteróides durante a CEC também foi extensamente investigada, pois podem reduzir a liberação de complemento (SANO e cols., 2003; LIAKOPoulos e cols., 2007). Apesar dos resultados positivos, em pacientes submetidos à CEC (WAN e cols., 1999; CHANEY e cols., 2001; ASIMAKOPoulos e cols., 2003; LEVY e TANAKA, 2003; RUBENS e cols., 2005), benefícios clínicos evidentes não foram confirmados e efeitos pouco desejáveis, como uma maior demora na extubação dos pacientes

(WAN e cols., 1999; CHANEY e cols., 2001; ASIMAKOPOULOS e cols., 2003; LEVY e TANAKA, 2003; RUBENS e cols., 2005) em doses expressivas.

A aprotinina, um polipeptídeo extraído de pulmão bovino, também pode limitar a fibrinólise associada à exposição sanguínea a superfícies estranhas, pois tem efeito anti-inflamatório pela redução da produção de NO (BRUDA e cols., 1998), podendo aumentar a liberação de IL-10 (HILL e cols., 1998). Clinicamente, a aprotinina reduziu a necessidade de uso de transfusão homóloga e o risco de Acidente Vascular Cerebral (AVC), se comparada ao placebo (SEDRAKYAN e cols., 2004; LEVI e cols., 1999), porém, estudos retrospectivos correlacionaram o seu uso a uma maior morbi-mortalidade no pós-operatório dos pacientes submetidos à cirurgia cardíaca (VAN DER LINDEN e cols., 2007).

O revestimento dos circuitos da CEC com heparina promove uma maior biocompatibilidade dos mesmos, por reduzir a ativação do complemento, pela inibição da ativação de granulócitos ou adesão plaquetária e por diminuir a liberação de TNF- α e IL-8 (KUTAY e cols., 2006; JESSEN, 2006), protegendo os pacientes de alterações cognitivas e disfunções renais durante a CEC (HEYER e cols., 2002; de VROEGE e cols., 2005). Em estudo prospectivo e randomizado, Goudeau e colaboradores (GOUDEAU e cols., 2007) demonstraram concentrações significativamente menores de PCR, IL-6, CPK-MB, troponina I, ácido lático e EROs em plasma de pacientes tratados com heparina, com redução de complicações pós-operatórias e menor tempo de permanência em unidade de tratamento intensivo (UTI). A heparina tem sido usada de rotina em procedimentos que envolvem a CEC.

Como são gerados vários mediadores inflamatórios durante a CEC, sugeriu-se que o uso de filtros para reter leucócitos e outros componentes celulares ativados também poderia reduzir a lesão pulmonar pós-operatória induzida pela CEC (WARREN e cols., 2007). Contudo, a remoção simultânea das plaquetas influencia negativamente a hemostasia trans-operatória (WARREN e cols., 2007). O uso de oxigenadores de membrana em lugar do oxigenador de borbulhas parece reduzir o trauma sanguíneo e a embolia gasosa (LAUTH e cols., 1990). As superfícies hidrofílicas, resistentes ao depósito de proteínas e células e o óxido de polietileno, pela sua configuração molecular favorável, têm sido estudados (TAN e cols., 2007) para uso na construção dos circuitos de CEC.

Como podemos observar, o mecanismo envolvido na resposta do sangue ao circuito de CEC é complexo e está longe de ser completamente elucidado. O estresse oxidativo e a inflamação, intimamente relacionados, correlacionam-se com uma pior evolução clínica dos pacientes submetidos à CRM o que leva à tentativa de adoção de estratégias para o equilíbrio destas respostas.

1.4 Imunomodulação trans-operatória pelos anestésicos

A imunomodulação trans-operatória pelos agentes anestésicos pode ser exemplificada por alguns estudos (CROZIER e cols., 1994; KEVIN e cols., 2005), a maioria deles sugerindo que os anestésicos possuem efeitos anti-inflamatórios, mas relacionados à imunossupressão e maior suscetibilidade dos pacientes a infecções (KRUMHOLZ e cols., 1994; HELLER e cols., 2008), exceção provavelmente feita aos agonistas (α)-2-adrenérgicos (Pandharipande e cols., 2007; PANDHARIPANDE e cols., 2008).

O uso do anestésico propofol (2-6 di-iso-propilfenol), cuja estrutura é semelhante aos anti-oxidantes fenólicos (KAHRAMAN e DEMIRYUREK, 1997), em modelos animais experimentais, é efetivo na proteção a vários órgãos (YOUNG e cols., 1997; NAVAPURKAR e cols., 1998). O propofol é bastante utilizado na indução e manutenção da anestesia geral, principalmente nas anestesias intravenosas totais (AIVT). Em concentração anestésica habitual, leva à redução da peroxidação lipídica (MURPHY e cols., 1992;) com efeito protetor miocárdico (CORCORAN e cols., 2006; XIA e cols., 2006). Porém, dados *in vivo* e *in vitro* (KRUMHOLZ e cols., 1994; HELLER e cols., 1998; KOTANI e cols., 1999) sugerem que os efeitos anti-inflamatórios do propofol podem, em caso de infecções associadas, levar a uma piora do quadro infeccioso.

Os opióides, outra classe de medicamentos que são importantes analgésicos utilizados em anestesia, tiveram seu efeito imunossupressor primeiramente demonstrado em 1898 (CANTACUZENE, 1898). A morfina foi a droga mais estudada, sendo que pouco se sabe sobre o efeito imunossupressor de outros opióides. Foi demonstrado que a morfina tem efeito anti-inflamatório *in vitro* e que aumenta a mortalidade em modelos de infecção em animais (WEINERT e cols., 2008). Interessante foi a observação de que a clonidina, um agonista (α)-2-adrenérgico, de maneira dose-dependente, em ratos, foi eficiente em modular a supressão imunológica causada pela morfina (WEST e cols., 1999). Para uma revisão dos efeitos imunomodulatórios dos sedativos/ anestésicos em geral, o trabalho de Sanders e colegas (SANDERS e cols., 2011) constitui-se ótima referência.

Os agonistas (α)-2-adrenérgicos podem exercer seus efeitos através de interações neuro-imunes, pela ação direta sobre o Sistema Nervoso Simpático (SNS), o que mostrou ter efeitos benéficos sobre o sistema imunológico (SMITH e cols., 1977; NANCE e SANDERS, 2007). De maneira controversa, o estímulo de receptores (α)-2-adrenérgicos parece provocar uma resposta pró- inflamatória *in vitro* (SPENGLER e cols., 1990) e *in vivo* (FLIERL e cols., 2007), porém, na maioria dos estudos em animais e humanos a sua administração induziu

resposta anti-inflamatória (NADER e cols., 2001; VENN e cols., 2001; Taniguchi e cols., 2004; MEMIS e cols., 2007; TANIGUCHI e cols., 2008). Esta dualidade parece ser decorrente dos diferentes efeitos, centrais e periféricos, destas drogas. Em nível periférico, os agonistas (α)-2-adrenérgicos parecem estimular a imunidade inata (MILES e cols., 1996; WEATHERBY e cols., 2003; GETS e MONROY, 2005) e a ação simpatolítica central induz a um aumento do tônus parassimpático que parece conseguir promover o controle do quadro infeccioso (STERNBERG, 2006; TRACEY, 2007). Em presença de inflamação, um agonista (α)-2-adrenérgico como a dexmedetomidina age de forma anti-inflamatória preferencialmente (SUD e cols., 2008).

A dexmedetomidina, um anestésico agonista (α)-2-adrenérgico, pode ser uma droga promissora como indutora de proteção celular, pela sua capacidade de ativação pré-sináptica de receptores (α)-2-adrenérgicos (Figura 4), atenuando a liberação excessiva de noradrenalina durante a isquemia (MATSUMOTO e cols., 1993) com diminuição do potencial para a formação de EROS (SCHOLZ e TONNER, 2000; ROCHA e cols., 2010).

A administração trans-operatória de dexmedetomidina tem demonstrado vantagens, como em reduzir as doses de outros anestésicos, em melhorar a estabilidade hemodinâmica e em manter a sedação dos pacientes no período de recuperação de cirurgias de grande porte (JALONEN e cols., 1997; HERR e cols., 2003; AANTAA e JALONEN, 2006) (Figura 5). Kang e colegas (KANG e cols., 2012), demonstraram que a dexmedetomidina (dose de 1 $\mu\text{g} \cdot \text{kg}^{-1}$ em bolus e infusão posterior de 0.6 $\mu\text{g} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$) reduziu a necessidade de propofol durante a anestesia venosa com o opióide remifentanil sem comprometer o padrão de recuperação, mantendo os pacientes hemodinamicamente mais estáveis. O efeito sedativo da dexmedetomidina é mediado no tronco cerebral, pelo *locus ceruleus*, onde a dexmedetomidina diminui o tônus simpático e aumenta o tônus parassimpático (ARCANGELI e cols., 2009) (Figura 5).

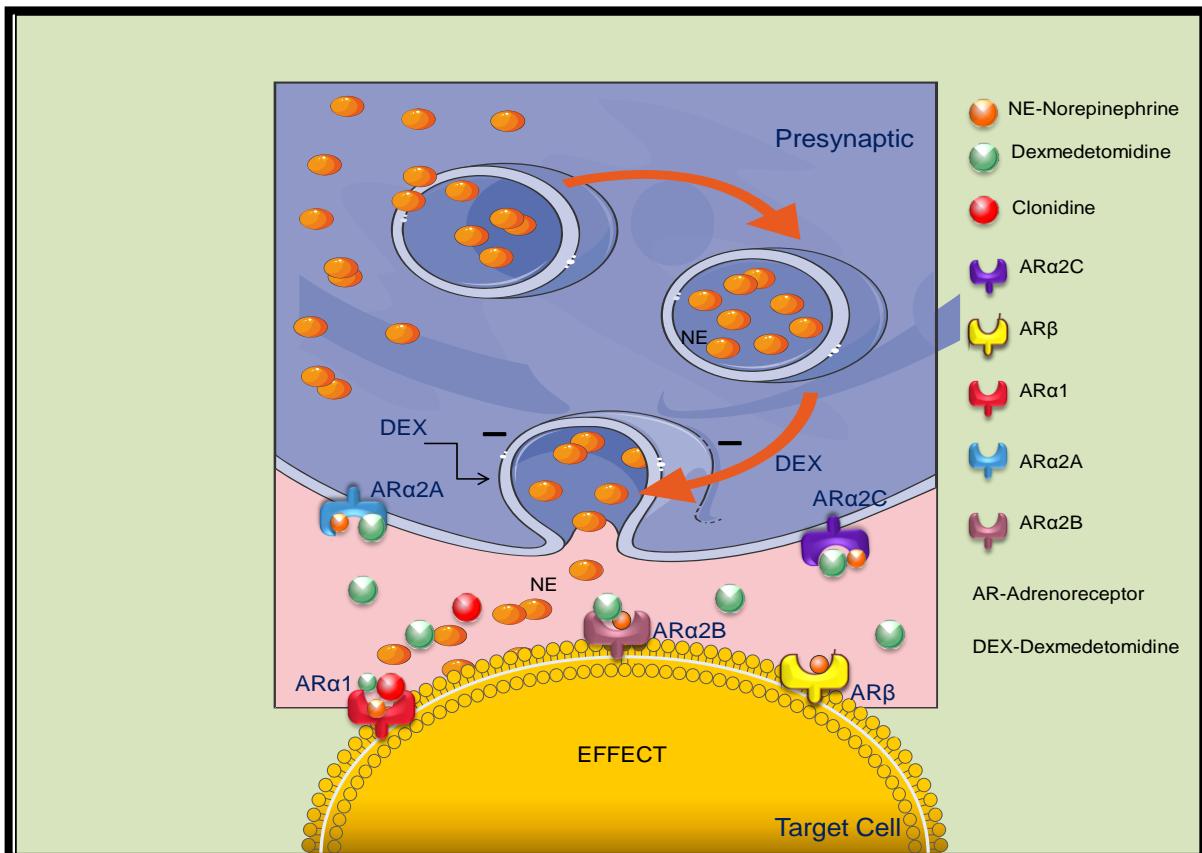


Figura 4 - Efeito da dexmedetomidina sobre os receptores (α)-2-adrenérgicos pré e pós-sinápticos. A dexmedetomidina pode exercer seus efeitos sobre três subtipos de (α)-2-adrenorreceptores. As subclasses pré-sinápticas de (α)-2-adrenorreceptores ((α)-2A e (α)-2C), estimulados pela dexmedetomidina, inibem a liberação de norepinefrina, por feed-back negativo. Localizados pós-sinapticamente estão a subclasse (α)-2B-adrenorreceptores e também (α)-1-adrenorreceptores, pois a dexmedetomidina não é (α)-2-adrenérgica específica. Os (α)-2-adrenorreceptores também existem extrasinapticamente.

Os mecanismos diferentes para produzir sedação entre a dexmedetomidina, o propofol e os opióides sugerem um sinergismo benéfico em sua combinação. O efeito poupadão de propofol que a dexmedetomidina apresenta pode ser atraente, por reduzir as doses necessárias do propofol, evitando os seus principais efeitos adversos, tais como a acidose metabólica, a depressão miocárdica, a alteração da agregação plaquetária e a demorada recuperação, relacionados ao seu uso em grandes doses ou por período prolongado (BOLLI e cols., 1988; De LA CRUZ e cols., 1997; AOKI e cols., 1998; BUROW e cols., 2004; SALENGROS e cols., 2004; LIOLIOS e cols., 2005; MERZ e cols., 2006; KAM e CARDONI, 2007).

Autores relataram um efeito anti-inflamatório da dexmedetomidina superior aos dos outros anestésicos (NADER e cols., 2001; VENN e cols., 2001; MEMIS e cols., 2007), atividade anti-apoptótica (MaD e cols., 2004) e capacidade de melhor modulação da função dos macrófagos (MILES e cols., 1996; WEATHERBY e cols., 2003; GETS e cols., 2005;

YANG e cols., 2008). Hofer e colegas (HOFER e cols., 2009) em modelo animal válido para sépsis em humanos, demonstraram que a administração prévia de clonidina e dexmedetomidina aumentou significativamente a sobrevida. Observou-se uma redução dos mediadores pró-inflamatórios IL-1 β , IL-6 e TNF- α , inibindo a reação inflamatória à sépsis. Estes resultados suportam a idéia de se considerar o uso de drogas simpatolíticas como adjuvantes importantes em pacientes que serão submetidos a cirurgias de grande porte e/ou em pacientes em unidade de tratamento intensivo (UTI) sujeitos a desenvolver um quadro séptico. Pandharipande e colegas (PANDHARIPANDE e cols., 2007) descreveram o aumento da sobrevida em pacientes críticos sedados em UTI, pelo efeito simpatolítico e vagomimético da dexmedetomidina. Spies e colegas (SPIES e cols., 1996) também demonstraram que pacientes tratados com clonidina para crises de abstinência ao álcool tiveram menor incidência de pneumonia se comparados àqueles tratados com agentes apenas simpatolíticos.

Apesar de que resultados de estudos em humanos continuem contraditórios (WIJEYSUNDERA e cols., 2003; SULEMANJI e cols., 2007; TASDOGAN e cols., 2009), em modelo animal, foi demonstrado também o efeito neuroprotetor da dexmedetomidina (MAIER e cols., 1993) após isquemia cerebral induzida e reperfusão. Estudos bem recentes sugerem que a dexmedetomidina possa ser o anestésico de eleição para pacientes sob o risco de desenvolver lesões neurológicas no período trans-operatório (CHEN e cols., 2013; PENG e cols., 2013).

Os fatores hematológicos também são sensíveis às mudanças metabólicas sendo que a capacidade de deformidade dos eritrócitos e a viscosidade plasmática podem afetar de maneira importante a perfusão dos órgãos e tecidos (SIMCHON e cols., 1987; ZINCHUK, 2001). A ocorrência de estresse oxidativo leva à peroxidação lipídica, com consequente comprometimento das funções e da integridade das membranas dos eritrócitos (KUYPERS, 1998; THEROND e cols., 2000; SIVIOTTI, 2004). Arslan e colegas (ARSLAN e cols., 2012) demonstraram a capacidade de proteção da deformidade dos eritrócitos pelo uso prévio de dexmedetomidina em ratos submetidos à isquemia/reperfusão hepática. O estudo mostrou que ocorre claramente uma alteração importante na capacidade para a deformidade dos eritrócitos no modelo experimental de isquemia/reperfusão hepática em ratos, relacionada à peroxidação lipídica, e que houve proteção pelo uso da dexmedetomidina.

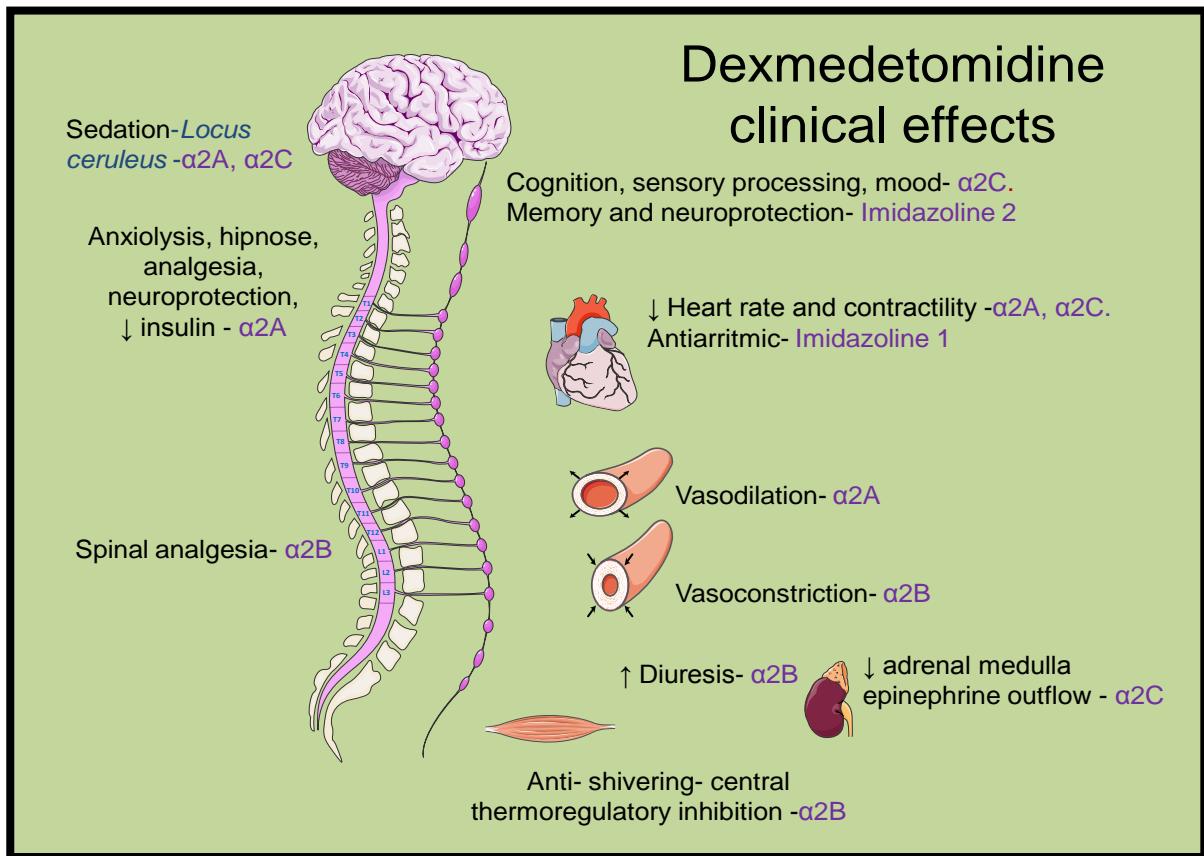


Figura 5 - Efeitos clínicos induzidos pelo uso da dexmedetomidina e os receptores específicos envolvidos. Através do agonismo a adrenorreceptores pré-sinápticos α_2A , a dexmedetomidina induz sedação, ansiolise, hipnose, analgesia, promove a neuroproteção, reduz a liberação de insulina, reduz a frequência cardíaca, reduz a contratilidade miocárdica e leva à vasodilatação. Pelo efeito pré-sináptico sobre os adrenorreceptores α_2C , induz sedação, modulação da cognição, memória e processamento sensório e reduz a liberação de epinefrina na medula adrenal. Pelo agonismo a adrenorreceptores α_2B pós-sinápticos, a dexmedetomidina leva à analgesia em nível espinhal, vasoconstrição (efeito pós-sináptico, com doses elevadas em *bolus*), aumento da diurese e inibição central dos tremores. A dexmedetomidina também atua sobre os receptores imidazolínicos, levando à neuroproteção (receptores imidazolínicos 2) e tendo efeito antiarritmico (receptores imidazolínicos 1).

1.5 Justificativa

A taxa de mortalidade dos pacientes submetidos à cirurgia de revascularização miocárdica (CRM), no Brasil é bastante elevada (6,2%) (PIEGAS e cols., 2009) se comparada a outros países como os Estados Unidos (2.9%) (HANNAN e cols., 2003) e Canadá (1.7%) (CARTIER e cols., 2008). Diversos estudos têm demonstrado a ocorrência da síndrome da resposta inflamatória sistêmica (SRIS) associada à CRM com circulação extracorpórea (CEC) e suas implicações clínicas indesejadas. Em nosso estudo, propomos investigar o efeito da dexmedetomidina, um anestésico agonista (α)-2-adrenérgico, sobre esta resposta inflamatória. Considerando os resultados positivos obtidos em estudos anteriores quanto à potencialidade da dexmedetomidina, acreditamos que ela possa ser capaz de modificar a resposta inflamatória ao trauma e a evolução clínica dos pacientes. Assim, pretendemos avaliar os níveis plasmáticos de citocinas pró e anti- inflamatórias e outros biomarcadores inflamatórios, além de avaliar a peroxidação lipídica pela dosagem das substâncias reativas ao ácido tiobarbitúrico (TBARS) (PUNTEL e cols., 2007) e o estresse oxidativo pela atividade da enzima delta aminolevulinato desidratase (δ -ALA-D) (BERLIN e SCHALLER, 1974).

1.6 Objetivos

1.6.1 Objetivo Geral

O presente estudo teve como objetivo geral investigar o efeito do uso da dexmedetomidina, uma droga agonista (α)-2-adrenérgica sobre os biomarcadores de inflamação e estresse oxidativo em pacientes submetidos à cirurgia de revascularização miocárdica com mini-circulação extracorpórea.

1.6.2 Objetivos Específicos

Manuscrito 1

1. Determinar o novo estado da arte da dexmedetomidina e suas potencialidades;
2. Discutir a capacidade da dexmedetomidina na proteção celular e de órgãos, visando promover o seu uso, ainda muito incipiente durante os procedimentos cirúrgicos.

Manuscrito 2

1. Comparar dois grupos de pacientes submetidos à cirurgia de revascularização miocárdica com mini-circulação extracorpórea, usando técnicas diferentes de anestesias venosas totais, uma delas com o uso associado da dexmedetomidina;
2. Avaliar o efeito da dexmedetomidina nos níveis dos marcadores inflamatórios, como de IL-1, IL-6, TNF- α , INF- γ e PCR e no nível do marcador antiinflamatório IL-10;
3. Estudar o efeito do uso da dexmedetomidina sobre as respostas ao estresse oxidativo, através da avaliação de TBARS e da atividade da enzima δ -ALA-D;
4. Determinar os níveis dos marcadores de lesão celular miocárdica, como CPK, CPK-MB e cTnI;
5. Avaliar o efeito da dexmedetomidina nos níveis de outros biomarcadores bioquímicos.

2 MANUSCRITOS

Os resultados que fazem parte desta tese estão apresentados sob a forma de manuscritos científicos, os quais se encontram aqui organizados. Uma referência ao estado da arte em relação resposta inflamatória em pacientes submetidos à CRM e potencialidades da dexmedetomidina sobre estas alterações encontram-se no **Manuscrito 1**. Os itens Materiais e Métodos, Resultados e Discussão dos Resultados encontram-se no **Manuscrito 2**. Os **Manuscrito 1** e **Manuscrito 2** estão dispostos na forma em que normalmente se submete para publicação.

- 2.1 MANUSCRITO 1 – Resposta inflamatória em pacientes submetidos à cirurgia de revascularização miocárdica (CRM) e implicações clínicas: uma revisão da relevância do uso de dexmedetomidina.**

Manuscrito 1

INFLAMMATORY RESPONSE IN PATIENTS UNDER CORONARY ARTERY BYPASS GRAFTING SURGERY (CABG) AND CLINICAL IMPLICATIONS: A REVIEW OF THE RELEVANCE OF DEXMEDETOMIDINE USE

NEUSA MARIA HEINZMANN BULOW, ELISÂNGELA COLPO,
EDUARDO FRANCISCO MAFASSIOLY CORREA, ROCHELLE SILVEIRA
SCHLOSSER, ANELISE LAUDA, IGE JOSEPH KADE, JOÃO BATISTA
TEIXEIRA ROCHA

Manuscrito 1

INFLAMMATORY RESPONSE IN PATIENTS UNDER CORONARY ARTERY BYPASS GRAFTING SURGERY (CABG) AND CLINICAL IMPLICATIONS: A REVIEW OF THE RELEVANCE OF DEXMEDETOMIDINE USE.

Neusa Maria Heinzmann Bulow¹, Elisângela Colpo², Eduardo Francisco Mafassiyol Correa^{3,b}, Rochelle Silveira Schlosser^{4,b}, Anelise Lauda^{5,b}, Ige Joseph Kade^{6,a}, João Batista Teixeira Rocha^{7,a}

Departamento de Química, Programa de Pós-graduação em Ciências Biológicas: Bioquímica Toxicológica, Centro de Ciências Naturais e Exatas, Universidade Federal de Santa Maria, Cep 97105-900, Santa Maria, RS, Brazil.

b Departamento de Cirurgia, Centro de Ciências da Saúde, Universidade Federal de Santa Maria, Cep 97105-900, Santa Maria, RS, Brazil.

Corresponding author:

Neusa Maria Heinzmann Bulow and João Batista Teixeira da Rocha

UFSM – CCNE – Dep. de Química

Cep 97105-900, Santa Maria, RS, Brasil.

Tel: #55-55-3220-8140

Fax: #55-55-3220-8978

2.1.1 Summary

Despite coronary artery bypass grafting surgery (CABG) with cardiopulmonary bypass (CPB) prolongs life and reduces symptoms in patients with severe coronary artery diseases, these benefits are accompanied by increased risks. Morbidity associated with cardiopulmonary bypass can be attributed to the generalized inflammatory response induced by blood-xenosurfaces interactions during extracorporeal circulation and the ischemia/reperfusion implications, including exacerbated inflammatory response resembling the systemic inflammatory response syndrome (SIRS). The use of specific anesthetic drugs that could, as antiinflammatory agents, these inflammatory response modulate and promote a postoperative recovery may be advantageous. It is known that the stress response to surgery can be attenuated by sympatholytic effects caused by activation of central (α)-2-adrenergic receptor, leading to reductions in blood pressure and heart rate, and more recently, that they can have antiinflammatory properties. This review discusses the clinical significance of the dexmedetomidine use, a selective (α)-2-adrenergic agonist, as a coadjuvant in general anesthesia. Actually, dexmedetomidine use is not in anesthetic routine, but this drug can be considered a particularly promising agent in perioperative multiple organ protection.

Keywords: Dexmedetomidine; Inflammation; Oxidative Stress; Total Intravenous Anesthesia (TIVA); Cardiopulmonary Bypass (CPB).

2.1.2 Introduction

2.1.2.1 Inflammatory response and ischemia/reperfusion in CABG surgery

Surgery induces a variety of metabolic, endocrine, and immune changes known as the "stress response", which may lead to prolonged in-hospital stay. The clinical manifestation of this reaction include postoperative complications such as respiratory failure, wound infections (1) myocardial damage with contractile dysfunction, renal impairment, coagulopathy, neurologic dysfunction (2) and altered liver function with an increased mortality (3).

Inflammatory response in cardiac surgical patients is produced by complex interactions with numerous pathways including generation or activation of complement, cytokines, neutrophils, thrombin, mast cells, and others multiple inflammatory mediators. Cardiopulmonary bypass responses has often been compared with the pathophysiologic

changes occurring in systemic inflammatory response syndrome (SIRS) (4) and remain not fully understood. Several interlinked mechanisms could play a role in the pathological effects associated with cardiopulmonary bypass, for instance, the exposure of blood to nonphysiologic surfaces, surgical trauma, anesthesia, changes in body temperature, increased intestinal permeability to endotoxins, and ischemia-reperfusion injury (5). It results in a complex immunologic reaction with the release into circulation of arachidonic acid metabolites, proinflammatory cytokines, endothelins, platelet-activating factors, endothelial and leukocyte adhesion molecules that stimulate the overproduction of reactive oxygen species (6, 7) (Scheme 1).

SCHEME 1 ABOUT HERE

Although it has been shown that, compared with clinical management alone, conventional coronary artery bypass grafting surgery with cardiopulmonary bypass prolongs life and reduces symptoms in patients with severe coronary artery diseases, these benefits are accompanied by increased risks of transfusions (30–90%), mortality (2–6%), stroke (2%), atrial fibrillation (30%), and neurocognitive dysfunction (50–60%) (8, 9). The adverse clinical consequences, associated with conventional coronary artery bypass surgery, have been largely attributed to the extracorporeal blood circulation (ECC) on cardiopulmonary bypass circuit, general systemic effects (including exacerbated inflammatory response resembling the SIRS, Scheme 1), hypothermic cardiac arrest, aortic cannulation, and cross-clamping (10,11,12). Consequently, it may be of interest to study the potential benefit of specific anesthetic drugs exhibiting anti-inflammatory mechanism. By modulating inflammatory response, anesthetic drugs could reduce the postoperative complications and mortality associated with CABG.

One potential candidate that has been little explored is dexmedetomidine. Dexmedetomidine, an (α)-2-adrenergic receptor agonist, can provide anxiolysis and sedation without respiratory depression (13). It decreases central nervous system sympathetic outflow in a dose-dependent manner and has analgesic effects described as opioid-sparing effect. There is increasing evidence that dexmedetomidine has organ protective effects against ischemic and hypoxic injury, including cardioprotection, neuroprotection, and renoprotection (13). However, little is known about the cellular and molecular mechanism(s) involved in dexmedetomidine protective effects. Here we will discuss the potential systemic antioxidant and anti-inflammatory action of dexmedetomidine and its possible relationship with cardio- and neuroprotective effects after coronary artery bypass grafting surgery (CABG).

2.1.3 Cardiopulmonary bypass (CPB)

Cardiopulmonary bypass (CPB) replaces the functions of the heart and lungs during cardiac surgery, allowing the heart to be opened and operated (Figure 1). The first successful human intracardiac operation was performed by Gibbon in 1953, using a mechanical extracorporeal pump oxygenator (14). Despite the long time since the first CPB surgery and numerous studies about CPB pathophysiological side-effects, the complex mechanisms involved in the responses of blood and tissues to cardiopulmonary bypass are still far from clear.

Clinical points of evidence suggest that morbidity associated with cardiopulmonary bypass can in part be attributed to the generalized inflammatory response induced by blood-xenosurfaces (from catheters and filtration membranes) interactions during extracorporeal circulation (4) (Scheme 1, Figure 1). Although conflicting data exist, the prominent hypothesis is that a metabolic unbalance occurs during extracorporeal blood recirculation involving every line of the inflammatory response including complement activation. Total perioperative values of inflammatory markers are probably less important than the balance between the oxide inflammatory cascade and antiinflammatory feedback mechanisms. Oxidative stress and inflammation are related and perhaps inseparable and, a reduced cytokine response may be directly translated into changes in clinical outcomes (15, 16).

FIGURE 1 ABOUT HERE

The pump and the oxygenator used for cardiopulmonary bypass function in a nonphysiologic manner, with altered vascular pressure and gas composition. Inflammation is the initial, nonspecific response of vascularized tissue to a variety of injuries, involving both the activation of humoral and cellular inflammatory pathways. Significant hemodilution also occur leading to a dilution and denaturation of plasma proteins. The blood exposition to nonendothelial surfaces activates the production of vasoactive mediators, altering capillary permeability and causing hemolysis (which increase the free concentration of the prooxidant heme and non-heme iron) and the coagulation system will be impaired (17). One important question that needs to be explored in details is whether or not the ECC induced hemolysis increases the concentration of iron in pathologically relevant tissues such as brain, heart and kidney. Increase in free hemoglobin, heme and iron can further feed the pro-oxidative-pro-inflammatory cycle in different tissues (18-24). The potential role of iron on “early and late phases” of inflammation associated with cardiopulmonary bypass (see below) should be

investigated in detail as well as the possibility of utilizing chelation therapy as co-adjuvant in patients at risk of developing SIRS-like response. Of particular significant, literature data have indicate a beneficial effect of desferoxamine in sepsis (25-27), which indicate that buffering of free iron can reduce the toxicity found in SIRS or SIRS-like situations (25-27).

The inflammatory response to cardiopulmonary bypass can be divided into 2 phases: “early” and “late” (Scheme 2). The early phase occurs as a result of the direct blood contact with nonendothelial surfaces, and the late phase is triggered by ischemia-reperfusion injury and endotoxemia (For a comprehensive review see the work of Warren and colleagues) (17).

SCHEME 2 ABOUT HERE

In the early phase, thrombosis becomes favorable, and it can be reduced or ameliorated with the administration of heparin before cardiopulmonary bypass initiation. When heparinized blood comes into cardiopulmonary bypass circuit, plasma proteins are adsorbed onto the circuit, leading to the activation of plasma protein systems and cell groups. These initiate a whole-body inflammatory response, associated with tissue edema, coagulopathy and organ dysfunction (28). With the course of cardiopulmonary bypass, the activation of the humoral and cellular components diminishes, but a second phase of inflammatory response initiates, which is related to ischemia-reperfusion injury and release of endotoxins from intestinal microflora (29). The ischemia-reperfusion injury is mediated by neutrophil-endotelial interactions (Fig. 2). High levels of endothelial injury occur during ischemic period, resulting in neutrophil activation and sequestration on reperfusion. Independent of leukocytes, production of toxic reactive oxygen species also occur, leading to release of arachidonic acid metabolites, proinflammatory cytokines by ischemic cells (eg, plasma tumor necrosis factor-alpha and interleukins like IL-1, IL-6 and IL-8) and activation of the humoral protein systems (30). The reintroduction of oxygen during reperfusion promote a high concentration of damaging reactive oxygen species in previously ischemic cells, and can damage cell membranes, denature proteins and act as second messengers to stimulate an acute inflammatory response (4) (Scheme 2 and scheme 3).

SCHEME 3 ABOUT HERE

There are many possible sources of endotoxin, including lipopolysaccharides from cell wall of gram-negative bacteria, release during bypass, with gut translocation as the primary source (31). The increased level of endotoxin related to cardiopulmonary bypass stimulate the release of nitric oxide and proinflammatory cytokines and increase levels of oxygen consumption (32). These stimuli and the complexity of this disequilibrium, the balance between the processes of activation and inhibition of these systems, suggest that the implementation of effective antiinflammatory and antioxidant strategies (though to be desirable in theory) can be a difficult challenge. Of particular pharmacological significance, recent experimental data have indicated that dexmedetomidine can attenuate sepsis-induced lung and kidney damage, in part by decreasing tissue migration of inflammatory cells in rats (33). These results may indicate a potential role of dexmedetomidine as a negative modulator of SIRS-like response in cardiopulmonary bypass.

2.1.3.1 Mini-extracorporeal circulation (MECC)

Biocompatible circuits designed to prevent the early activation of inflammatory cascades have been shown to affect some aspects of blood activation but not all. There have been some progresses in cardiopulmonary bypass design that has shown promising clinical outcomes, particularly, those aiming to reduce the incidence of SIRS-like response and its complications. Recently, a new cardiopulmonary bypass system, the mini-extracorporeal circulation (MECC), has been developed and it use has been associated with a reduced inflammatory response, when compared with the conventional system (standard cardiopulmonary bypass or extracorporeal circulation). It has no venous reservoir, a reduced priming volume, and less blood-synthetic interface contact (Figure 3).

FIGURE 3 ABOUT HERE

In a review, Vohra and colleagues have consolidated the current literature on the mini-extracorporeal circulation system (34). They have paid particular attention to the role that cardiopulmonary bypass has in generating a systemic inflammatory response and have outlined ways in which MECC may be superior to standard cardiopulmonary bypass. The MECC system has shown promising results with regard to cardiac damage and end-organ dysfunction. Many studies cited by this author have also shown that changes in blood markers of inflammation (for instance, C-reactive protein, leucocytes, and cytokines) were lower when

MECC is used. Of clinical significance, utilization of MECC has been associated with a decrease in complications found more frequently in standard ECC, particularly arrhythmias and thromboembolic events.

2.1.3.2 Oxidative stress and inflammation associated with coronary artery bypass grafting surgery (CABG)

Reactive oxygen species are recognized as critical mediators of cardiac and neurologic injury during ischemia and reperfusion. Sources of these reactive oxygen species are the mitochondrial electron transport chain, the enzymes xanthine oxidase, NADPH oxidase, lipoxygenase/cyclooxygenase and nitric oxide synthase (NOS), and auto-oxidation of various substances, such as catecholamines. An unpaired electron usually makes the species highly reactive. There are endogenous antioxidant systems that counteract the potential for injury to cellular structures by regulating the balance of reactive oxygen species. These endogenous antioxidants are upregulated when exposure of the cell to the reactive oxygen species is increased. Under pathologic conditions, such as ischemia-reperfusion, their formation can rapidly overcome antioxidant defenses and cellular injury ensues. It is known that the cardiopulmonary bypass can be responsible for activating neutrophils that represents a prominent source of systemic primary reactive oxygen species production (Figure 2). The synergism of damages related to reactive oxygen species, activation and infiltration of neutrophils in reperfused tissues, has been well recognized for many years (4).

Some investigators suggested that strategies of neurological and myocardial protection must not be limited to interventions targeted at the heart or brain itself, but should take in account the systemic response of organism to cardiopulmonary bypass (35, 36). These concepts should be particularly relevant for high-risk patients, who are more prone to organ injuries. But, despite the improvement in the medical cardiac treatment, for instance, endovascular interventions and robotic surgery, cardiopulmonary bypass remains an essential part of many cardiovascular procedures. The multifactorial nature of inflammatory response suggests that no single pharmaceutical or technical intervention can in isolation inhibit the adverse clinical outcomes of such type of surgery. And more, theoretical and experimental data supporting that negative modulation of systemic inflammatory response (observed during and after cardiopulmonary bypass) might ameliorate brain injury found after cardiac surgery, are not clear. Accordingly, the association between cardiopulmonary bypass-induced inflammation with neurocognitive deficits is still a matter of controversy.

Clermont et al. (37) demonstrated with electron spin resonance spectroscopy measurements the time course and origin of reactive oxygen species release, derived from myocardial source or not, in patients undergoing coronary artery surgery involving cardiopulmonary bypass. Their results demonstrated that systemic oxidative stress occurs in patients undergoing open heart surgery, illustrated by the increased alkyl and alkoxy radicals detected and quantified by electron spin resonance spectroscopy. Other studies have already taken an interest in classical indirect oxidative stress markers such as vitamins, antioxidant plasma status, or thiobarbituric acid reactive substances and the results were controversial. The concept that oxidative stress could influence post-operative outcome in patients subjected to coronary artery bypass surgery also remains a controversial and inconclusive issue (38, 39, 40).

Oxidative stress (measured by lipid peroxidation) also has been compared in patients undergoing coronary artery surgery with or without cardiopulmonary bypass (on-pump or off-pump) (41), and it has been shown to be lesser in the off-pump (without cardiopulmonary bypass) than in the on-pump (with cardiopulmonary bypass) group. These results are not surprising since it is clear that the ischemia and reperfusion involved in on-pump surgery are expected to induce oxidative stress. However, there are some results in the literature indicating that glutathione levels decreased and catalase activity increased to similar values between on-pump or off-pump groups with a little difference between them (41). These observations may support the assumptions of Milei and colleagues (42), that the induction of oxidative stress could be relatively benign. It is interesting to note that, the patients in this study were at low risk with good ventricular function; it is expected, therefore, that these patients could have minimal increases in oxidative stress. The study of Milei and colleagues (42) investigated markers of oxidative stress in a small number of low risk patients (24 in total) undergoing coronary artery bypass surgery. They measure myocardial release of glutathione, myocardial antioxidants (vitamin E and ubiquinol) and lipid peroxidation markers (TBARS) in blood, as well as ultrastructural assessment of tissue injury (from myocardial biopsies) and evaluation of post- ischemic hemodynamic function and clinical outcome. The results show that there was evidence of increased glutathione release in the initial 20 min of reperfusion, and a decrease in tissue antioxidant levels of ubiquinol (but not vitamin E), and minimal increase in tissue lipid peroxidation or any ultrastructural damage. The study indicates that, for the majority of low risk patients undergoing coronary artery bypass surgery, oxidative stress remains a constant underlying factor, unlikely to significantly influence clinical outcome as long as myocardial protection is provided and the ischemic

duration is kept as short as possible. However, in critically ill patients an intervention to attenuate oxidative stress might be considered beneficial, because reactive oxygen species may contribute to myocardial stunning, infarction and apoptosis and vascular dysfunction (1, 2, 3).

2.1.3.3 Neuroinflammation associated with coronary artery bypass grafting surgery (CABG)

It is largely suggested that neurocognitive decline after cardiopulmonary bypass results from an inflammatory response that is initiated by extracorporeal circulation (43, 44, 45). However, a more comprehensive review of the literature does not consistently support this hypothesis. For example, in an animal model that included both elderly rats and diabetic rats, de Lange and colleagues (46) found no differences in short-term neurocognitive performance (8 –14 days after surgery) in rats undergoing surgery with cardiopulmonary bypass, compared with those undergoing a sham operation. They noted an increase in cytokine release (interleukin-6) after cardiopulmonary bypass in diabetic rats, but not in elderly rats. In humans, Westaby and colleagues (47) did not find an association between maximal levels of inflammatory markers (complement C4a and C5b-9) with early or late neurocognitive function after coronary artery bypass graft surgery with cardiopulmonary bypass. Furthermore, Parolari and colleagues (48) demonstrated that postoperative levels of inflammatory markers, including interleukin-6, plasma tumor necrosis factor-alpha, C-reactive protein, and fibrinogen, differed little in patients undergoing coronary artery bypass graft surgery with or without cardiopulmonary bypass.

Nevertheless, it is largely known that necrosis and apoptosis after an acute ischemic event are accompanied by other processes which lead to a posterior neurodegeneration. It was demonstrated that release of cytokines, such as tumor necrosis factor alpha and interleukins, as mediated by oxidative stress, and prolonged microglial activation by interleukin-1 induce to a neuronal degeneration that follow cerebral ischemia (49) and that excessive formation of reactive oxygen species, induce to direct tissue damage and stimulate inflammatory and proapoptotic cascades (50). Central norepinephrine release during brain ischemia also increases neuronal metabolism and carry to the formation of reactive oxygen species from auto-oxidation of neurotransmitters, induce damage caused by glutamate during ischemia and can exacerbate the underlying disease of patients (49).

If the inflammatory response is or not the primary cause of neurocognitive injury after cardiopulmonary bypass, the question is whether or not neurocognitive decline in adult cardiac surgical patients could be related to the cardiopulmonary bypass pump. Van Harten and colleagues (51) discuss the evidence for cardiopulmonary bypass-related neuronal injuries in adult cardiac surgery patients, and review the evidence that immune priming is a key factor in the pathogenesis of cognitive dysfunction after cardiac surgery. They suggest further studies about pathophysiology of post-operative cognitive dysfunction (POCD) that may lead to strategies and therapies to prevent or attenuate POCD and also define the better choice of hypnotic, and dose of opioid, on the inflammatory response to surgery and on the incidence of POCD. These studies could determine the benefit, if any, of immune system modulation, by antiinflammatory agents and also by other drugs that may exert beneficial effects on the balance between pro and antiinflammatory mediators, such as interleukin-6 or tumor necrosis factor-alpha and interleukin-10, respectively.

A comparison of coronary artery bypass graft surgery with percutaneous coronary intervention failed to show difference in cognitive decline in patients undergoing cardiac revascularization (52). Age is considered to be the strongest predictive factor of post-operative cognitive dysfunction (POCD) and coronary artery bypass grafting without the use of cardiopulmonary bypass could be considered less harmful to the these patient group, especially in terms of neurological complications. Although an increasing number of patients with advanced age and other risk factors for neurocognitive injuries have been referred for coronary artery bypass grafting, Jensen and colleagues (53), in a randomized trial, investigated the effect of avoiding the heart-lung machine on cognitive function 1 year after surgery in aged patient population. They did not detect differences in cognitive outcomes in elderly high-risk patients 1 year after the operation between subjects which underwent coronary artery bypass grafting surgery without cardiopulmonary bypass with those subjected to extracorporeal circulation. The study of Jensen and colleagues (53) are in line with other randomized study about late cognitive outcome in younger patients with less advanced coronary artery disease and lower preoperative risk (54). In Jensen study (53), postoperative cognitive dysfunction, unexpectedly, tended to be less common in the on-pump group. This could be further suggestive that many other factors such as inflammatory processes including sternotomy, heparin administration and hemodynamic variations may be responsible for cognitive dysfunction observed after surgery (55). It seems that patient characteristics, such as the presence of atherosclerosis, are more relevant than the type of intervention as predictive factor of neurocognitive injury in patients with severe coronary artery disease (56). In

addition, late cognitive decline occurring 5 to 6 years after coronary artery bypass graft surgery did not differ in degree from longitudinal cognitive decline observed in patients of similar age either with coronary artery disease (57) or without coronary artery disease (46, 58). Perhaps, decline in neuropsychological tests with time is related to progression of underlying cardiovascular disease or simply to natural aging (59, 60, 61). In fact, neurocognitive impairment in many patients undergoing cardiac surgery may be preexisting, although subclinical (62).

2.1.3.4 S-100B as a marker and modulator of neuroinflammation

S100B protein might also participate in the brain inflammatory response. At the nanomolar concentrations found in the brain extracellular space, under normal conditions, S100B acts as neurotrophic factor, promoting neuronal survival under stress conditions and neurite outgrowth (63) and stimulating the uptake of the cytotoxic glutamate by astrocytes (64). The level of S100B in blood is considered a clinical marker of brain cell damage and/or increased permeability of the blood/brain/barrier. Moreover, S100B release by astrocytes can be augmented upon stimulation by the proinflammatory cytokines tumor necrosis factor-alpha, interleukin-1 (IL-1) and interleukin-6 (IL-6) (65, 66, 67, 68). As demonstrated before, cytokines contribute to a cascade of events typical of inflammation and especially proinflammatory cytokines, such as IL-6 and IL-8, are thought to contribute to the development of sickness behavior (69). Trophic effects of the S100B protein on neurons depend on interaction with the receptor for advanced glycation end products (RAGE) (70), a multiligand receptor belonging to the immunoglobulin family that has been implicated in both neuroprotection and neurodegeneration, and in the inflammatory response (71). Acute stimulation of RAGE with high doses of S100B causes neuronal apoptosis via overproduction of reactive oxygen species (72) and stimulates inducible nitric oxide synthase in astrocytes and microglia (73, 74, 75), which might contribute to astrocytic and neuronal apoptosis (75). Moreover, S100B also stimulates interleukin-1 (IL-1) release from microglia (76).

S100B protein increases 50 to 100 fold after cardiac surgery using standard cardiopulmonary bypass (77, 78), a finding that could support association between cardiopulmonary bypass and brain damage. The postoperative serum concentration of S-100B appear to increase with the duration of cardiopulmonary bypass and with the number of cerebral emboli detected by transcranial doppler imaging (79). Several studies have suggested

that, in the absence of clear neurologic signs, transient elevations in serum S100B protein can reflect subclinical cerebral damage (80, 81). But, early release of S100B after cardiopulmonary bypass has not been associated with adverse neurological outcome. In contrast, cerebral complications such confusion, delayed awakening and stroke have been correlated with late increase in S100B detected 5 to 48 hours after cardiopulmonary bypass (82). The increase in plasma S100B could also be linked with postoperative delirium incidence (83) and be a consequence of S100B release by astrocytes stimulated by circulating proinflammatory mediators (IL-6, IL-8, etc) (83, 84). Such complex and not fully well characterized relationship between S100B, inflammatory markers and neurobehavioral changes has been studied in more detail in elderly hip fracture patients (83).

Of particular clinical significance, many studies have demonstrated that the development of delirium in critically ill patients increases morbidity, mortality, and healthcare costs (85, 86). It has been hypothesized also a higher frequency of dementia in patients who presented with delirium at the end of the surgery (87). The neurocognitive impairments might reflect an irreversible brain damage triggered by surgery pathophysiological effects. Consequently, it could be supposed that the higher the level of S100B in a delirious patient, the higher the risk of dementia after delirium, and thus cerebral damage. This cerebral damage could be mediated via neuroinflammatory mechanisms because the level of S100B and the incidence of neurodegeneration are higher in patients with an infectious disease (which normally is associated with inflammatory response) as compared to non infected subjects (88, 89).

2.1.4 Alpha (α)-2-adrenergic receptor agonists

Alpha (α)-2-adrenergic receptor agonists have been utilized in surgery because they have sedative, analgesic, hemodynamic-stabilizing properties and sympatholytic pharmacologic effects (90, 91) (Figure 4).

FIGURE 4 ABOUT HERE

The stress response to surgery can be attenuated by sympatholytic effects caused by postsynaptic activation of central (α)-2-adrenergic receptor, leading to reductions in blood pressure and heart rate (89). Of clinical significance, two adrenergic agonists have been used as coadjuvant in general anesthesia or even as anesthetic agents by themselves, i.e., clonidine and dexmedetomidine (Figure 5). Here we will briefly discuss the use of clonidine, because

clonidine has a smaller selectivity for (α)-2-adrenergic receptor than dexmedetomidine, consequently, a low efficacy as an anesthetic agent.

FIGURE 5 ABOUT HERE

2.1.4.1 Clonidine

Clonidine was first used for postoperative pain relief and regional anesthesia (93,94, 95). In effect, clonidine has antinociceptive properties and reduces anesthetic requirements by attenuating sympathoadrenal responses during surgery and plasma concentrations of norepinephrine by stimulating presynaptic (α)-2-adrenergic receptors. While the use of clonidine during coronary artery bypass graft surgery did not appear to influence the perioperative stress response (96), its immunomodulatory effects remains to be characterized. Of clinical significance, perioperative use of clonidine was associated with reduction in the incidence of myocardial ischemia and death after non-cardiac surgery in patients at risk of coronary disease (97, 98, 99). Von Dossow and colleagues (100) investigated the influence of perioperative clonidine infusion on the early T-cell immune response, in patients undergoing elective coronary artery bypass graft surgery, and demonstrated early T-cell response ratios in the clonidine group 6 h after cardiac surgery. No differences were found with respect to plasma cytokine levels. In contrast to these findings, Ellis and Pedlow (101) reported no influence of clonidine on lymphocytes, but a significant decrease in plasma norepinephrine levels in patients undergoing major noncardiac surgery. The decreased norepinephrine plasma levels after clonidine administration have been previously reported (102), especially in patients undergoing cardiac surgery (103). It has been hypothesized that the major effect of (α)-2-adrenergic receptor agonists is on tonic activity, while sympathetic nervous system responsiveness to stressful stimuli appears to be unaffected.

Here it is important to emphasize that there are few studies about the modulation of inflammatory response after systemic use of clonidine in anesthesia. In a study with 7 patients, preoperative administration of clonidine was associated with a reduction in plasma and cerebrospinal fluid levels of TNF-alpha (104) Similarly, perioperative epidural clonidine administration caused a decrease in blood IL-6 and suggested that (α)-2-adrenergic receptor stimulation can modulate systemic inflammatory response in human (105).

2.1.4.2 Dexmedetomidine

Dexmedetomidine is a selective (α)-2-adrenergic receptor agonist with an increased ratio of (α)-2 to (α)-1 activity of 1.620:1, as compared to clonidine (220:1). In 1999, dexmedetomidine was approved by the United States of America (USA) Food and Drug Administration (FDA) only for sedation of patients. In 2008, based on two randomized, double-blind, placebo-controlled, multicenter trials (106), FDA approved the update labeling use for dexmedetomidine, including the indication for sedation in surgery or other procedures.

Dexmedetomidine is the dextro enantiomer of medetomidine, the methylated derivative of etomidine, and specific (α)-2 adrenergic receptor subtypes mediate its pharmacodynamic effects (Figure 4). Agonism at the (α)-2A adrenergic receptor appears to promote sedation, hypnosis, analgesia, sympatholysis, neuroprotection (107) and inhibition of insulin secretion (108). Agonism at the (α)-2B adrenergic receptor suppresses shivering centrally (109), induces analgesia at spinal cord and vasoconstriction in peripheral arteries. The (α)-2C adrenergic receptor is associated with cognition, sensory processing, mood and regulation of epinephrine outflow from the adrenal medulla (110). Inhibition of norepinephrine release appears to be equally affected by all three alpha-2 receptor subtypes (111) (Figure 6). Dexmedetomidine also binds to imidazoline receptors and this activity may explain some of the non-(α)-2 adrenergic receptor effects of this drug, and receptor subtypes have also been identified. Imidazoline-1 receptors are linked to G-proteins and modulate blood pressure and have anti-arrhythmic effects (90). Imidazoline-2 receptors have been implicated in neuroprotection in a cerebral ischemia model in animals and in acquisition and retention of memory. They are not G-protein coupled receptors and located on the mitochondrial outer membrane and probably exert their effects by decreasing tissue norepinephrine levels (90, 112).

FIGURE 6 ABOUT HERE

2.1.4.3 Dexmedetomidine pharmacokinetics

After intravenous injection, dexmedetomidine has an onset of action after 15 minutes and peak concentrations are achieved within 1 hour after continuous intravenous infusion. Rapid distribution occurs away from the central neurological system with an alpha half-life ($t_{1/2} \alpha$) of 6 minutes and a terminal elimination half-life ($t_{1/2} \beta$) between 2.0 and 2.5 hours. The drug is highly protein-bound, with a 6% free fraction, and has a large steady state volume of

distribution (V_{dss} , 1.33 L. kg^{-1}). Total plasma clearance and protein binding is age independent (113).

Hepatic clearance may be decreased to 50% of normal with severe liver disease. Pharmacokinetics is not significantly altered in patients with severe renal impairment, but patients remained sedated for longer than normal controls, suggesting an enhanced pharmacodynamic effect (114). Thus, dosages should be decreased in the presence of either hepatic or renal diseases. There are no recognized active or toxic hepatic derivatives of dexmedetomidine after its metabolism via glucuronide conjugation and biotransformation by cytochrome P450 enzymes.

Intravascular doses of dexmedetomidine induced dose-dependent decreases in systolic and diastolic blood pressure and in heart rate with important decreases in plasma norepinephrine levels. However, at high-bolus intravascular doses (50-75 μg), a transient initial hypertensive response may be seen, because an activation of peripheral vascular (α)-2B adrenergic receptors before the central sympatholytic effect on the vasomotor center occur (115). Dexmedetomidine apparently does not induce alterations in plasma renin activity, atrial natriuretic peptide or arginine vasopressin levels (116).

Targeted plasma dexmedetomidine levels revealed desirable pharmacodynamic effects between 0.5 and 1.2 ng. mL^{-1} . Subsequent clinical studies designed to achieve these effects used a loading dose of 1 $\mu\text{g. kg}^{-1}$ during a period of 10 minutes, followed by a continuous intravenous infusion rate of 0.2 to 0.7 $\mu\text{g. kg}^{-1}. \text{h}^{-1}$, the dosing regimen originally approved by the USA Food and Drug Administration in 1999. Studies examining very high dexmedetomidine plasma levels (up to 8.0 ng. mL^{-1}) demonstrate that the (α)-2B peripheral vasoconstrictor effects become predominant, with increasing systemic vascular resistance and decreasing cardiac index, associated with marked catecholamine suppression and deepening sedation. Even at these very high plasma levels of dexmedetomidine, there was no clinically significant respiratory depression (117) and it appears to be safe. Case reports of large accidental overdoses of dexmedetomidine describe oversedation as the only important effect, with resolution within an hour of discontinuation (118). There are reports of dexmedetomidine safely use as the sole agent at high rates of infusion (5-15 $\mu\text{g. kg}^{-1}. \text{h}^{-1}$) to anesthetize patients with tracheal stenosis while preserving spontaneous ventilation (119). In October 2008, the US Food and Drug Administration approved an increased dose of dexmedetomidine (up to 1.5 $\mu\text{g. kg}^{-1}. \text{h}^{-1}$) for surgical procedures.

2.1.4.4 Dexmedetomidine analgesic and sedative effects

Dexmedetomidine possesses analgesic properties and other advantageous pharmacological effects that make it a potential useful and safe adjunct in several clinical applications, as demonstrated Sleigh in a recent review (120). When used as an adjunct to general anesthesia, dexmedetomidine can reduce both the minimum alveolar concentration requirement of inhalation agents and provide opiate-sparing properties up to 90% (121).

The mechanism by which (α)-2-adrenergic receptor agonists produce analgesia and sedation is multi-factorial. Both, hypnotic and supra-spinal analgesic effects of dexmedetomidine are mediated by noradrenergic neurons. Dexmedetomidine causes inhibition of norepinephrine release and its neuron associated activity in the descending medullo-spinal noradrenergic pathway and suppresses neuronal activity in the *locus coeruleus* (122). Suppression of these inhibitory controls causes release of mediators and neurotransmitters that decrease the secretion of histamine and produce hypnosis, similar to normal sleep, without evidence of depression of ventilation (123). The suppression of activity along the descending noradrenergic pathway terminates propagation of pain signals, resulting in analgesia or decreased awareness at noxious stimuli. In neurons of the superficial dorsal horn of the spinal cord, dexmedetomidine suppresses and reduces pain transmission by inhibiting the release of glutamate and substance P (nociceptive transmitters) from primary afferent terminals and with G-protein-mediated activation of potassium channels causing hyperpolarization of inter-spinal neurons. Antinociception may also be provided by non-spinal mechanisms, as demonstrated in intra-articular administration of dexmedetomidine during knee surgery, which was associated with improved postoperative analgesia, with less sedation than the intravenous route (124). The suggested mechanisms are activation of alpha-2A adrenoreceptors (125) inhibition of the conduction of nerve signals through C and A δ fibers, and the local release of encephalin. Figure 7 demonstrated the possible effector mechanisms of the (α)-2-adrenoreceptors, linked to G proteins.

FIGURE 7 ABOUT HERE

Dexmedetomidine provides dose-dependent increases in anxiolysis and sedation that appears to be unique in comparison with GABAergic agents such as midazolam or propofol. Arousalability is maintained at deep levels of sedation (126) and once aroused, patients normally performed well the tests of vigilance (127), and they can cooperate with nursing, radiologic, and airway procedures (128). There appears to be particular value in a drug such

as dexmedetomidine that facilitates the arousal and rapid orientation of a sedated patient. The amnestic effects of dexmedetomidine are far less than the benzodiazepines, which provide profound anterograde amnesia that may contribute to confused states on emergence. In contrast, amnesia is achieved with dexmedetomidine only at high plasma levels ($\geq 1.9 \text{ ng. mL}^{-1}$), without retrograde amnesia (117).

Unlike opioids, dexmedetomidine achieves its sedative, hypnotic, and analgesic effects without causing any clinically relevant respiratory depression, even when dosed to plasma levels up to 15 times those normally recommended for therapy (117). Sedation induced by dexmedetomidine has the respiratory pattern and electroencephalogram (EEG) changes comparable with natural sleep. Compared with remifentanil, hypercapnic arousal is preserved (129) and functional magnetic resonance imaging studies show that unlike GABAergic agents, dexmedetomidine preserves a cerebral blood flow pattern from natural sleep (130).

Administration of dexmedetomidine during sevoflurane or desflurane anesthesia with spontaneous ventilation has no effect on end-tidal carbon dioxide (131) and arterial saturation is better preserved with dexmedetomidine than propofol under magnetic resonance imaging procedures (132). In contrast to infusions of opioids, benzodiazepines, or propofol, dexmedetomidine can safely be infused through tracheal extubation and beyond. It has been used successfully to facilitate tracheal extubation in patients who had previously failed extubation because of excessive agitation (133).

2.1.4.5 Antiinflammatory effects of dexmedetomidine

Reactive oxygen species (ROS) are considered as key regulatory molecules vital for life, but they cause cellular and organ damage when produced in excess or when antioxidant defenses are overwhelmed such in cardiac and neurologic ischemic and reperfusion injury. ROS can contribute to myocardial stunning, infarction and apoptosis, to the genesis of arrhythmias and neurologic deficits. Several intravascular anesthetic drugs can act as reactive oxygen species scavengers. It was demonstrated in patients with impaired preoperative left ventricular function undergoing elective coronary artery bypass surgery with cardiopulmonary bypass, that the administration and maintenance of a clinically relevant dose of propofol from before aortic cross-clamp release, maintained until 4 hours after reperfusion, attenuate myocardial lipid peroxidation, associated with a decrease in IL-6 production and a late increase of IL-10 release (134).

Recently, Arslan and colleagues (2012) concluded that dexmedetomidine protected liver from lipid peroxidation, when given before induction of ischemia in an experimental model (135). Rocha and colleagues have also indicated the protective potential of dexmedetomidine in women which underwent pelvic videolaparoscopic surgery (136). In their study dexmedetomidine protected blood aminolevulinate dehydratase (ALA-D) from inactivation caused by hyperoxygenation in total intravenous anesthesia. The results of the investigation indicated that blood ALA-D from patients anesthetized with dexmedetomidine was not modified by exposure to high concentrations of oxygen, whereas the activity of enzyme from those patients anesthetized with remifentanil exhibited a statistical significant decrease in activity. Regarding the dexmedetomidine group, it is possible that the anesthetic has protected the enzyme from oxidation by hyperoxygenation process.

Several investigators have published reports about the effects of dexmedetomidine and other (α)-2-adrenergic receptors agonists on cytokines (137) and on (α)-2-agonists modulated lipopolissacaride-induced tumor necrosis factor- α production by macrophages (138). Taniguchi and colleagues (139) demonstrated that dexmedetomidine has an inhibitory effect on cytokine responses to endotoxemia. These findings suggest that one of the mechanisms of antiinflammatory effects of dexmedetomidine may be *via* modulation of cytokine production by macrophages and monocytes. Hofer and colleagues demonstrated that dexmedetomidine infusion decreased cytokine production in sepsis (140), which is in accordance with a recent study showing reno and pulmonary protective effect of dexmedetomidine in an experimental model of sepsis in rats (33). They have shown that preventive administration of clonidine or dexmedetomidine improved survival in induced sepsis. This was accompanied by a reduction in the proinflammatory mediators IL-1 β , IL- 6 and tumor necrosis factor- α . Furthermore, they suggested that, administration of a central acting (α)-2- adrenoreceptor agonist might be considered as a preventive therapeutic option in high-risk patients undergoing major surgery. In another animal study, dexmedetomidine treatment was equally effective to methylprednisolone in reducing TNF- α and IL-6 levels induced by spinal cord injury. Apparently, dexmedetomidine treatment reduced neutrophils' infiltration at the site of spinal cord injury (141). Dexmedetomidine inhibited cortisol synthesis at supratherapeutic concentrations but this has not been reported in short-term use in humans (142,143). Our study group, have the influence of dexmedetomidine on cortisol levels evaluated (144). At this study, we measured cortisol concentrations before anesthetic induction, 5 minutes after intubation, and 30 minutes after surgical incision in patients undergoing gynecologic videolaparoscopic surgery, receiving dexmedetomidine or remifentanil. After intubation, there

was a significant decrease in cortisol concentrations from baseline in both groups ($-4.3 \pm 1.4 \mu\text{g. dL}^{-1}$ and $-4.6 \pm 1.6 \mu\text{g. dL}^{-1}$, respectively) but only in the remifentanil group at 30 minutes after incision ($2.6 \pm 1.8 \mu\text{g. dL}^{-1}$ and $-7.1 \pm 2.1 \mu\text{g. dL}^{-1}$) and we could conclude that dexmedetomidine did not suppress steroidogenesis (144). In the MENDS trial (145), cortisol concentrations were determined at baseline and 2 days after stopping dexmedetomidine infusion, and there was no statistically significant difference in cortisol concentrations. At high doses as $1.5 \mu\text{g. kg}^{-1}. \text{h}^{-1}$ dexmedetomidine does not appear to cause clinically significant adrenal suppression (146).

Laringoscopy and endotracheal intubation also provoke marked sympathetic and sympathoadrenal response that increase the risk of perioperative myocardial ischemia and infarction. The perioperative use of dexmedetomidine may improve endocardial perfusion and decrease heart rate with attenuation of stress response (147). Dexmedetomidine increases the hemodynamic stability by altering the stress-induced sympatho-adrenal responses to intubation, during surgery and emergence from anesthesia (148) and this reflect a better outcome.

In a recent study, Sukegawa and colleagues (149) described the potent inhibitory effect of dexmedetomidine on inflammatory reactions, including edema, accumulation of inflammatory cells, and production of tumor necrosis factor-alpha and cyclooxygenase-2 (COX-2), induced by an injection of carrageenin into the paw of mice. They have also demonstrated a potent antiinflammatory effect of dexmedetomidine at a high dose on endotoxin-induced inflammation in murine macrophages (150).

Yagmurdur and colleagues (151) have examined the effect of dexmedetomidine on ischemia-reperfusion injury due to tourniquet during upper-extremity surgery by determining blood malondialdehyde and hypoxanthine levels. Dexmedetomidine significantly attenuated plasma hypoxanthine production in the ischemia and plasma malondialdehyde production in the reperfusion periods. They suggest that dexmedetomidine can have advantages over other anesthetic agents (for instance, opioids and propofol) by inhibiting lipid peroxidation in the case of anticipated ischemia-reperfusion injury, such as would occur in upper-extremity surgery requiring tourniquet application. Bekker and colleagues (152) hypothesized that the intraoperative administration of dexmedetomidine could reduce the stress response and improve the quality of recovery in patients undergoing major spinal surgery. They compare a propofol/fentanyl/dexmedetomidine anesthesia group with propofol/fentanyl/placebo-saline anesthesia. In both groups, plasma cortisol levels were elevated in the postanesthesia care unit, whereas C-reactive protein levels were elevated only in the first postoperative day.

Dexmedetomidine significantly reduced the levels of cortisol, but not those of C-reactive protein. Levels of cytokines IL-6 and IL-8 were significantly higher immediately after surgery and at first postoperative day. Dexmedetomidine delayed postoperative rise of IL-10 but not of IL-6 or IL-8. Plasma levels of others cytokines were not affected by surgery. Clinically, dexmedetomidine infusion moderately improved the quality of recovery (106).

Gu and colleagues also conducted a study (153) to investigate dexmedetomidine antiinflammatory capacity. They utilized an animal model of renal ischemia-reperfusion that induced an acute lung injury and either pre-treated mice with dexmedetomidine ($25\mu\text{g. kg}^{-1}$ before ischemia) or gave it after reperfusion. Renal ischemia/reperfusion induced an increase of inflammatory markers in lungs (mieloperoxidase (MPO) activity, intercellular adhesion molecule-1(ICAM-1) and TNF- α mRNA level). Both pre- and post-treatment with dexmedetomidine markedly reduced lung edema and inflammatory response and lowered MPO activity and ICAM-1 and TNF- α mRNA expression. Other study explored the antiinflammatory effects of dexmedetomidine in rats, using an intravenous infusion of dexmedetomidine at the rate of $5.0\ \mu\text{g. kg}^{-1}\cdot\text{h}^{-1}$ after bilateral blunt chest trauma-induced pulmonary contusion (154). Dexmedetomidine not only significantly modified hemodynamics and relieved the infiltration of inflammatory cells into alveolar spaces but also inhibited the injury-induced increase in plasma TNF- α and IL-1 β production.

In humans, Kang and colleagues (155) demonstrated the antiinflammatory dexmedetomidine effects in patients subjected to laparoscopic cholecystectomy. Patients in the dexmedetomidine group received a loading dose of dexmedetomidine ($1.0\ \mu\text{g. kg}^{-1}$), followed by infusion of dexmedetomidine at $0.5\ \mu\text{g. kg}^{-1}\cdot\text{h}^{-1}$. Dexmedetomidine decreased the plasma level of IL-1 β , TNF- α , and IL-10, when compared to saline group. The C-reactive protein (CRP) level and leukocyte count on post-operative day 1 were also lower in dexmedetomidine group. Tasdogan and colleagues (156) conducted other study to compare the effects of an intravenous infusion of propofol and dexmedetomidine, on inflammatory responses and intra-abdominal pressure in severe sepsis after abdominal surgery. Dexmedetomidine infusion decreases tumor necrosis factor-alpha, IL-1, and IL-6 levels and intra-abdominal pressure significantly more than a propofol infusion.

2.1.4.6 Neuroprotective effects of dexmedetomidine

The brain has a high requirement for oxygen and glucose, but is unable to store these substrates and rapid necrosis occurs to hypoxic-ischemic injury. It results in dysfunction of

adenosine triphosphate (ATP) dependent ion channels and pumps, leading to cellular depolarization and the release of extracellular excitatory neurotransmitters. The most important neurotransmitter is glutamate, which activates the N-methyl-D-aspartate receptor (NMDA), increasing intracellular calcium and sodium, contributing further to depolarization and neuronal activation. Excess of calcium promotes activation of pathways which disrupt ionic homeostasis, leading to membrane degeneration and excitotoxic cell death (157). Apoptotic mechanisms are also activated in response to ischemic brain injury, days to weeks after ischemic insult, especially in the region surrounding the necrotic area (158).

Neurological injury remains a major cause of morbidity in cardiac surgery patients and, in an extensive review, Hogue and colleagues (43) concluded that about 60% of patients have evidence of cognitive decline one month after cardiac surgery. Central nervous system deficits after cardiopulmonary bypass ranging from postoperative cognitive dysfunction (POCD), with incidence of 30- 60% (159, 160) to stroke, over 1-5% of patients (161). Adverse cerebral outcomes after cardiac surgery have been studied for a long time and literature data suggest that modalities modifying the systemic inflammatory response to cardiopulmonary bypass might protect brain against potential injury after cardiac surgery (44, 162, 45). But, the association between cardiopulmonary bypass-induced inflammation and neurocognitive deficits itself remains less than clear. A review of the literature did not support neurocognitive decline after cardiopulmonary bypass as a result of an exacerbated inflammatory response initiated by extracorporeal circulation. In recent issue, Jungwirth and colleagues (61) have published a well-controlled study in a rat model that fails to demonstrate a relationship between neurologic injury and the foreign surface area of cardiopulmonary bypass or donor blood used to prime the cardiopulmonary pump. They have suggested that other factors than cardiopulmonary bypass lead to adverse neurocognitive outcomes after cardiac surgery. Elsewhere, neurocognitive impairment in many patients undergoing cardiac surgery may be preexisting, although subclinical (62, 163, 164, 165, 166, 167), and the cognitive outcomes for patients needing cardiac surgery with cardiopulmonary bypass appear to depend little on the perfusion technique, but rather on the underlying diseases (168). However, inflammatory response, oxidative stress and massive extracellular catecholamine release may lead to additional neurodegeneration (168).

Nevertheless, Singh and colleagues (169) concluded that anesthetic choice in patients under cardiac surgery may have implications on S100B protein serum levels, a neuroinflammatory component, that could be a marker for brain injury on serum (170) and/or

damage to blood-brain barrier (76). In other trials (171, 145, 172), patients receiving dexmedetomidine developed significantly less delirium compared with patients receiving other drugs, such midazolam or propofol in intensive central unit. The pathogenesis of postoperative delirium is not completely clear but appears to be related, in part, to increased release of inflammatory mediators and the binding to the gama-aminobutyric acid (GABA) receptor (172). Dexmedetomidine does not bind to the GABA receptor and hence may minimize the development of delirium by decreasing release of norepinephrine. Because of conflicting results (173) more studies are needed to determine whether dexmedetomidine can really prevent or treat postoperative associated delirium. Many anesthetics act as gama-aminobutyric acid (GABA) receptor agonists, and in animal models, a GABA agonist can suppress neural cell proliferation, whereas GABA antagonist can enhance neurogenesis (174, 175). Dexmedetomidine acts by reducing noradrenergic output from the *locus coeruleus*, and decreasing brain norepinephrine levels, and in animals, manipulations that decrease brain norepinephrine also suppress cell proliferation (176, 177). Inhaled anesthetics such as isoflurane inhibit the cholinergic basal forebrain, and suppress hippocampal neurogenesis in animals (178). However, Tung and colleagues (179) found no effect of prolonged (8 hours) anesthesia with isoflurane, propofol, or dexmedetomidine on hippocampal cell proliferation in 3 or 12-months-old Sprague-Dawley rats. These results suggest that the sum of the many potential mechanisms linking cell proliferation to the anesthetized state (vigilance state, environmental stimuli, adrenal effects of anesthesia, direct pharmacologic effects), result in no overall effect and that suppression of adult hippocampal cell proliferation is unlikely to be an effect of brief or prolonged anesthesia, and thus unlikely to cause postoperative cognitive dysfunction in humans.

The neuroprotective effects of dexmedetomidine have been also demonstrated *in vivo* and *in vitro* in a variety of models of ischemia. These include models of incomplete ischemia in the rat (180, 181), transient focal ischemia in rabbits (182) and transient global ischemia in gerbils (183). *In vitro* studies of neuronal injury, using hippocampal slices (184) and neuronal and cortical cell cultures (185) also support dexmedetomidine as a neuroprotectant drug.

Originally, all dexmedetomidine neuroprotective activities were supposed to be caused by inactivation of presynaptic (α)-2-adrenergic receptors, inhibiting noradrenergic activity. However, dexmedetomidine concentrations well below 100 nM exert prominent effects on cultured astrocytes (185, 186) and the (α)-2-adrenoceptor is densely expressed in astrocytes freshly isolated from mouse brain by fluorescence-activated cell sorting (187). Thus, instead

of receiving a subtype-mixed noradrenergic signal from *locus coeruleus* the cells can be directly activated at their (α)-2-adrenoceptor sites by the drug.

The (α)-2-adrenergic signaling pathway has been studied in cultured astrocytes (186, 188). It connects activation of (α)-2-adrenoceptors with extracellular signal-regulated kinase (ERK) phosphorylation in two-stages, separated by trans-activation of the epidermal growth factor (EGF) receptor. This receptor is highly expressed in both neurons and astrocytes. In the first stage, the $\beta\gamma$ subunits of the activated heterotrimeric Gi protein lead, *via* activation of cytosolic Src tyrosine kinases, to metalloproteinase-mediated ‘shedding’ of heparin-binding epidermal growth factor (HB-EGF) from its transmembrane-spanning HB-EGF precursor. In the second stage, released HB-EGF ‘transactivates’ EGF receptors in the same and adjacent cells (including neurons) by phosphorylating EGF receptors, leading to Ras- and Raf-dependent ERK phosphorylation (186, 188). The astrocytic effects may contribute also to dexmedetomidine’s analgesic effects, at least in the spinal cord (189, 190).

FIGURE 8 ABOUT HERE

Despite dexmedetomidine has repeatedly been found to have neuroprotective effects against ischemia in experimental models (191) and could be able to protect against trauma in hippocampal organotypic cultures (192), these neuroprotection capacity have not been confirmed clinically.

More recently, Zhang and colleagues (193) described a possible mechanism through which dexmedetomidine induces neuroprotection. Based on knowledge that oxidative damage contributes greatly to post-traumatic brain injury (194) they induced oxidative neuronal injury with H_2O_2 in the glutamatergic cerebellar granule neurons. The hypothesis was tested that ‘conditioned’ medium from dexmedetomidine-treated astrocyte cultures would enhance neuronal viability due to release of an epidermal growth factor (EGF) receptor agonist, whereas direct administration of dexmedetomidine to neurons or treatment with non-conditioned medium would have no effect. Furthermore, it was examined if the protection found after addition of medium from dexmedetomidine treated astrocytes was abolished by treatment of the astrocytes with the specific (α)-2-adrenergic antagonist atipamezole. This was confirmed, but atipamezole addition directly to H_2O_2 - exposed neurons treated with dexmedetomidine had no effect. They demonstrated, that dexmedetomidine at clinically relevant concentrations, was neuro-protective against oxidative damage by stimulating directly the astrocytic (α)-2-adrenoceptors, causing release of heparin-binding epidermal

growth factor (HB-EGF). HB-EGF in turn activates neuronal epidermal growth factor (EGF) receptors. At these concentrations however, dexmedetomidine has no direct neuronal effect (193).

The practice of neuroprotection is difficult because the process of neuronal damage and cell death is complex and not completely understood. The complexity of pathophysiologic mechanisms suggests that neuroprotection may have a multimodal approach and it is unlikely that a single pharmaceutical agent will be effective in improving neurological outcome. Recent evidence indicates, indeed that new neurons are produced in the adult hippocampus (195), and play a functional role in cognitive processes such as learning and memory (196, 197). Because anesthetics also affected these factors, it can be suspected that anesthetics or the anesthetized state also affected adult hippocampal cell proliferation. Anesthetic management may thus improve the quality of recovery in patients undergoing coronary artery bypass graft surgery, affecting the postoperative course, reduce the stress response and possibly reduce neurological deficits onset.

2.1.4.7 Dexmedetomidine as protective agent against ischemia

Regarding the cerebral circulation in humans, during cardiopulmonary bypass, relatively little information was available until Henriksen and colleagues (198) reported evidence of cerebral hyperemia in 1983. In 1984, Govier and colleagues (199) incite controversy and debate with their observations of ischemic threshold levels of cerebral blood flow during cardiopulmonary bypass. Murkin and colleagues (200), subsequently, reported a decrease in cerebral blood flow and metabolic rate (oxygen consumption) during hypothermic cardiopulmonary bypass in humans. These low values were restored to control levels shortly after separation from cardiopulmonary bypass system. This study demonstrated a physiological basis for the embolic theory of central nervous system impairment after cardiac surgery. Ganushchak and colleagues (201) tested with a retrospective study the hypothesis that combinations of hemodynamic events from apparently normal cardiopulmonary bypass procedures are related to the development of postoperative neurological complications and affect the impact of patient common clinical risk factors on postoperative neurological complications. Patients who underwent cardiopulmonary bypass procedures with large fluctuations in hemodynamic parameters particularly showed an increased risk for the development of postoperative neurological complications (201).

There are increasing points of evidence both *in vitro* and *in vivo* which indicates that dexmedetomidine has a cell-protective effect on nervous tissue under ischemic conditions (202, 203, 204, 205). Considering that ischemia enhances the formation of reactive oxygen species in brain tissue and the activation of brain cells as microglia to synthesize cytokines, Eser and colleagues (206) investigated the neuroprotective effects of dexmedetomidine on an animal model of transient global cerebral ischemia-reperfusion injury. They showed a lower number of apoptotic neurons at hippocampus and decreased levels of cytokines on dexmedetomidine group as compared to the saline control group. These results indicated a clear neuroprotective effect of dexmedetomidine after transient global cerebral ischemia-reperfusion injury.

In a recent review, Afonso and Reis (207) observed that dexmedetomidine seems to have promising applications on neuro- and cardioprotection, and may confer this protection by targeting a number of different areas. The attenuation of ischemia-elicited increase in blood catecholamine levels and a limitation of excitotoxicity from glutamate might be involved in the protective underlying mechanism of dexmedetomidine. But, more evidence has been obtained suggesting that this effect can be mediated also by the stimulation of imidazoline-receptors (208). The signal transduction cascade linked to these receptors comprises extracellular signal-regulated protein kinase 1 and 2 and is known to be an important regulator for cell survival and mediator of neuroprotective effects of various agents (209). Dexmedetomidine was reported also to be effective in protecting against focal ischemia in rabbits, in cardiac ischemia-reperfusion injury in rats, in kidney ischemia-reperfusion injury in rats, and in incomplete forebrain ischemia in rats (210, 211, 202).

There is considerably more experimental evidence that dexmedetomidine has neuroprotective effects by sympatholysis, preconditioning, and attenuation of ischemia-reperfusion injury (112) and under decreases on cerebral blood flow (213, 214, 215) with its ratio with cerebral metabolic rate to be preserved (216).

Schoeler and colleagues (217) found that dexmedetomidine has a protective effect on hippocampal slice cultures subjected to a focal mechanical trauma, with the observed trauma reduction being significantly more pronounced than observed in slices treated with hypothermia. But other studies have indicated conflicting results (218). These authors investigated twenty-four patients, aged 50-70 years, undergoing coronary artery bypass graft surgery, randomized into two groups: those receiving dexmedetomidine (group D) and those which did not receive it (group C). As basal blood samples from arterial and jugular bulb catheters were drawn, dexmedetomidine ($1 \mu\text{g. kg}^{-1}$ bolus and infusion at a rate of $0.7 \mu\text{g. kg}^{-1}$

$^1 \cdot h^{-1}$) was administered to patients in group D. Arterial and jugular venous blood gas analyses, serum S-100B protein (S-100B), neuron-specific enolase (NSE) and lactate measurements were performed after induction, 10 minute after the initiation of cardiopulmonary bypass, 1 minute after declamping, at the end of cardiopulmonary bypass, at the end of the surgery and at 24 hours after surgery. No significant differences between-groups were found regarding arterial and jugular venous pH, PO₂, PCO₂ and O₂ saturations. S-100B, NSE and lactate levels were also similar between groups D and C. During the postoperative period, there were no clinically overt neurological complications in any patient. Cerebral ischemia marker (S-100B, NSE and lactate) patterns were increased during cardiopulmonary bypass, as expected; however, there were no differences between the groups, which led to believe that during coronary artery bypass graft surgery dexmedetomidine has no neuroprotective effects (218).

At periféric level, in spinal cord, dexmedetomidine can preserve neurologic function in mice after aortic cross-clamping, as demonstrated by Bell and colleagues (219). It was also observed that mice exhibited almost complete reversal of the protective effect with the administration of the (α)-2A receptor antagonist atipamezole. Dexmedetomidine appears to attenuate spinal cord ischemia-reperfusion injury via (α)-2A receptor-mediated agonism (219).

At renal site, Gu and colleagues investigated whether the (α)-2-adrenoceptor agonist dexmedetomidine provides protection against ischemia-reperfusion induced kidney injury *in vitro* and *in vivo* (220). Pre- or post-treatment with dexmedetomidine provided cytoprotection, improved tubular architecture and function following renal ischemia. Associated with this cytoprotection, dexmedetomidine reduced plasma high-mobility group protein B1 (HMGB-1) elevation when given prior to or after kidney ischemia-reperfusion, and pre-treatment also decreased toll-like receptor 4 (TLR4) expression in tubular cells. Dexmedetomidine treatment promoted long-term functional renoprotection, and even increased survival following nephrectomy. However, prospective human studies establishing a benefit of dexmedetomidine against kidney damage are not yet available.

Despite its increased clinical use and potential benefits, the effect of dexmedetomidine on inflammation and neuroprotection remains limited and somewhat controversial. Future investigators may examine the clinical benefits of the use of dexmedetomidine, and the correlation of better neurological outcome with anesthesia choice.

2.1.4.8 Dexmedetomidine hemodynamic and myocardial protective effects

Dexmedetomidine has complex hemodynamic effects specific to its activation of pre- and postsynaptic (α)-2-adrenergic receptors. These effects are dose-dependent and biphasic: vasodilation at lower dosages, vasoconstriction at higher dosages and an initial short-term increase in blood pressure followed by a longer lasting reduction in blood pressure and heart rate. Several investigations have identified the cardiovascular effects of dexmedetomidine (221, 222, 117), however its effect on intraoperative hemodynamics during a propofol-supplemented remifentanil-based anesthesia regimen, which produces a strong vasodilatory effect, has not been well investigated.

There is a latent risk for excessive bradycardia and even sinus arrest when dexmedetomidine is administered in combination with sympatholytic or cholinergic agents (beta-blockers, fentanyl, neostigmine), especially with concomitant vagal stimulation (sternal separation) (223, 224, 225). Dexmedetomidine causes dose-dependent decreases in heart rate and blood pressure, concomitant with decreasing plasma catecholamines. This is of considerable benefit in tachycardic, hypertensive patients with improvement of hemodynamic stability in the perioperative period. These effects, however, may be unwanted in patients with congestive heart failure, whose cardiac output is rate dependent, or with conduction system disease. As mentioned, a high-dose bolus may result in a biphasic response, with bradycardia and hypertension consequent to initial stimulation of peripheral (α)-2B vascular receptors, followed by central sympatholysis and a decline in blood pressure (226). Unlike clonidine, cessation of dexmedetomidine administration does not appear to be associated with rebound hypertension or agitation.

The ability of (α)-2 receptor agonists to decrease tachycardia and hypertension suggests that they may play a role in cardioprotection by enhancing myocardial oxygen balance. There is little evidence that dexmedetomidine could enhance myocardial ischemic preconditioning or attenuates reperfusion injury, for example, when used after cardiac surgery, dexmedetomidine decreased the incidence of ventricular arrhythmias from 5% to zero, compared with propofol (117). Other authors have described the safe use of dexmedetomidine as an anesthetic adjunct in coronary artery bypass grafting, improving a stable hemodynamic status (227, 228).

Guo and colleagues (228) investigated the protective effects of dexmedetomidine on left ventricular contractile performance under myocardial hypoxia. Their study indicates that hypoxia immediately impairs left ventricular function with a rapid increase in coronary flow

followed by a gradual decrease, with poor recovery of left ventricular function at reoxygenation. Dexmedetomidine administration only in prehypoxia, improve recovery of left ventricular function and coronary flow, and the protective effects are antagonized by yohimbine. The mechanisms are not clear, but there are several possibilities.

Dexmedetomidine might exert the protective effect on left ventricular dysfunction through inhibition of the release of norepinephrine as suggested also by Chen and colleagues (229) that showed the post-ischemic heart had a large amount of coronary norepinephrine overflow and that its reduction significantly improved the recovery of postischemic left ventricular function in an isolated working heart preparation. As demonstrated before, it is believed that high interstitial concentrations of norepinephrine result in myocyte calcium overload and cell death causing development of cardiac dysfunction (230). The high plasma concentrations of catecholamines (norepinephrine and epinephrine) would lead to a calcium overload into the myocardial cells, increased cytosolic and intramitochondrial calcium, reactive oxygen species release, and adenosine triphosphate (ATP) depletion, with resulting electrocardiogram (ECG) changes, failing in myocardial contraction and possible cell death (231, 232). In patients under extreme sympathetic discharge caused by an acute stress a tissue lesion characterized by contraction of sarcomeric myofibrilles, and interstitial mononuclear infiltration was described (233). Ebert and colleagues (234) reported that dexmedetomidine diminished the hemodynamic and norepinephrine response to the activation of cardiac sympathetic nerves by the cold pressor test. Dexmedetomidine could also prevent a myocardial ischemia-induced norepinephrine release in anesthetized dogs (235).

Dexmedetomidine increase the cyclic adenosine monophosphate (cAMP) level in the coronary artery. Guo and colleagues (228) demonstrated that hypoxia caused an immediate increase in coronary flow followed by a gradual decrease, similarly to the results of Karmazyn and colleagues (236), and reoxygenation resulted in poor recovery. Pinsky and colleagues (237) reported that the graft vasculature with hypoxia impaired vascular function and decreased blood flow after transplantation, and it enhanced phosphodiesterase activity and caused a time dependent decline in cAMP levels in the vascular smooth muscle cells. Kitakaze and colleagues (238) reported that an increase in cAMP level by stimulation of adenosine receptors was amplified by the (α)-2-adrenergic stimulation in the coronary artery. Thus it is possible that dexmedetomidine could increase the cAMP level and attenuate coronary vascular damage of an adenosine-induced coronary vasodilative effect and preserve coronary flow. Kitakaze and colleagues (238) provided evidence that alpha-2-adrenergic stimulation increased coronary flow during ischemia as a result of enhancement of adenosine-

induced coronary vasodilation, although (α) -2-adrenergic stimulation exerted prominent vasoconstriction in nonischemic hearts. However, in Guo study, dexmedetomidine did not significantly improve the coronary flow during hypoxia, contesting this hypothesis.

A large European study demonstrated that perioperative infusion of mivazerol, another (α) -2-adrenergic agonist, significantly decreased cardiac death after vascular surgery in patients with known coronary artery disease (239). And, a meta-analysis of noncardiac vascular surgery patients receiving any (α) -2-adrenergic agonist agent demonstrated decreased risk of myocardial infarction and death (240), but dexmedetomidine alone on cardiovascular outcomes after noncardiac surgery did not show statistical significance (241). Therefore, larger studies are required to clearly ascertain the cardioprotective effect of dexmedetomidine and they whether or not should be included in patients at high cardiac risk.

2.1.4.9 Dexmedetomidine other potential effects

The effects of dexmedetomidine on renal function are complex. Alpha-2 agonists exert a diuretic effect with decreased salt and water reabsorption (242). There are experimental evidence that dexmedetomidine attenuates murine radiocontrast nephropathy by preserving cortical blood flow (243). This mechanism is supported by the observation that dexmedetomidine decreases the renal cortical release of norepinephrine (211). There are also evidence that dexmedetomidine attenuates murine ischemia-reperfusion injury (171). However, prospective human studies cannot establish renal benefits of dexmedetomidine.

Based on preliminary studies, the USA Food and Drug Administration approved duration of infusion of dexmedetomidine remains 24 hours. However, there are several studies that have demonstrated safe use for a week or longer in mechanically ventilated critically ill patients (244). With prolonged administration, tolerance to dexmedetomidine's hypnotic effects has been demonstrated in animals (245), but it does not appear to be clinically significant.

Dexmedetomidine also suppress shivering, possibly by their activity at (α) -2B receptors in the hypothalamic thermoregulatory center of the brain (246). Low-dose dexmedetomidine has an additive effect with meperidine on lowering the shivering threshold, when these drugs are combined (247). Dexmedetomidine may be beneficial in decreasing patient discomfort and oxygen consumption that occur on postoperative shivering (248).

Alpha-2 adrenergic agonists, such as clonidine, have also an established role in the treatment of central hyperadrenergic states induced by withdrawal of drugs, including cocaine, alcohol, or opioids. Numerous case reports of successful treatment of withdrawal using dexmedetomidine have been published (226, 249, 250, 251, 252), but to date, no randomized trials have been performed.

2.1.5 Conclusions

Until December 2007, when results of the MENDS (Maximizing Efficacy of Targeted Sedation and Reducing Neurologic Dysfunction) trial were published, most of the data published on dexmedetomidine were from its use in surgical patients unique as a coadjuvant anesthetic (106). Administered intravenously, dexmedetomidine has been used for sedation and anxiolysis in the intensive care unit and as others additional perioperative uses: premedication, to reduce emergence delirium and postoperative pain, and to attenuate the stress responses associated with surgery and anesthesia. But, it will be necessary to explore the pharmacological mechanisms for the actions of these (α)-2-adrenergic receptor agonist in more detail. The incipient clinical use of dexmedetomidine can be ascribed to its recent introduction as anesthetic in human and veterinary practices. Since inflammation is normally a component of surgery-associated injuries, it would be valuable to have a safe and effective means of preventing inflammatory response to major surgery, especially to coronary artery bypass grafting, and its complications, with the beneficial actions of anesthetic drugs. We believe that dexmedetomidine can be considered a particularly promising agent. Other anesthetic approaches will be required to test the efficacy of dexmedetomidine as an anti-inflammatory agent and to further clarify both safety and efficacy to dexmedetomidine use in patients undergoing extremely invasive surgeries, such as cardiopulmonary bypass.

2.1.6 References

1. Sander M, von Heymann C, von Dossow V, Spaethe C, Konertz, Uday Jain WF, Spies CD. Increased interleukin-6 after cardiac surgery predicts infection. Anesth Analg 2006; **102**: 1623- 9.
2. Murkin JM. Panvascular inflammation and mechanisms of injury in perioperative CNS outcomes. S Cardioth Vasc Anesth 2010; **14**: 190- 195.

3. Plomondon ME, Cleveland JC Jr, Ludwig ST, Grunwald, GK, Kiefe, CI, Grover, FL, Shroyer, AL. Off-pump coronary artery bypass is associated with improved risk-adjusted outcomes. *Ann Thorac Surg* 2001; **72**: 114- 119.
4. Wan S, LeClerc JL, Vincent JL. Inflammatory response to cardiopulmonary bypass: mechanisms involved and possible therapeutic strategies. *Chest* 1997; **112**: 676- 92.
5. Elahi MM, Khan JS, Matata BM. Deleterious effects of cardiopulmonary bypass in coronary artery surgery and scientific interpretation of off-pump's logic. *Acute Cardiac Care* 2006; **8**: 196- 209.
6. Matata BM, Sosnowski AW, Galinanes M. Off-pump bypass graft operation significantly reduces oxidative stress and inflammation. *Ann Thorac Surg* 2000; **69**: 785- 791.
7. Matata BM, Galinanes M. Cardiopulmonary bypass exacerbates oxidative stress but does not increase proinflammatory cytokine release in patients with diabetes compared with patients without diabetes: Regulatory effects of exogenous nitric oxide. *J Thorac Cardiovasc Surg* 2000; **120**: 1- 11.
8. Stover EP, Siegel LC, Parks R, Levin J, Body SC, Maddi R, D'Ambra MN, Mangano DT, Spiess BD. Variability in transfusion practice for coronary artery bypass surgery persists despite national consensus guidelines: A 24-institution study. Institutions of the Multicenter Study of Perioperative Ischemia Research Group. *Anesthesiology* 1998; **88**: 327- 33.
9. Stamou SC, Hill PC, Dangas G, Pfister AJ, Boyce SW, Dullum MK, Bafi AS, Corso PJ. Stroke after coronary artery bypass: Incidence, predictors, and clinical outcome. *Stroke* 2001; **32**: 1508- 13.
10. Mathew JP, Parks R, Savino JS, Friedman AS, Koch C, Mangano DT, Browner WS. Atrial fibrillation following coronary artery bypass graft surgery: Predictors, outcomes, and resource utilization. Multi Center Study of Perioperative Ischemia Research Group. *JAMA* 1996; **276**: 300- 6.
11. Rose EA. Off-pump coronary-artery bypass surgery. *N Engl J Med* 2003; **348**: 379- 80.
12. Ascione R, Caputo M, Angelini GD. Off-pump coronary artery bypass grafting: not a flash in the pan. *Ann Thorac Surg* 2003; **75**: 306- 13.
13. Hall JE, Uhrich TD, Barney JA, Arain SR, Ebert TJ. Sedative, amnestic, and analgesic properties of small-dose dexmedetomidine infusions. *Anesth Analg* 2000; **90**: 699- 705.
14. Gibbon JH Jr. Application of a mechanical heart and lung apparatus to cardiac surgery. *Minn Med* 1954; **37**: 171- 185.

15. Chen YF, Tsai WC, Lin CC, Tsai LY, Lee CS, Huang CH, Pan PC, Chen ML. Effect of leukocyte depletion on endothelial cell activation and trans endothelial migration of leukocytes during cardiopulmonary bypass. *Ann Thorac Surg* 2004; **78**: 634- 42.
16. Okamura T, Ishibashi N, Zurakowski D, Jonas RA. Cardiopulmonary bypass increases permeability of the blood-cerebrospinal fluid barrier. *Ann Thorac Surg* 2010; **89**: 187- 94.
17. Warren OJ, Smith AJ, Alexiou C, Rogers PLB, Jawad N, Vincent C, Darzi AW, Athanasiou T. The Inflammatory Response to Cardiopulmonary Bypass: Part 1-Mechanisms of Pathogenesis. *J Cardioth Vasc Anesth* 2009; **23**: 223- 231.
18. Wood KC, Hsu LL, Gladwin MT. Sickle cell disease vasculopathy: a state of nitric oxide resistance. *Free Radic Biol Med* 2008; **44**: 1506-1528.
19. Sanjay K, Bandyopadhyay U. Free heme toxicity and its detoxification systems in human. *Toxicology letters* 2005; **157**: 175- 188.
20. Schaer DJ, Buehler PW, Alayash AI, Belcher JD, Vercellotti GM. Hemolysis and free hemoglobin revisited: exploring hemoglobin and hemin scavengers as a novel class of therapeutic proteins. *Blood* 2013; **121**: 1276-84.
21. Vermeulen WIC, Hanssen SJ, Buurman WA, Jacobs MJ. Cardiovascular surgery and organ damage: time to reconsider the role of hemolysis. *J Thorac Cardiovasc Surg* 2011; **142**:1-11.
22. Haase M, Bellomo R, Haase-Fielitz A. Novel biomarkers, oxidative stress, and the role of labile iron toxicity in cardiopulmonary bypass-associated acute kidney injury. *J Am Coll Cardiol*. 2010; **55**: 2024-33.
23. Farina F, Davila DS, Rocha JBT, Aschner M Farina M, Avila DS, da Rocha JB, Aschner M. Metals, oxidative stress and neurodegeneration: A focus on iron, manganese and mercury. *Neurochem Int* 2012; **21**. doi:pii: S0197-0186(12)00396-8. 10.1016/j.neuint.2012.12.006. [Epub ahead of print] PubMed PMID: 23266600.
24. Haase M, Haase-Fielitz A, Bellomo R. Cardiopulmonary bypass, hemolysis, free iron, acute kidney injury and the impact of bicarbonate. *Contrib Nephrol* 2010; **165**: 28- 32.
25. Ritter C, Andrades ME, Reinke A, Menna-Barreto S, Moreira JCF, Dal-Pizzol F. Treatment with N-acetylcysteine plus deferoxamine protects rats against oxidative stress and improves survival in sepsis. *Crit Care Med* 2004, **32**: 342-349.
26. Vulcano M, Meiss RP, Ithuriz MA. Deferoxamine reduces tissue injury and lethality in LPS-treated mice. *Inter J Immunopharmacol* 2000, **22**: 635-644.
27. Vlahakos D, Arkadopoulos N, Kostopanagiotou G, Siasiakou S, Kaklamanis L, Degiannis D, Demonakou M, Smyrniotis V. Deferoxamine attenuates lipid peroxidation, blocks

- interleukin-6 production, ameliorates sepsis inflammatory response syndrome, and confers renoprotection after acute hepatic ischemia in pigs. *Artif Organs.* 2012; **36:** 400-408.
28. Edmunds LH: Extracorporeal perfusion. In: Edmunds LH editors. *Cardiac Surgery in the Adult.* New York, NY: McGraw-Hill; 1997; 255–294.
29. Rossi M, Sganga G, Mazzone M, Valenza, V, Guarneri, S, Portale, G, Carbone, L, Gatta, L, Pioli, C, Sanguinetti, M, Montaldo, M, Glieca, F, Fadda, G, Schiavello, R, Silveri, NG. Cardiopulmonary bypass in man: Role of the intestine in a self-limiting inflammatory response with demonstrable bacterial translocation. *Ann Thorac Surg* 2004; **77:** 612- 618.
30. Krishnadasam B, Griscavage-Ennis J, Aldea GS. Reperfusion injury during cardiopulmonary bypass. In: Matheis G, Moritz A, Scholz M editor. *Leukocyte Depletion in Cardiac Surgery and Cardiology.* Basel, Switzerland: Karger 2002; 54.
31. Aydin NB, Gercekoglu H, Aksu B, Ozkul, V, Sener, T, Kiygil, I, Turkoglu, T, Cimen, S, Babacan, F, Demirtas, M. Endotoxemia in coronary artery bypass surgery: A comparison of the off-pump technique and conventional cardiopulmonary bypass. *J Thorac Cardiovasc Surg* 2003; **125:** 843- 848.
32. Oudemans- van Straaten HM, Jansen PG, Hoek FJ, van Deventer, JH, Sturk, A, Stoutenbeek, CP, Tytgat1, NJ, Wildevuur, RH, Eysman, L. Intestinal permeability, circulating endotoxin, and postoperative systemic responses in cardiac surgery patients. *J Cardiothorac Vasc Anesth* 1996; **10:** 187- 194.
33. Koca U, Olguner ÇG, Ergür BU, Altekin E, Taşdögen A, Duru S, Girgin P, Gündüz K, Cilaker Micili S, Güzelda S, Akkuş M. The effects of dexmedetomidine on secondary acute lung and kidney injuries in the rat model of intra-abdominal sepsis. *The Scientific World Journal* 2013; art. no. 292687.
34. Vohra HA, Whistance R, Modi A, Ohri SK. The inflammatory response to miniaturised extracorporeal circulation: a review of the literature. *Mediators of Inflammation* Volume 2009; Article ID 707042, 7 pages doi:10.1155/2009/707042.
35. Menasché P. The inflammatory response to cardiopulmonary bypass and its impact on post-operative myocardial function. *Curr Opin Cardiol* 1995; **10:** 597- 604.
36. Journois D. Hemofiltration during cardiopulmonary bypass. *Kidney Int* 1998; **53:** 174- 7.
37. Clermont G, Vergely C, Jazayeri S, Lahet J-J, Goudeau J-J, Lecour S, David M, Rochette L, Girard C. Systemic free radical activation is a major event involved in myocardial oxidative stress related to cardiopulmonary bypass. *Anesthesiology* 2002; **96:** 80- 7.

38. Davies S, Duffy J, Wickens D, Underwood S, Hill A, Alladine F, Feneck R, Dormandy T, Walesby R: Time-course of free radical activity during coronary artery operations with cardiopulmonary bypass. *J Thorac Cardiovasc Surg* 1993; **105**: 979- 87.
39. Balmer P, Reihart W, Jordan P, Buhler E, Moser U, Gey F. Depletion of plasma vitamin C but not of vitamin E in response to cardiac operations. *J Thorac Cardiovasc Surg* 1994; **108**: 311- 20.
40. Toivonen HJ, Ahotupa M. Free radical reaction products and antioxidant capacity in arterial plasma during artery bypass grafting. *J Thorac Cardiovasc Surg* 1994; **108**: 140- 7.
41. Akila D'Souza B, Vishwanath P, D'Souza V. Oxidative injury and antioxidants in coronary artery bypass graft surgery: off-pump CABG significantly reduces oxidative stress. *Clin Chim Acta* 2007; **375**: 147- 152.
42. Milei J, Forcada P, Fraga CG, Grana DR, Iannelli G, Chiarello M, et al. Relationship between oxidative stress, lipid peroxidation, and ultrastructural damage in patients with coronary artery disease undergoing cardioplegic arrest/reperfusion. *Cardiovasc Res* 2007; **73**: 710- 719.
43. Hogue CW Jr, Palin CA, Arrowsmith JE. Cardiopulmonary bypass management and neurologic outcomes: an evidence based appraisal of current practices. *Anesth Analg* 2006; **102**: 21- 37.
44. Smith PLC. The systemic inflammatory response to cardiopulmonary bypass and the brain. *Perfusion* 1996; **11**: 196 – 9.
45. Murkin JM. Etiology and incidence of brain dysfunction after cardiac surgery. *J Cardiothorac Vasc Anesth* 1999; **13**: 12- 7.
46. De Lange F, Dieleman JM, Jungwirth B, Kalkman CJ. Effects of cardiopulmonary bypass on neurocognitive performance and cytokine release in old and diabetic rats. *Br J Anaesth* 2007; **99**: 177- 83.
47. Westaby S, Saatvedt K, White S, Katsumata T, van Oeveren W, Halligan PW. Is there a relationship between cognitive dysfunction and systemic inflammatory response after cardiopulmonary bypass? *Ann Thorac Surg* 2001; **667**- 72.
48. Parolari A, Camera M, Alamanni F, Natiato M, Polvani L, Agrifoglio M, Brambilla M, biancardi C, Mussoni L, Biglioli P, Tremoli E. Systemic inflammation after on-pump and off-pump coronary bypass surgery: a one-month follow-up. *Ann Thorac Surg* 2007; **84**: 823- 8.

49. Coimba C, Drake M, Boris-Moller F, et al: Long-lasting neuroprotective effect of postischemic hypothermia and treatment with an antiinflammatory/antipyretic drug. *Stroke* 1996; **27**: 1578- 1585.
50. Warner DS, Sheng H, Batinic-Haberle I. Oxidants, antioxidants and the ischemic brain. *J Exp Biol* 2004; **207**: 3221- 3231.
51. Van Harten AE, Scheeren TWL, Absalom AR . A review of postoperative cognitive dysfunction and neuroinflammation associated with cardiac surgery and anaesthesia *Anaesthesia* 2012; **66**: 280- 293.
52. Wahrborg P, Booth JE, Clayton T, Nugara F, Pepper J, Weintraub WS, Sigwart U, Stables RH. SOS Neuropsychology substudy investigators: neuropsychological outcome after percutaneous coronary intervention or coronary artery bypass grafting: results from the stent or surgery (SOS) trial. 2004; **110**: 3411- 7.
53. Jensen BO, Rasmussen LS, Steinbruchel DA. Cognitive outcomes in elderly high-risk patients 1 year after off-pump versus on-pump coronary artery bypass grafting. A randomized Trial. *Eur Jour Cardioth Surg* 2008; **34**: 1016- 102.
54. Van Dijk D, Spoor M, Hijman R, Nathoe HM, Borst C, Jansen EW, Grobbee DE, de Jaegere PP, Kalkman CJ. Octopus Study Group: Cognitive and cardiac outcomes 5 years after off-pump vs on-pump coronary artery bypass graft surgery. *JAMA* 2007; **297**: 701- 8.
55. Newman MF, Mathew JP, Grocott HP, Mackensen GB, Monk T, Welsh- Bohmer KA, Blumenthal JA, Laskowitz DT, Mark DB. Central nervous system injury associated with cardiac surgery *Lancet* 2006; **368**: 694- 703.
56. Ernest CS, Murphy BM, Worcester MU, Higgins RO, Elliott PC, Goble AJ, Le Grande MR, Genardini N, Tatoulis J. Cognitive function in candidates for coronary artery bypass graft surgery. *Ann Thorac Surg* 2006; **82**: 812- 8.
57. Selnes OA, Grega MA, Bailey MM, Pham LD, Zeger SL, Baumgartner WA, McKhann GM. Cognition 6 years after surgical or medical therapy for coronary artery disease. *Ann Neurol* 2008; **63**: 581- 90.
58. Van Dijk D, Moons KGM, Nathoe HM, van Aarnhem EHL, Borst C, Keizer AMA, Kalkman CJ, Hijman R. Cognitive outcomes five years after not undergoing coronary artery bypass graft surgery. *Ann Thorac Surg* 2008; **85**: 60- 4.
59. Selnes OA, Pham L, Zeger S, McKhann GM. Defining cognitive change after CABG: decline versus normal variability. *Ann Thorac Surg* 2006; **82**: 388- 90.
60. Van Dijk D, Kalkman CJ. Why are cerebral microemboli not associated with cognitive decline? *Anesth Analg* 2009; **109**: 1006- 8.

61. Jungwirth B, Eckel B, Blobner M, Kellermann K, Kochs EF, Mackensen GB. Impact of cardiopulmonary bypass on systemic interleukin-6 release, cerebral NFB expression and neurocognitive outcome in rats. *Anesth Analg* 2010; **110**: 312- 20.
62. Silbert BS, Scott DA, Ewered LA, Lewis MS, Maruff PT. Preexisting cognitive impairment in patients scheduled for elective coronary artery bypass graft surgery. *Anesth Analg* 2007; **104**: 1023- 8.
63. Winningham-Major F, Staeker JL, Barger SW, Coats S, Van Eldik LJ. Neurite extension and neuronal survival activities of recombinant S100 proteins that differ in the content and position of cysteine residues. *J Cell Biol* 1989; **109**: 3063- 3071.
64. Tramontina F, Leite MC, Goncalves D, Tramontina AC, Souza DF, Frizzo JK, Nardin P, Gottfried C, Wofchuk ST, Goncalves CA. High glutamate decreases S100B secretion by a mechanism dependent on the glutamate transporter. *Neurochem Res* 2006; **31**: 815- 820.
65. Donato R, Sorci G, Riuzzi F, Arcuri C, Bianchi R, Brozzi F, Tubaro C, Giambanco I. S100B's double life: Intracellular regulator and extracellular signal. *Biochim Biophys Acta* 2009; **1793**: 1008 -1022.
66. De Souza DF, Wartchow K, Hansen F, Lunardi P, Guerra, MC, Nardin P, Gonçalves,C-A. Interleukin-6-induced S100B secretion is inhibited by haloperidol and risperidone. *Prog NeuroPsychophar Biol Psych* 2013; **43**: 14- 22.
67. De Souza DF, Leite MC, Quincozes-Santos A, Nardin P, Tortorelli LS, Rigo MM, Gottfried C, Leal RB, Gonçalves C-A. S100B secretion is stimulated by IL-1 β in glial cultures and hippocampal slices of rats: Likely involvement of MAPK pathway. *J Neuroimmunology* 2009; **206**: 52- 57.
68. Beer C, Blacker D, Bynevelt M, Hankey GJ, Pudsey IB. Systemic markers of inflammation are independently associated with S100B concentration: results of an observational study in subjects with acute ischaemic stroke. *J Neuroinflammation* 2010; **29**: 7:71.
69. Reichenberg A, Yirmiya R, Schuld A et al. Cytokine-associated emotional and cognitive disturbances in humans. *Arch Gen Psychiatry* 2001; **58**: 445- 452.
70. Huttunen HJ, Kuja-Panula J, Sorci G, Agnelli AL, Donato R, Rauvala H: Coregulation of neurite out growth and cell survival by amphotericin and S100 proteins through receptor for advanced glycation end products (RAGE) activation. 2000; **275**: 40096-105.
71. Bierhaus A, Humpert PM, Morcos M, Wendt T, Chavakis T, Arnold B, Stern DM, Nawroth PP. Understanding rage, the receptor for advanced glycation end products. *Journ Mol Med* 2005; **83**: 876- 86.

72. Adami C, Sorci G, Blasi E, Agnelli AL, Bistoni F, Donato R: S100B expression in and effects on microglia. *Glia* 2001; **33**: 131- 142.
73. Hu J, Castets F, Guevara JL, Van Eldik, LJ. S100B stimulates inducible nitric oxide synthase activity and mRNA levels in rat cortical astrocytes. *J Biol Chem* 1996; **271**: 2543- 2547.
74. Petrova TV, Hu J, Van Eldik LJ. Modulation of glial activation by astrocyte-derived protein S100B: differential responses of astrocyte and microglial cultures. *Brain research* 2000; **853**: 74- 80.
75. Hu J, Ferreira A, Van Eldik LJ. S100B induces neuronal cell death through nitric oxide release from astrocytes. *J Neurochem* 1997; **69**: 2294 - 2301.
76. Kim SH, Smith CJ, Van Eldik LJ. Importance of mapk pathways for microglial pro-inflammatory cytokine IL-1 β production. *Neurobiol Aging* 2004; **25**: 431- 439.
77. Westaby S, Johnsson P, Parry AJ, et al. Serum S100 protein: a potential marker for cerebral events during cardiopulmonary bypass. *Ann Thorac Surg* 1996; **61**: 88- 92.
78. Blomquist S, Johnsson P, Luhrs C, et al. The appearance of S-100 protein in serum during and immediately after cardiopulmonary bypass surgery: a possible marker for cerebral injury. *J Cardiothorac Vasc Anesth* 1997; **11**: 699- 703.
79. Barbut D, Yao FS, Hager DN, Kavanagh P, Trifiletti RR, Gold JP. Comparison of transcranial doppler ultrasonography and transesophageal echocardiography to monitor emboli during coronary artery bypass surgery. *Stroke* 1996; **27**: 87- 90.
80. Jonsson H, Johnsson P, Alling C, Westaby S, Blomquist S. Significance of serum S100 release after coronary artery bypass grafting. *Ann Thorac Surg* 1998; **65**: 1639- 44.
81. Grocott HP, Croughwell ND, Amory DW, White WD, Kirchner JL, Newmann MF. Cerebral emboli and serum S100 during cardiac operations. *Ann Thorac Surg* 1998; **65**: 1645- 50.
82. Jonsson, A. As compared to neuron-specific enolase, S100B protein correlate more specific to the stroke volume and clinical outcome in ischemic stroke. *Kaka-Orinska* 2010.
83. Van Munster BC, Korevaar JC, Zwinderman AH et al. Time-course of cytokines during delirium in elderly patients with hip fractures. *J Am Geriatr Soc* 2008; **56**: 1704- 1709.
84. Worthmann H, Tryc AB, Goldbecker A, Ma YT, Tountopoulou A, Hahn A, Dengler R, Lichtenhagen R, Weissenborn K. The temporal profile of inflammatory markers and mediators in blood after acute ischemic stroke differs depending on stroke outcome. *Cerebrovasc Dis* 2010; **30**: 85- 92.

85. Thompson JS, Santini A, Peterson JF, Pun BT, Jackson JC, Ely EW. Intensive care unit delirium is an independent predictor of longer hospital stay: a prospective analysis of 261 non-ventilated patients. *Crit Care* 2005; **9**: R375- 81.
86. Pandharipande P, Cotton BA, Shintini A, et al. Prevalence and risk factors for development of delirium in surgical and trauma intensive care unit patients. *J Trauma* 2008; **65**: 34- 41.
87. Lundstrom M, Edlund A, Bucht G et al. Dementia after delirium in patients with femoral neck fractures. *J Am Geriatr Soc* 2003; **51**: 1002- 1006.
88. Perry VH. The influence of systemic inflammation on inflammation in the brain: implications for chronic neurodegenerative disease. *Brain Behav Immun* 2004; **18**: 407- 413.
89. Herrmann M, Ebert AD, Galazky I et al. Neurobehavioral outcome prediction after cardiac surgery: role of neurobiochemical markers of damage to neuronal and glial brain tissue. *Stroke* 2000; **31**: 645- 650.
90. Khan ZP, Ferguson CN, Jones RM. Alpha-2 and imidazoline receptor agonists. Their pharmacology and therapeutic role. *Anesthesia* 1999; **54**: 146- 65.
91. Carollo DS, Nossaman BD, Ramadhyani U. Dexmedetomidine: A review of clinical applications. *Curr Opin Anaesthesiol* 2008; **21**: 457- 61.
92. Candiotti KA, Bergese SD, Bokesch PM, Feldman MA, Wisemandle W, Bekker AY. Monitored anesthesia care with dexmedetomidine: A prospective, randomized, double-blind, multicenter trial. *Anesth Analg* 2010; **110**: 47- 56.
93. Eisenach, J, De Kock, M, Klinscha, W. α 2-Adrenergic agonists for regional anesthesia: a clinical review of clonidine (1984 - 1995). *Anesthesiology* 1996; **85**: 655- 674.
94. Maze M, Tranquilli W. Alpha-2 Adrenoceptor agonists: defining the role in clinical anesthesia. *Anesthesiology* 1991; **74**: 581- 605.
95. Fürst S. Transmitters involved in antinociception in the spinal cord. *Brain Research Bulletin* 1999; **48**: 129-141.
96. Loick HM, Schmidt C, Van Aken H, Junker R, Erren M, Berendes E, Rolf N, Meissner A, Schmid C, Scheld HH, Möllhoff T. High thoracic epidural anesthesia, but not clonidine, attenuates the perioperative stress response via sympatholysis and reduces the release of troponin T in patients undergoing coronary artery bypass grafting. *Anesth Analg* 1999; **88**: 701- 9.
97. Wallace A, Galindez D, Salahieh A, Layug, EL, Lazo EA, Haratonik KA, Boisvert DM, Kardatzke, D. Effect of clonidine on cardiovascular morbidity and mortality after noncardiac surgery. *Anesthesiology* 2004; **101**: 284- 293.

98. Stevens RD, Burri H, Tramèr MR. Efficacy of clonidine for prevention of perioperative myocardial ischemia: A critical appraisal and meta-analysis of the literature. *Anesth Analg* 2003; **97**: 623- 633.
99. Nishina K, Mikawa K, Uesugi T, Obara H, Maekawa M, Kamae I, Nishi N. Efficacy of clonidine for prevention of perioperative myocardial ischemia: a critical appraisal and meta-analysis of the literature. *Anesthesiology* 2002; **96**: 323- 329.
100. Von Dossow V, Baehr N, Moshirzadeh M, von Heymann C, Braun JP, Hein OV, Sander M, Wernecke KD, Konertz W, Spies CD. Clonidine Attenuated Early Proinflammatory Response in T-Cell Subsets After Cardiac Surgery. *Anesth Analg* 2006; **103**: 809- 814.
101. Ellis JE, Pedlow S. Premedication with clonidine does not attenuate suppression of certain lymphocyte subsets after surgery. *Anesth Analg* 1998; **87**: 1426 – 30.
102. Dorman T, Clarkson K, Rosenfeld B, et al. Effects of clonidine on prolonged sympathetic response. *Crit Care Med* 1997; **25**: 1147- 52.
103. Kulka P, Tryba M, Zenz M. Dose-response effects of intravenous clonidine on stress response during induction of anesthesia in coronary artery bypass grafting. *Anesth Analg* 1995; **80**: 263- 8.
104. Nader ND, Ignatowski TA, Kurek CJ, Knight PR, Spengler RN. Clonidine suppresses plasma and cerebrospinal fluid concentrations of TNF- α during the perioperative period. *Anesth Analg* 2001; **93**: 363-369.
105. Persec J, Persec Z, Husedzinovic I. Postoperative pain and systemic inflammatory stress response after preoperative analgesia with clonidine or levobupivacaine: A randomized controlled trial. *Wiener Klinische Wochenschrift* 2009; **121**: 558- 563.
106. Guerlach AT, Dasta JF. Dexmedetomidine: an updated review. *Ann Pharmacother* 2007; **41**: 245-252.
107. Ma D, Rajakumaraswamy N, Maze M: Alpha-2-Adrenoreceptor agonists: shedding light on neuroprotection? *Br Med Bull* 71:77-92, 2005
108. Fagerholm V, Scheinin M, Haaparanta M. Alpha-2A-adrenoceptor antagonism increases insulin secretion and synergistically augments the insulinotropic effect of glibenclamide in mice. *Br J Pharmacol* 2008; **154**: 1287- 1296.
109. Takada K, Clark DJ, Davies MF, et al. Meperidine exerts agonist activity at the alpha (2B)-adrenoceptor subtype. *Anesthesiology* 2002; **96**: 1420- 1426.
110. Fagerholm V, Rokka J, Nyman L, et al. Autoradiographic characterization of alpha (2C)-adrenoceptors in the human striatum. *Synapse* 2008; **62**: 508- 515.

111. Moura E, Afonso J, Hein L, Vieira-Coelho MA. Alpha2-adrenoceptor subtypes involved in the regulation of catecholamine release from the adrenal medulla of mice. *Br J Pharmacol* 2006; **149**: 1049- 58.
112. Takamatsu I, Iwase A, Ozaki M, et al. Dexmedetomidine reduces long-term potentiation in mouse hippocampus. *Anesthesiology* 2008; **108**: 94- 102.
113. Venn RM, Karol MD, Grounds RM. Pharmacokinetics of dexmedetomidine infusions for sedation of postoperative patients requiring intensive care. *Br J Anaesth* 2002; **88**: 669- 675.
114. De Wolf AM, Fragen RJ, Avram MJ, et al. The pharmacokinetics of dexmedetomidine in volunteers with severe renal impairment. *Anesth Analg* 2001; **93**: 1205- 1209.
115. Talke P, Richardson CA, Scheinin M, et al. Postoperative pharmacokinetics and sympatholytic effects of dexmedetomidine. *Anesth Analg* 1997; **85**: 1136- 1142.
116. Kallio A, Scheinin M, Koulu M, et al. Effects of dexmedetomidine, a selective alpha 2-adrenoceptor agonist, on hemodynamic control mechanisms. *Clin Pharmacol Ther* 1989; **46**: 33- 42.
117. Ebert TJ, Hall JE, Barney JA, et al. The effects of increasing plasma concentrations of dexmedetomidine in humans. *Anesthesiology* 2000; **93**: 382- 394.
118. Jorden VS, Pousman RM, Sanford MM, Thorborg PA, Hutchens MP. Dexmedetomidine overdose in the perioperative setting. *Ann Pharmacother* 2004; **38**: 803- 807.
119. Ramsay MA, Luterman DL. Dexmedetomidine as a total intravenous anesthetic agent. *Anesthesiology* 2004; **101**: 787- 790.
120. Sleigh J. All hands on dex. *Anaesthesia* 2012; **67**: 1193–1197.
121. Aho M, Erkola O, Kallio A, Scheinin H, Korttila K. Dexmedetomidine infusion for maintenance of anesthesia in patients undergoing abdominal hysterectomy. *Anesth Analg* 1992; **75**: 940- 6.
122. Ishii H, Kohno T, Yamakura T, Ikoma M, Baba H. Action of dexmedetomidine on the substantia gelatinosa neurons of the rat spinal cord. *Eur J Neurosci* 2008; **27**: 3182- 90.
123. Nelson LE, You T, Maze M, Franks NP. Evidence that the mechanism of hypnotic action in dexmedetomidine and muscimol-induced anesthesia converges on the endogenous sleep pathway. *Anesthesiology*. 2001; **95**: A1368.
124. Al-Metwalli RR, Mowafi HA, Ismail SA, et al. Effect of intra-articular dexmedetomidine on postoperative analgesia after arthroscopic knee surgery. *Br J Anaesth* 2008; **101**: 395- 399.

125. Yoshitomi T, Kohjitani A, Maeda S, et al. Dexmedetomidine enhances the local anesthetic action of lidocaine via an alpha-2A adrenoceptor. *Anesth Analg* 2008; **107**: 96-101.
126. Turkmen A, Altan A, Turgut N, et al. The correlation between the Richmond agitation-sedation scale and bispectral index during dexmedetomidine sedation. *Eur J Anaesthesiol* 2006; **23**: 300-304.
127. Aantaa R: Assessment of the sedative effects of dexmedetomidine, an alpha 2-adrenoceptor agonist, with analysis of saccadic eye movements. *Pharmacol Toxicol* 1991; **68**: 394- 398.
128. Elias WJ, Durieux ME, Huss D, Frysinger RC. Dexmedetomidine and arousal affect subthalamic neurons. *Mov Disord* 2008; **23**: 1317-1320.
129. Hsu YW, Cortinez LI, Robertson KM, et al. Dexmedetomidine pharmacodynamics: part I: crossover comparison of the respiratory effects of dexmedetomidine and remifentanil in healthy volunteers. *Anesthesiology* 2004; **101**: 1066- 1076.
130. Coull JT, Jones ME, Egan TD, et al. Attentional effects of noradrenaline vary with arousal level: selective activation of thalamic pulvinar in humans. *Neuroimage* 2004; **22**: 315-322.
131. Deutsch E, Tobias JD. Hemodynamic and respiratory changes following dexmedetomidine administration during general anesthesia: sevoflurane vs desflurane. *Paediatr Anaesth* 2007; **17**: 438- 444.
132. Koroglu A, Teksan H, Sagir O, et al. A comparison of the sedative, hemodynamic, and respiratory effects of dexmedetomidine and propofol in children undergoing magnetic resonance imaging. *Anesth Analg* 2006; **103**: 63- 67.
133. Arpino PA, Kalafatas K, Thompson BT. Feasibility of dexmedetomidine in facilitating extubation in the intensive care unit. *J Clin Pharm Ther* 2008; **33**: 25-30.
134. Corcoran TB, Engel A, Sakamoto H, O'Shea A, O'Callaghan-Enright S. Shorten The effects of propofol on neutrophil function, lipid peroxidation and inflammatory response during elective coronary artery bypass grafting in patients with impaired ventricular function *British Journal of Anaesthesia* 2006; **97**: 825- 31.
135. Arslan M, Çomu FM, Kuçuk A, Ozturk L, Yaylak F. Dexmedetomidine protects against lipid peroxidation and erythrocyte deformability alterations in experimental hepatic ischemia reperfusion injury. *Libyan J Med* 2012; **7**: 18185.
136. Rocha JBT, Bulow NMH, Correa EFM, Scholze C, Nogueira CW, Barbosa NBV. Dexmedetomidine protects blood d-aminolevulinate dehydratase from inactivation caused by

- hyperoxygenation in total intravenous anesthesia. Human and Experimental Toxicology 2010; **30**: 289- 295.
137. Straub RH, Herrmann M, Berkmler G, et al. Neuronal regulation of interleukin 6 secretion in murine spleen: adrenergic and opioidergic control. J Neurochem 1997; **68**: 1633- 9.
138. Szelenyi J, Kiss JP, Vizi ES. Differential involvement of sympathetic nervous system and immune system in the modulation of TNF-alpha production by alpha2- and beta-adrenoceptors in mice. J Neuroimmunol 2000; **103**: 34- 40.
139. Taniguchi T, Kidani Y, Kanakura H, et al. Effects of dexmedetomidine on mortality rate and inflammatory responses to endotoxin-induced shock in rats. Crit Care Med 2004; **32**: 1322- 6.
140. Hofer S, Steppan J, Wagner T, Funke B, Lichtenstern C, Martin E, Graf BM, Bierhaus A, Weigand MA. Central sympatholytics prolong survival in experimental sepsis. Crit Care Vol 13 No 1.
141. Can M, Gul S, Bektas S, Hanci V, Acikgos S. Effects of dexmedetomidine or methylprednisolone on inflammatory responses in spinal cord injury Acta Anaesth Scand 2009; **53**: 1068-1072.
142. Maze M, Virtanen R, Daunt D, Banks SJ, Stover EP, Feldman D. Effects of dexmedetomidine, a novel imidazole sedative-anesthetic agent, on adrenal steroidogenesis: in-vivo and in-vitro studies. Anesth Analg 1991; **73**: 204- 8.
143. Venn RM, Bryant A, Hall GM, Grounds RM. Effects of dexmedetomidine on adrenal cortical function, and the cardiovascular, endocrine, and inflammatory responses in post-operative patients needing sedation in the intensive care unit. Br J Anaesth 2001; **86**: 650-6.
144. Bulow NMH, Barbosa NBV, Rocha JBT. Opioid consumption in total anesthesia is reduced with dexmedetomidine: a comparative study with remifentanil in gynecologic videolaparoscopic surgery. J Clin Anesth 2007; **19**: 280- 5.
145. Pandharipande PP, Pun BT, Herr DL, et al. Effects of sedation with dexmedetomidine vs lorazepam on acute brain dysfunction in mechanically ventilated patients: the MENDS randomized controlled trial. JAMA 2007; **298**: 2644- 53.
146. Gerlach AT, Murphy CV, Dasta JF. An updated focused review of dexmedetomidine in adults. Ann Pharmacother 2009; **43**: 245-52.
147. Sulaiman S, Karthekeyan RB, Vakamundi M, Sundair AS, Ravullapalli H, Gandhan R. The effects of dexmedetomidine on attenuation of stress response to endotracheal intubation

- in patients undergoing elective off pump coronary artery bypass grafting. Ann Card Anaesth 2012; **15**: 39- 43.
148. Scheinin B, Lindgren L, Randell T, Scheinin H, Scheinin M. Dexmedetomidine attenuates sympathoadrenal responses to tracheal intubation and reduces the need for thiopentone and preoperative fentanyl. Br J Anaesth 1992; **68**: 126- 31.
149. Sukegawa S, Inoue M, Higuchi H, Tomoyasu Y, Maeda S, Miyawaki T. Locally Injected Dexmedetomidine Inhibits Carrageenin-induced Inflammatory Reactions in Injected Region. ASA annual meeting 2011; A1590.
150. Lai YC, Tsai PS, Huang CJ. Effects of dexmedetomidine on regulating endotoxin-induced up-regulation of inflammatory molecules in murine macrophages. J Surg Res 2009; **154**: 212- 219.
151. Yagmurdu H, Ozcan N, Dokumaci F, Kilinc K, Yilmaz F, Basar H. Dexmedetomidine reduces the ischemia-reperfusion injury markers during upper extremity surgery with tourniquet. J Hand Surg 2008; **33**: 941- 947.
152. Bekker A, Haile M, Kline R, Didehvar S, Babu R, Martiniuk F, Urban M. The effect of intraoperative infusion of dexmedetomidine on the quality of recovery after major spinal surgery. J Neurosurg Anesthesiol 2012; doi: 10.1097/ANA.0b013e31826318af.
153. Gu J, Chen J, Xia P, Tao G, Zhao H, Ma D. Dexmedetomidine attenuates remote lung injury induced by renal ischemia-reperfusion in mice. Acta Anaesth Scand 2011; **55**: 1272- 1278.
154. Wu X, Song X, Li N, Zhan L, Meng Q, Xia Z. Protective effects of dexmedetomidine on blunt chest trauma-induced pulmonary contusion in rats. J Trau Acute Care Surg 2013; **74**: 524-530.
155. Kang S-H, Kim Y-S, Hong T-H, Chae M-S, Cho M-L, Her Y-M, Lee J. Effects of dexmedetomidine on inflammatory responses in patients undergoing laparoscopic cholecystectomy. Acta Anaesth Scand 2013; **57**: 480-487.
156. Tasdogan M, Memis D, Sut N, Yuksel M. Results of a pilot study on the effects of propofol and dexmedetomidine on inflammatory responses and intraabdominal pressure in severe sepsis. J Clin Anesth 2009; **21**: 394-400.
157. Sanders RD, Ma D, Maze M. Anaesthesia induced neuroprotection. Best Pract Res Clin Anaesthesiol 2005; **19**: 461- 474.
158. Janke EL, Samra S. Dexmedetomidine and neuroprotection. Sem Anesth, Perioperative Medicine and Pain 2006; **25**: No 2.

159. Newman M. Open heart surgery and cognitive decline. Cleve Clin J Med 2007; **74**: S52-5.
160. Stroobant N, van Nooten G, De Bacquer D, Van Belleghem Y, Vingerhoets G. Neuropsychological functioning 3-5 years after coronary artery bypass grafting: does the pump make a difference? Eur J Cardiothorac Surg 2008; **34**: 396- 401.
161. Grocott HP. Genetic influences on cerebral outcome after cardiac surgery. Semin Cardiothorac Vasc Anesth 2006; **10**: 291- 296.
162. Taylor KM. Central nervous system effects of cardiopulmonary bypass. Ann Thorac Surg 1998; **66**: S20-4.
163. Hogue CW Jr, Hershey T, Dixon D, Fucetola R, Nassief A, Freedland KE, Thomas B, Schechtman K. Pre-existing cognitive impairment in women before cardiac surgery and its relationship with C-reactive protein concentrations. Anesth Analg 2006; **102**: 1602- 8.
164. Ho PM, Arciniegas DB, Grigsby J, McCarthy M, McDonald GO, Moritz TE, Shroyer AL, Sethi GK, Henderson WG, London MJ, VillaNueva CB, Grover FL, Hammermeister KE. Predictors of cognitive decline following coronary artery bypass graft surgery. Ann Thorac Surg 2004; **77**: 597- 603.
165. Koch CG, Li L, Shishehbor M, Nissen S, Sabik J, Starr NJ, Blackstone EH. Socioeconomic status and comorbidity as predictors of preoperative quality of life in cardiac surgery. J Thorac Cardiovasc Surg 2008; **136**: 665- 72.
166. Stroobant N, Vingerhoets G. Depression, anxiety, and neuropsychological performance in coronary artery bypass graft patients: a follow-up study. Psychosomatics 2008; **49**: 326- 31.
167. Ille R, Lahousen T, Schweiger S, Hofmann P, Kapfhammer HP. Influence of patient-related and surgery-related risk factors on cognitive performance, emotional state, and convalescence after cardiac surgery. Cardiovasc Revasc Med 2007; **8**: 166- 9.
168. Nussmeier NA, Searles BE. Inflammatory brain injury after cardiopulmonary bypass: is it real? Anesth Analg 2010; **110**: 288- 290.
169. Singh SP, Kapoor PM, Chowdhury U, Kiran U. Comparison of S100B levels, and their correlation with hemodynamic indices in patients undergoing coronary artery bypass grafting with three different anesthetic techniques. Ann Cardiac Anaesth 2011; **14**: 197- 202.
170. Kleindienst, A. et al. The neurotrophic protein S100B: value as a marker of brain damage and possible therapeutic implications. Prog. Brain Res 2007; **161**: 317- 325.
171. Riker RR, Shehabi Y, Bokesch PM, et al. Dexmedetomidine vs midazolam for sedation of critically ill patients: a randomized trial. JAMA 2009; **301**: 489- 99.

172. Maldonado JR, Wysong A, van der Starre PJA, Block T, Miller C, Reitz BA. Dexmedetomidine and the reduction of postoperative delirium after cardiac surgery. *Psychosomatics* 2009; **50**: 206- 17.
173. Ruokonen E, Parviainen I, Jakob SM, et al. Dexmedetomidine versus propofol/midazolam for long-term sedation during mechanical ventilation. *Intensive Care Med* 2009; **35**: 282- 90.
174. Mayo W, Lemaire V, Malaterre J, Rodriguez JJ, Cayre M, Stewart MG, Kharouby M, Rougon G, Le Moal M, Piazza PV, Abrous DN. Pregnanolone sulfate enhances neurogenesis and PSA-NCAM in young and aged hippocampus. *Neurobiol Aging* 2005; **26**: 103- 1411.
175. Keller EA, Zamparini A, Borodinsky LN, Gravielle MC, Fiszman ML. Role of allopregnanolone on cerebellar granule cells neurogenesis. *Brain Res Dev Brain Res* 2004; **153**: 13- 7.
176. Correa-Sales C, Rabin BC, Maze M. A hypnotic response to dexmedetomidine, an alpha-2 agonist, is mediated in the locus coeruleus in rats. *Anesthesiology* 1992; **76**: 948- 52.
177. Kulkarni VA, Jha S, Vaidya VA. Depletion of norepinephrine decreases the proliferation, but does not influence the survival and differentiation, of granule cell progenitors in the adult rat hippocampus. *Eur J Neurosci* 2002; **16**: 2008- 12.
178. Mohapel P, Leanza G, Kokaia M, Lindvall O. Forebrain acetylcholine regulates adult hippocampal neurogenesis and learning. *Neurobiol Aging* 2005; **26**: 939- 46.
179. Tung A, Herrera S, Fornal CA, Jacobs BL. The effect of prolonged anesthesia with isoflurane, propofol, dexmedetomidine, or ketamine on neural cell proliferation in the adult rat. *Anesth Analg* 2008; **106**: 1772- 7.
180. Hoffman WE, Kochs E. Dexmedetomidine improves neurologic outcome from incomplete ischemia in the rat. Reversal by the alpha2-adrenergic antagonist atipamezole. *Anesthesiology* 1991; **75**: 328- 332.
181. Hoffman WE, Baughman VL, Albrecht RF. Interaction of catecholamines and nitrous oxide ventilation during incomplete brain ischemia in rats. *Anesth Analg* 1993; **77**: 908-912.
182. Maier C, Steinberg GK, Sun GH, et al. Neuroprotection by the α -2- adrenoreceptor agonist dexmedetomidine in a focal model of cerebral ischemia. *Anesthesiology* 1993; **79**: 306- 312.
183. Kuhmonen J, Porkorney J, Miettinen R, et al. Neuroprotective effects of dexmedetomidine in the gerbil hippocampus after transient global ischemia. *Anesthesiology* 1997; **87**: 371- 377.

184. Laudenbach V, Mantz J, Lagercrantz H, et al. Effects of α -2-adrenoreceptor agonists on perinatal excitotoxic brain injury. *Anesthesiology* 2002; **96**: 134- 141.
185. Peng L, Yu AC, Fung KY, Prevot V, Hertz L. Alpha-adrenergic stimulation of ERK phosphorylation in astrocytes is alpha(2)-specific and may be mediated by transactivation. *Brain Res* 2003; **978**: 65- 71.
186. Li B, Du T, Li H, Gu L, Zhang H, Huang J, Hertz L, Peng L. Signalling pathways for transactivation by dexmedetomidine of epidermal growth factor receptors in astrocytes and its paracrine effect on neurons. *Br J Pharmacol* 2008; **154**:191-203.
187. Hertz L, Lovatt D, Goldman SA, Nedergaard M. Adrenoceptors in brain: cellular gene expression and effects on astrocytic metabolism and $[Ca(2+)]_i$. *Neurochem Int* 2010; **57**: 411-420.
188. Peng L, Du T, Xu J et al. Adrenergic and V1-ergic agonists/antagonists affecting recovery from Brain Trauma in the Lund project Act on astrocytes. *Curr Signal Transd Ther* 2012; **7**: 43- 55.
189. Liu L, Ji F, Liang J, He H, Fu Y, Cao M. Inhibition by dexmedetomidine of the activation of spinal dorsal horn glial and the intracellular ERK signaling pathway induced by nerve injury. *Brain Res* 2012; **1427**: 1- 9.
190. Xu B, Zhang WS, Yang JL, Lu N, Deng XM, Xu H, Zhang YQ. Evidence for suppression of spinal glial activation by dexmedetomidine in a rat model of monoarthritis. *Clin Exp Pharmacol Physiol* 2010; **37**: 158-166.
191. Chrysostomou C, Schmitt CG. Dexmedetomidine: sedation, analgesia and beyond. *Expert Opin Drug Metab Toxicol* 2008; **4**: 619-627.
192. Schoeler M, Loetscher PD, Rossaint R, Fahlenkamp AV, Eberhardt G, Rex S, Weis J, Coburn M. Dexmedetomidine is neuroprotective in an in vitro model for traumatic brain injury. *BMC Neurol* 2012; **12**: 20.
193. Zhang M, Shan X, Gu L, Hertz L, Peng L. Dexmedetomidine causes neuroprotection via astrocytic α 2-adrenergic receptor stimulation and HB-EGF release. *J Anesth Clin Science* 2013; doi:10.7243/2049-9752-2-6.
194. Hall ED, Vaishnav RA, Mustafa AG. Antioxidant therapies for traumatic brain injury. *Neurotherapeutics* 2010; **7**: 51-61.
195. Eriksson PS, Perfilieva E, Bjork-Eriksson T, Alborn AM, Nordborg C, Peterson DA, Gage FH. Neurogenesis in the adult human hippocampus. *Nat Med* 1998; **4**: 1313- 7.
196. Shors TJ, Miesegaes G, Beylin A, Zhao M, Rydel T, Gould E. Neurogenesis in the adult is involved in the formation of trace memories. *Nature* 2001; **410**: 372- 6.

197. Leuner B, Gould E, Shors TJ. Is there a link between adult neurogenesis and learning? *Hippocampus* 2006; **16**: 216- 24.
198. Henriksen L, Hjelms E, Lindeburgh T. Brain hyperperfusion during cardiac operations. Cerebral blood flow measured in man by intra-arterial injection of xenon 133: evidence suggestive of intraoperative microembolism. *J Thorac Cardiovasc Surg* 1983; **86**: 202- 208.
199. Govier AV, Reves JG, McKay RD, et al. Factors and their influence on regional cerebral blood flow during nonpulsatile cardiopulmonary bypass. *Ann Thorac Surg* 1984; **38**: 592- 600.
200. Murkin JM, Farrar JK, Tweed WA, McKenzie FN, Guiraudon G. Cerebral autoregulation and flow/metabolism coupling during cardiopulmonary bypass: the influence of PaCO₂. *Anesth Analg* 1987; **66**: 825- 832.
201. Ganushchak YM, Fransen EJ, Visser C, De Jong DS, Maessen JG. Neurological complications after coronary artery bypass grafting related to the performance of cardiopulmonary bypass. *Chest* 2004; **125**: 2196- 2205.
202. Hoffman WE, Kochs E, Werner C, Thomas C, Albrecht RF. Dexmedetomidine improves neurologic outcome from incomplete ischemia in the rat. Reversal by the alpha 2-adrenergic antagonist atipamezole. *Anesthesiology* 1991; **75**: 328- 332.
203. Dahmani S, Rouelle D, Gressens P, Mantz J. Characterization of the post-conditioning effect of dexmedetomidine in mouse organotypic hippocampal slice cultures exposed to oxygen and glucose deprivation. *Anesthesiology* 2010; **112**: 373- 383.
204. Engelhard K, Werner C, Eberspacher E, Bachl M, Blobner M, Hildt E, Hutzler P, Kochs E. The effect of the alpha 2-agonist dexmedetomidine and the N-methyl-D-aspartate antagonist S(+)-ketamine on the expression of apoptosis-regulating proteins after incomplete cerebral ischemia and reperfusion in rats. *Anesth Analg* 2003; **96**: 524- 531.
205. Sato K, Kimura T, Nishikawa T, Tobe Y, Masaki Y. Neuroprotective effects of a combination of dexmedetomidine and hypothermia after incomplete cerebral ischemia in rats. *Acta Anaesthesiol Scand* 2010; **54**: 377- 382
206. Eser O, Fidan H, Sahin O, Cosar M, Yaman M, Mollaoglu H, Songur A, Buyukbas S. The influence of dexmedetomidine on ischemic rat hippocampus. *Brain Res.* 2008; **1218**: 250- 6184.
207. Afonso J, Reis F. Dexmedetomidine: current role in anesthesia and intensive care. *Rev Bras Anestesiol* 2012; **62**: 118- 33.
208. Dahmani S, Paris A, Jannier V, Hein L, Rouelle D, Scholz J, Gressens P, Mantz J. Dexmedetomidine increases hippocampal phosphorylated extracellular signal-regulated

- protein kinase 1 and 2 content by an alpha 2-adrenoceptor-independent mechanism: evidence for the involvement of imidazoline II receptors. *Anesthesiology* 2008; **108**: 457- 466.
209. Shen J, Wu Y, Xu JY, Zhang J, Sinclair SH, Yanoff M, Xu G, Li W, Xu GT. ERK and Akt-dependent neuroprotection by erythropoietin (EPO) against glyoxal-AGEs via modulation of Bcl-xL, Bax, and BAD. *Invest Ophthalmol Vis Sci* 2010; **51**: 35- 46.
210. Kocoglu H, Karaaslan K, Gonca E, Bozdogan O, Gulcu N. Preconditioning effects of dexmedetomidine on myocardial ischemia/reperfusion injury in rats. *Curr Ther Res Clin Exp* 2008; **69**: 150- 8.
211. Kocoglu H, Ozturk H, Ozturk H, Yilmaz F, Gulcu N. Effect of dexmedetomidine on ischemia-reperfusion injury in rat kidney: a histopathologic study. *Ren Fail* 2009; **31**: 70- 4.
212. Bloor BC, Ward DS, Belleville JP, Maze M. Effects of intravenous dexmedetomidine in humans: II. hemodynamic changes. *Anesthesiology* 1992; **77**: 1134- 42.
213. Dahmani S, Rouelle D, Gressens P, et al. Effects of dexmedetomidine on hippocampal focal adhesion kinase tyrosine phosphorylation in physiologic and ischemic conditions. *Anesthesiology* 2005; **103**: 969- 977.
214. Prielipp RC, Wall MH, Tobin JR, et al. Dexmedetomidine-induced sedation in volunteers decreases regional and global cerebral blood flow. *Anesth Analg* 2002; **95**: 1052- 1059.
215. Zornow MH, Maze M, Dyck JB, et al. Dexmedetomidine decreases cerebral blood flow velocity in humans. *J Cereb Blood Flow Metab* 1993; **13**: 350- 353.
216. Drummond JC, Dao AV, Roth DM, et al. Effect of dexmedetomidine on cerebral blood flow velocity, cerebral metabolic rate, and carbon dioxide response in normal humans. *Anesthesiology* 2008; **108**: 225- 232.
217. Schoeler M, Loetscher PD, Rossaint R, Fahlenkamp AV, Eberhardt G, Rex S, Weis J, Coburn M. Dexmedetomidine is neuroprotective in an in vitro model for traumatic brain injury. *BMC Neurology* 2012; **12**: 20.
218. Sulemanji DS, Dönmez A, Aldemir D, Sezgin A, Türkoglu S. Dexmedetomidine during coronary artery bypass grafting surgery: is it neuroprotective? – A preliminary study. *Acta Anaesth Scand* 2007; **51**: 1093- 1098.
219. Bell MT, Ferenc Puskas F, Smith PD, Agoston VA, Fullerton DA, Meng X, Weyant MJ, Reece TB. Attenuation of spinal cord ischemia-reperfusion injury by specific α -2a receptor activation with dexmedetomidine. *J Vasc Surg* 2012; **56**: 1398-1402.

220. Gu J, Sun P, Zhao H, Watts HR, Sanders RD, Terrando N, Xia P, Maze M, Ma D. Dexmedetomidine provides renoprotection against ischemia-reperfusion injury in mice. *Critical Care* 2011; **15**: R153.
221. Tufanogullari B, White PF, Peixoto MP, Kianpour D, Lacour T, Griffin J, et al. Dexmedetomidine infusion during laparoscopic bariatric surgery: the effect on recovery outcome variables. *Anesth Analg* 2008; **106**: 1741- 8.
222. Jalowiecki P, Rudner R, Gonciarz M, et al. Sole use of dexmedetomidine has limited utility for conscious sedation during outpatient colonoscopy. *Anesthesiology* 2005; **103**: 269- 273.
223. Muntazar M, Kumar FC. Cardiac arrest, a preventable yet a possible risk of dexmedetomidine: fact or fiction? *Anesthesiology* 2004; **101**: 1478- 1479.
224. Shah AN, Koneru J, Nicoara A, et al. Dexmedetomidine related cardiac arrest in a patient with permanent pacemaker; a cautionary tale. *Pacing Clin Electrophysiol* 2007; **30**: 1158-1160.
225. Herr DL, Sum-Ping ST, England M. ICU sedation after coronary artery bypass graft surgery: dexmedetomidine-based versus propofol-based sedation regimens. *J Cardiothorac Vasc Anesth* 2003; **17**: 576- 584.
226. Jaionen J, Hynynen M, Kuitunen A, Heikkila H, Perttila J, Salmenpera M, Valtonen M, Aantaa R, Kallio A. Dexmedetomidine as an anesthetic adjunct in coronary artery bypass grafting. *Anesthesiology* 1997; **86**: 331-45.
227. Karakaya KH, Sahin N, Temel Y, Aydogdu TT. Hemodynamics in coronary artery bypass surgery: effects of intraoperative dexmedetomidine administration. *Anaesthetist* 2011; PMID: 21271232.
228. Guo H, Takahashi S, Cho S, Hara T, Tomiyasu S, Sumikawa K. The effects of dexmedetomidine on left ventricular function during hypoxia and reoxygenation in isolated rat hearts. *Anesth Analg* 2005; **100**: 629- 35.
229. Chen H, Higashino H, Maeda K, et al. Reduction of cardiac norepinephrine improves post-ischemic heart function in strokeprone spontaneously hypertensive rats. *J Cardiovasc Pharmacol* 2001; **38**: 821- 32.
230. Mertes PM, Carteaux JP, Jaboin Y, et al. Estimation of myocardial interstitial norepinephrine release after brain death using cardiac microdialysis. *Transplantation* 1994; **57**: 371-7.
231. Bybee KA, Prasad A. Stress-related cardiomyopathy syndromes. *Circulation* 2008; **118**: 397- 409.

232. Hachinski VC, Smith KE, Silver MD, Gibson CJ, Ciriello J. Acute myocardial and plasma catecholamine changes in experimental stroke. *Stroke* 1986; **17**: 387- 90.
233. Lee VH, Oh JK, Mulvagh SL, Wijdicks EF. Mechanisms in neurogenic stress cardiomyopathy after aneurysmal subarachnoid hemorrhage. *Neurocrit Care* 2006; **5**: 243- 9.
234. Ebert TJ, Hall JE, Barney JA, et al. The effects of increasing plasma concentrations of dexmedetomidine in humans. *Anesthesiology* 2000; **93**: 382- 94.
235. Willigers HM, Prinzen FW, Roekaerts PM, et al. Dexmedetomidine decreases perioperative myocardial lactate release in dogs. *Anesth Analg* 2003; **96**: 657- 64.
236. Karmazyn M, Beamish R, Dhalla N. Involvement of calcium in coronary vasoconstriction due to prolonged hypoxia. *Am Heart J* 1984; **107**: 293- 7.
237. Pinsky D, Oz M, Liao H, et al. Restoration of the cAMP second messenger pathway enhances cardiac preservation for transplantation in a heterotopic rat model. *J Clin Invest* 1993; **92**: 2994 – 3002.
238. Kitakaze M, Hori M, Gotoh K, et al. Beneficial effects of alpha-2 adrenoceptor activity on ischemic myocardium during coronary hypoperfusion in dogs. *Cir Res* 1989; **65**: 1632- 45.
239. Oliver MF, Goldman L, Julian DG, et al. Effect of mivazerol on perioperative cardiac complications during non-cardiac surgery in patients with coronary heart disease: the European Mivazerol Trial (EMIT). *Anesthesiology* 1999; **91**: 951- 961.
240. Wijeysundera DN, Naik JS, Beattie WS. Alpha-2 adrenergic agonists to prevent perioperative cardiovascular complications: a meta-analysis. *Am J Med* 2003; **114**: 742- 752.
241. Rouch AJ, Kudo LH, Hebert C. Dexmedetomidine inhibits osmotic water permeability in the rat cortical collecting duct. *J Pharmacol Exp Ther* 1997; **281**: 62- 69.
242. Billings T, Chen SW, Kim M, et al: Alpha2-Adrenergic agonists protect against radiocontrast-induced nephropathy in mice. *Am J Physiol Renal Physiol* 2008; **295**: 741- 748.
243. Taoda M, Adachi YU, Uchihashi Y, et al. Effect of dexmedetomidine on the release of [³H]-noradrenaline from rat kidney cortex slices: characterization of alpha₂-adrenoceptor. *Neurochem Int* 2001; **38**: 317- 322.
244. Venn M, Newman J, Grounds M. A phase II study to evaluate the efficacy of dexmedetomidine for sedation in the medical intensive care unit. *Intensive Care Med* 2003; **29**: 201- 207.
245. Reid K, Hayashi Y, Guo TZ, et al. Chronic administration of an alpha-2 adrenergic agonist desensitizes rats to the anesthetic effects of dexmedetomidine. *Pharmacol Biochem Behav* 1994; **47**: 171- 175.

246. Doufas AG, Lin CM, Suleman MI, Liem EB, Lenhardt R, Morioka N, Akça O, Shah YM, Bjorksten A, Sessler DI: Dexmedetomidine and meperidine additively reduce the shivering threshold in humans. *Stroke* 2003; **34**: 1218- 1223.
247. Elvan EG, Oç B, Uzun S, Karabulut E, Coskun F, Aypar U. Dexmedetomidine and post-operative shivering in patients undergoing elective abdominal hysterectomy. *Eur Journ Anaesth* 2008; **25**: 357- 364.
248. Maccioli GA. Dexmedetomidine to facilitate drug withdrawal. *Anesthesiology* 2003; **98**: 575- 577.
249. Multz AS. Prolonged dexmedetomidine infusion as an adjunct in treating sedation-induced withdrawal. *Anesth Analg* 2003; **96**: 1054- 1055.
250. Baddigam K, Russo P, Russo J, Tobias JD. Dexmedetomidine in the treatment of withdrawal syndromes in cardiothoracic surgery patients. *J Intensive Care Med* 2005; **20**: 118- 123.
251. Rovasalo A, Tohmo H, Aantaa R, Kettunen E, Palojoki R. Dexmedetomidine as an adjuvant in the treatment of alcohol withdrawal delirium: a case report. *Gen Hosp Psychiatry* 2006; **28**: 362- 363.
252. Darrouj J, Puri N, Prince E, Lomonaco A, Spevetz A, Gerber DR. Dexmedetomidine infusion as adjunctive therapy to benzodiazepines for acute alcohol withdrawal. *2008 Ann Pharmacother* 2008; **42**: 1703- 1705.

Legends of Figures

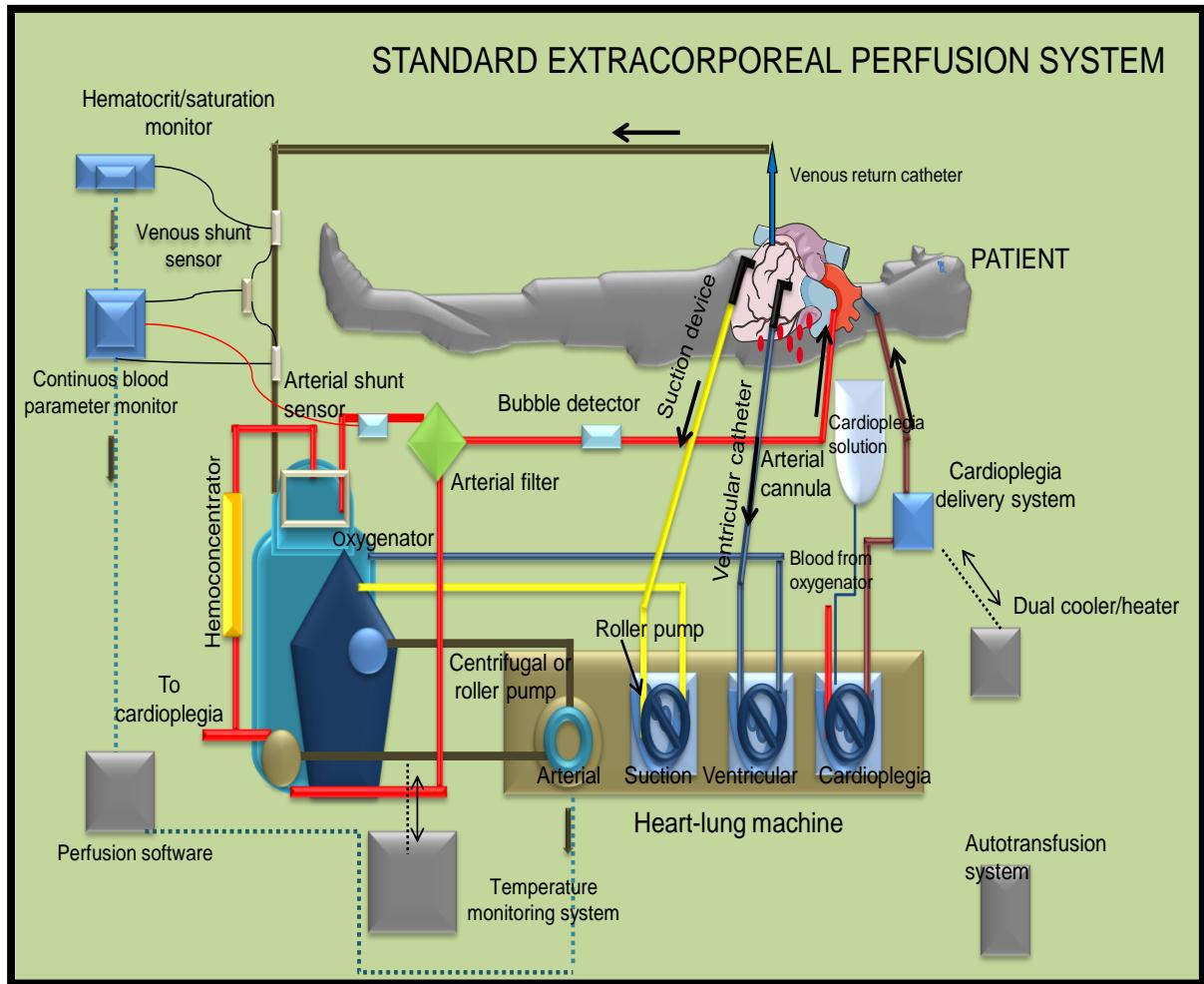


Figure 1. Standard Extracorporeal Circulation System. At coronary artery bypass grafting surgery, blood is diverted by pumps of heart and lungs, through an oxygenator, and circulating at an extensive tubulation device to permit the oxygenation of tissue during the ischemia period.

Legends of Figures

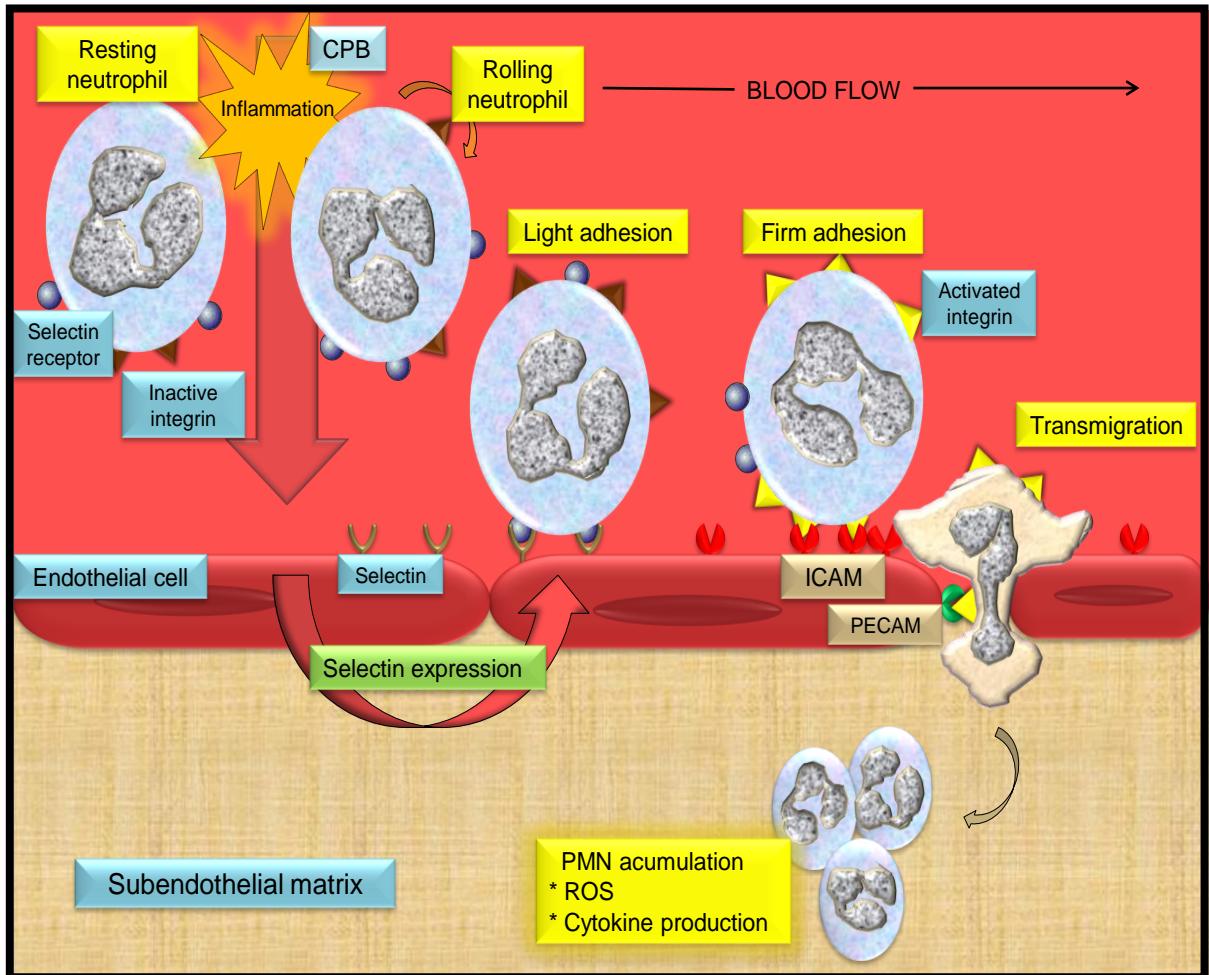


Figure 2. Cardiopulmonary Bypass neutrophil activation. At neutrophil activation by inflammatory mediators a neutrophil rolling phase occur, with posterior endothelial adhesivity, initially light, thus firm and culminating with the neutrophil endothelial transmigration. It leads to neutrophil accumulation, reactive oxygen species (ROS) production and cytokine release, maintaining the vicious circles. CPB: Cardiopulmonary Bypass; PMN: polymorphonuclears; ROS: Reactive Oxygen Species; ICAM: Intercellular adhesion molecule PECAM: Platelet/endothelial cell adhesion molecule.

Legends of Figures

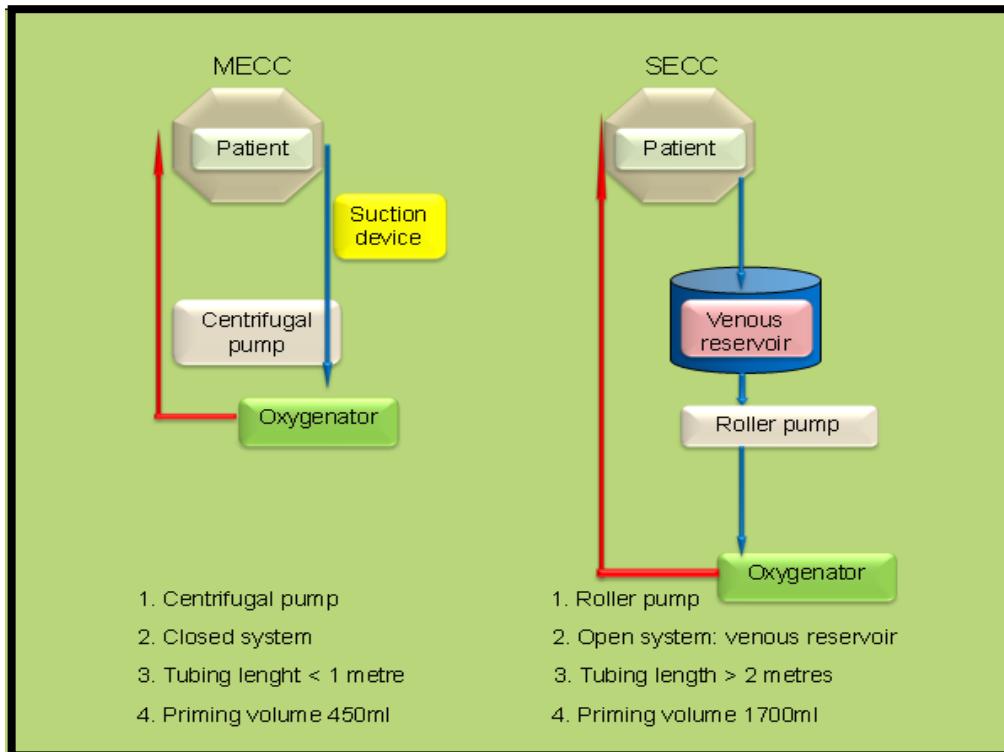


Figure 3. Mini-Extracorporeal Circulation System compared to Standard Extracorporeal Circulation System. In MECC system they exist various advantageous, such a shorter tubular circuit, only a centrifugal pump use, a closed system when blood no contact air and a smaller priming volume resulting in lesser hemodilution as compared to SECC. MECC: mini-extracorporeal circulation; SECC: standard extracorporeal circulation.

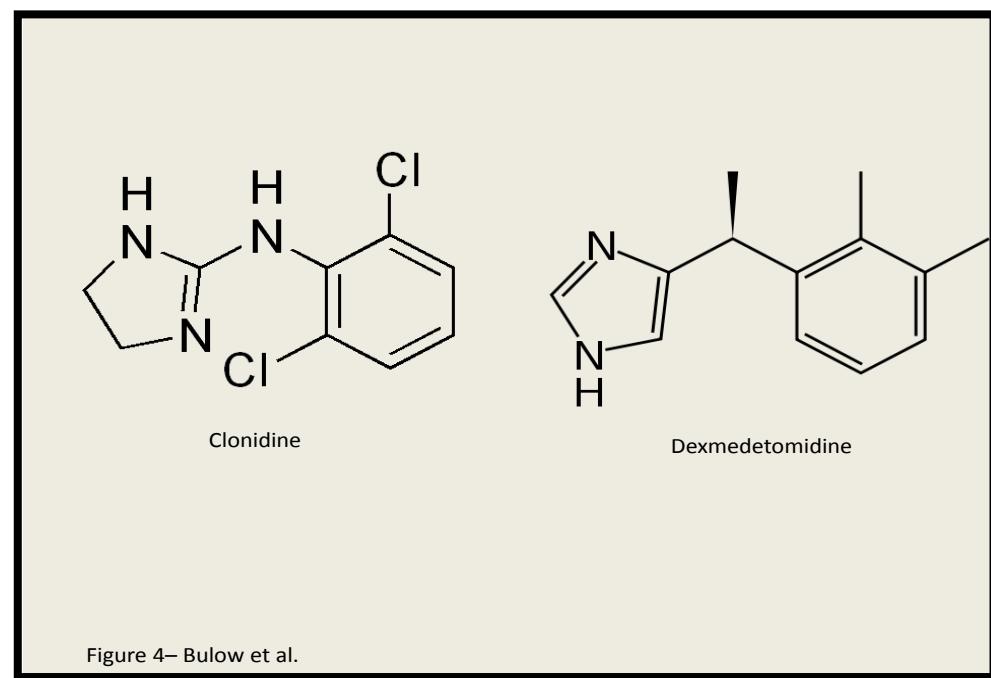


Figura 4. Dexmedetomidine and Clonidine structural formulae.

Legends of Figures

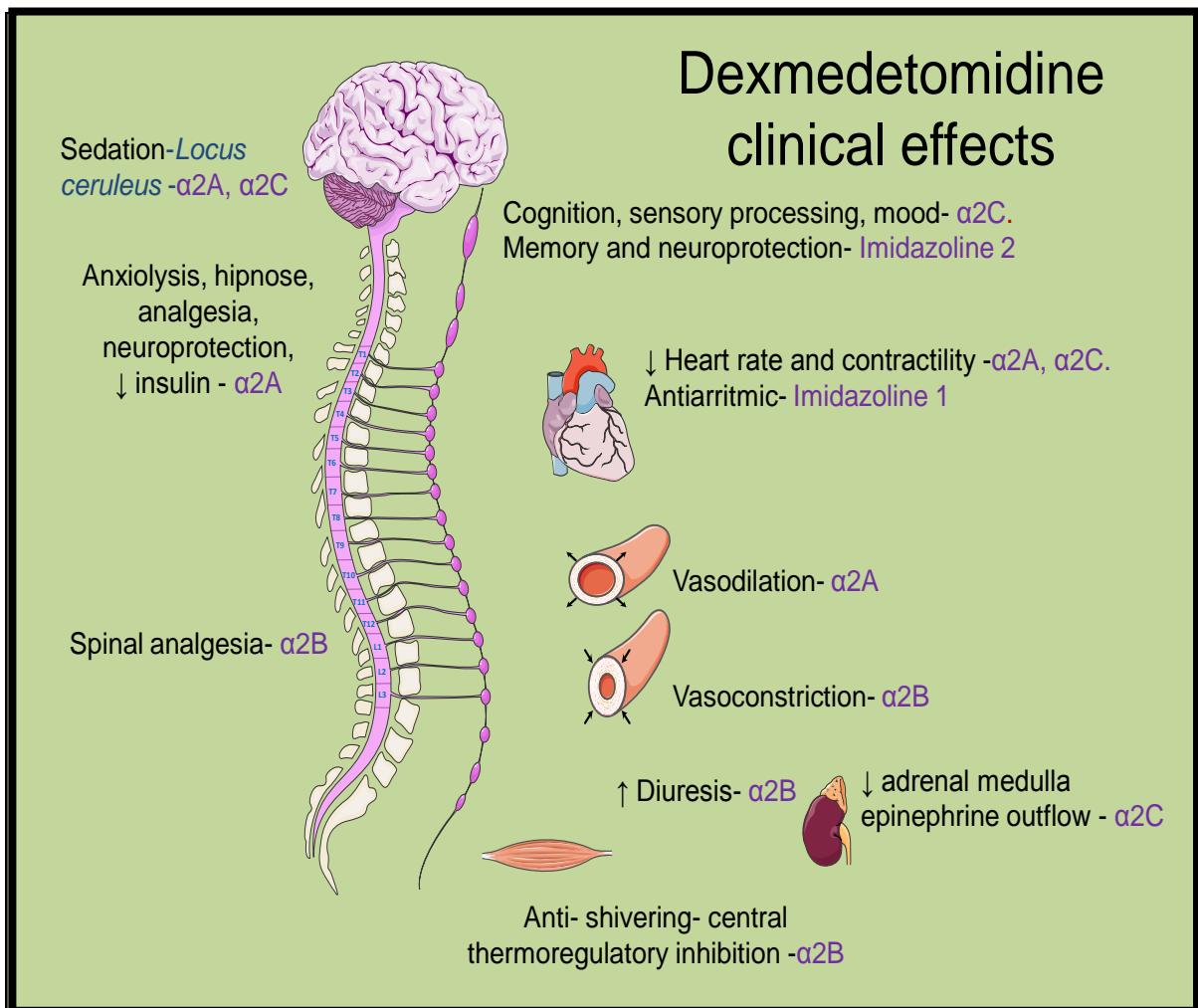


Figure 5. Dexmedetomidine clinical effects mediated via activation of (α)-2-adrenergic and imidazoline receptors. Through the presynaptic α -2A-adrenoreceptors agonism, dexmedetomidine induce sedation, anxiolysis, hipnose, analgesia, neuroprotection, reduce insulin release, reduce heart rate and myocardic contractility and lead to vasodilation. Also by presynaptic agonistic effect on α -2C-adrenoreceptors induce sedation, mood and cognition modulation, sensorial processing and reduction of adrenal medulla epinephrine. By the postsynaptic α -2B-adrenoreceptors agonism, dexmedetomidine cause analgesia at spinal level, vasoconstriction (with high bolus doses), improve of diuresis and central inhibition of shivering. Dexmedetomidine act also at imidazoline receptors, with a neuroprotection mechanism (imidazoline-2) and with antiarrhythmic effect (imidazoline-1).

Legends of Figures

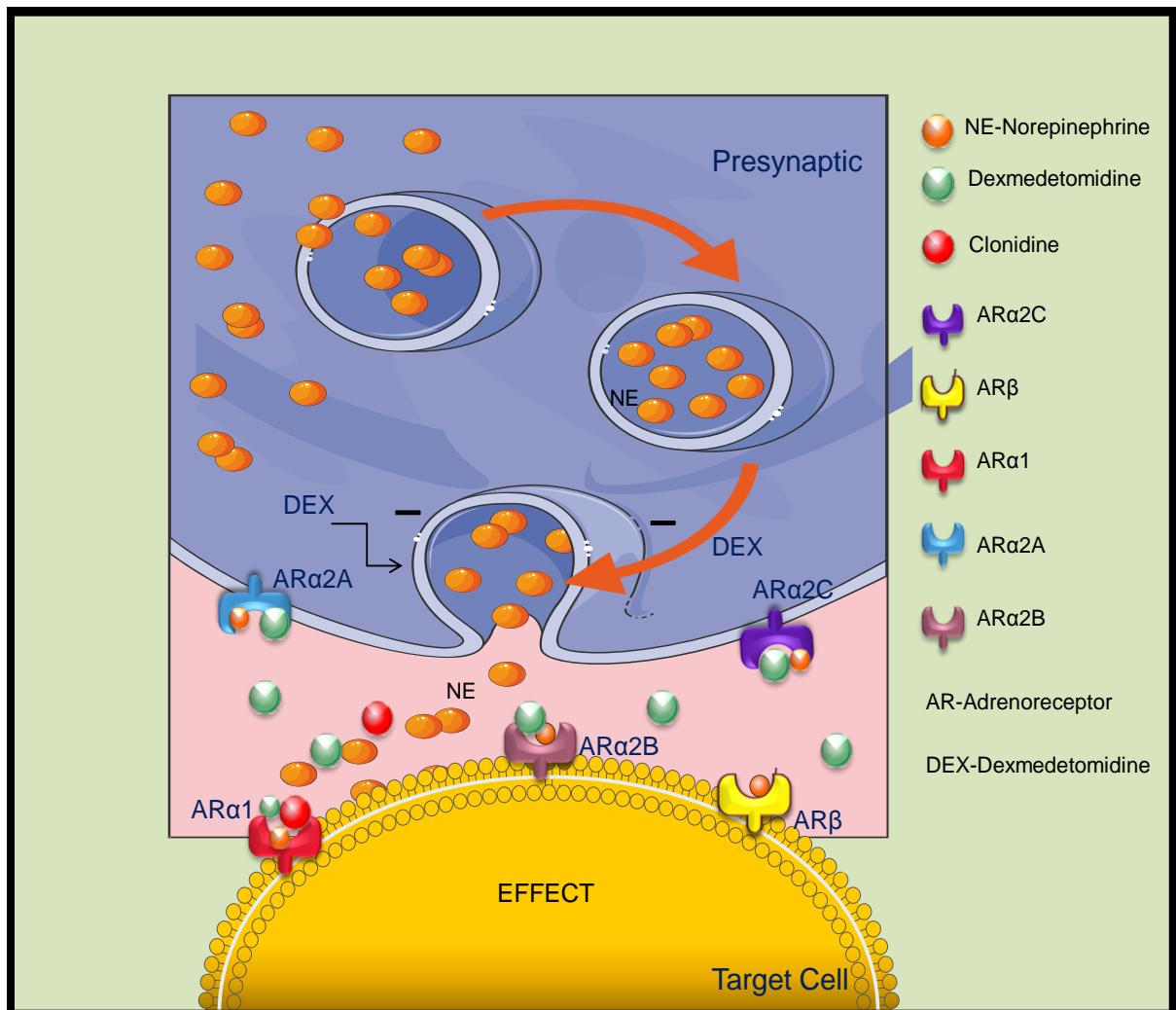


Figure 6. Dexmedetomidine can exert its effects via activation of three (α -2-adrenoreceptor subtypes. A subclass of (α -2-adrenoreceptor located presynaptically ((α -2A and (α -2C) regulated the release of neurotransmitter (norepinephrine). Located postsynaptically, a subclass of (α -2B-adrenoreceptor, and also the (α -1-adrenoceptor, while dexmedetomidine is not (α -2-adrenoceptor selective. The (α -2-adrenoceptors could exist also extrasynaptically.

Legends of Figures

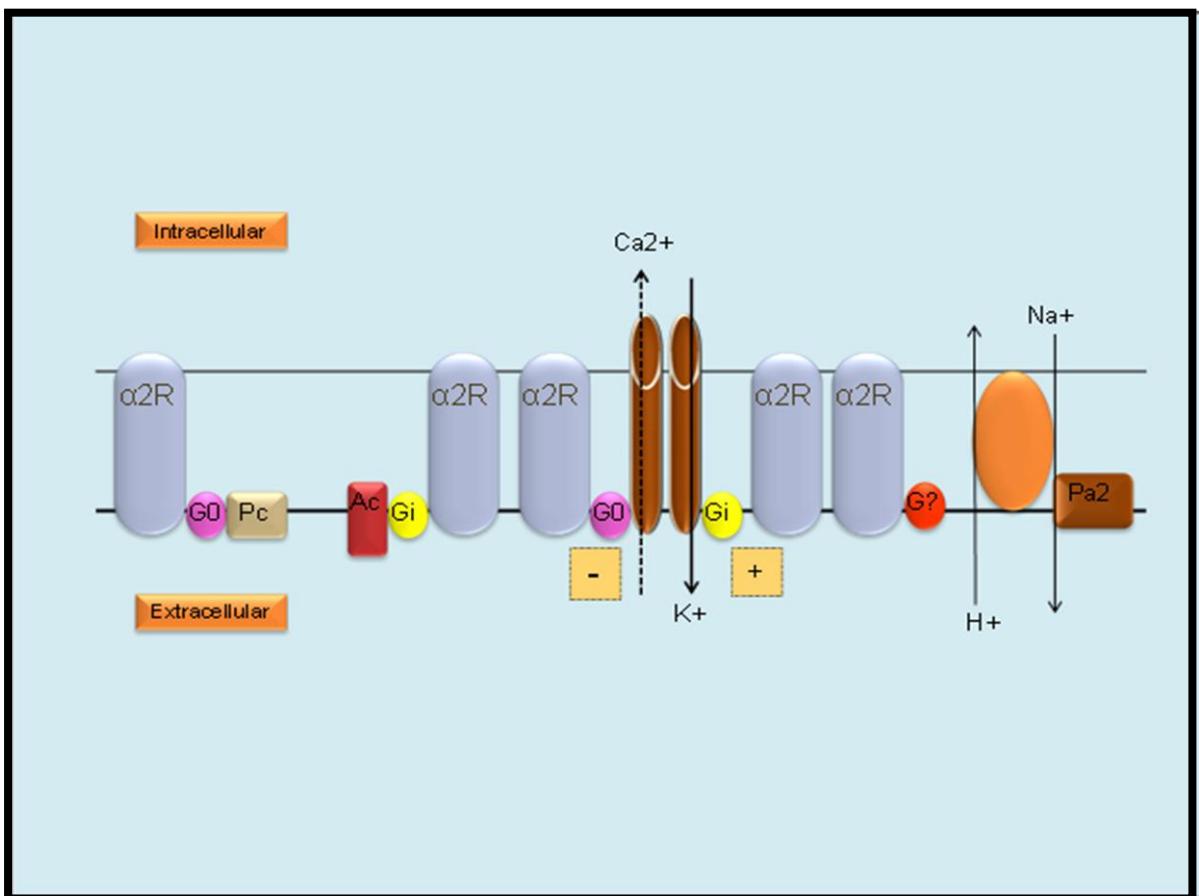


Figure 7. Putative Intracellular Mechanisms Involved in the (α) -2-adrenoreceptors Activation. The (α) -2-adrenoreceptor subtypes are transmembrane receptors that can be coupled to different classes of G-protein. The activation of (α) -2-adrenoreceptor (α_2 R) inhibits adenyl cyclase (Ac) via activation of receptor-coupled Gi protein. This causes outward opening of the K⁺ channel via Gi protein, which results in cell hyperpolarization. The coupling of adrenoreceptors to G0 can either inhibit Ca²⁺ translocation or modulate phospholipase C (Pc). The coupling with an undertermined class of G protein (G?) stimulates an exchange of H⁺ and Na⁺ (Modified from Ma D, Rajakumaraswamy N, Maze M: 2-Adrenoreceptor agonists: shedding light on neuroprotection? Br Med Bull 71:77-92, 2005¹⁰⁷).

Legends of Figures

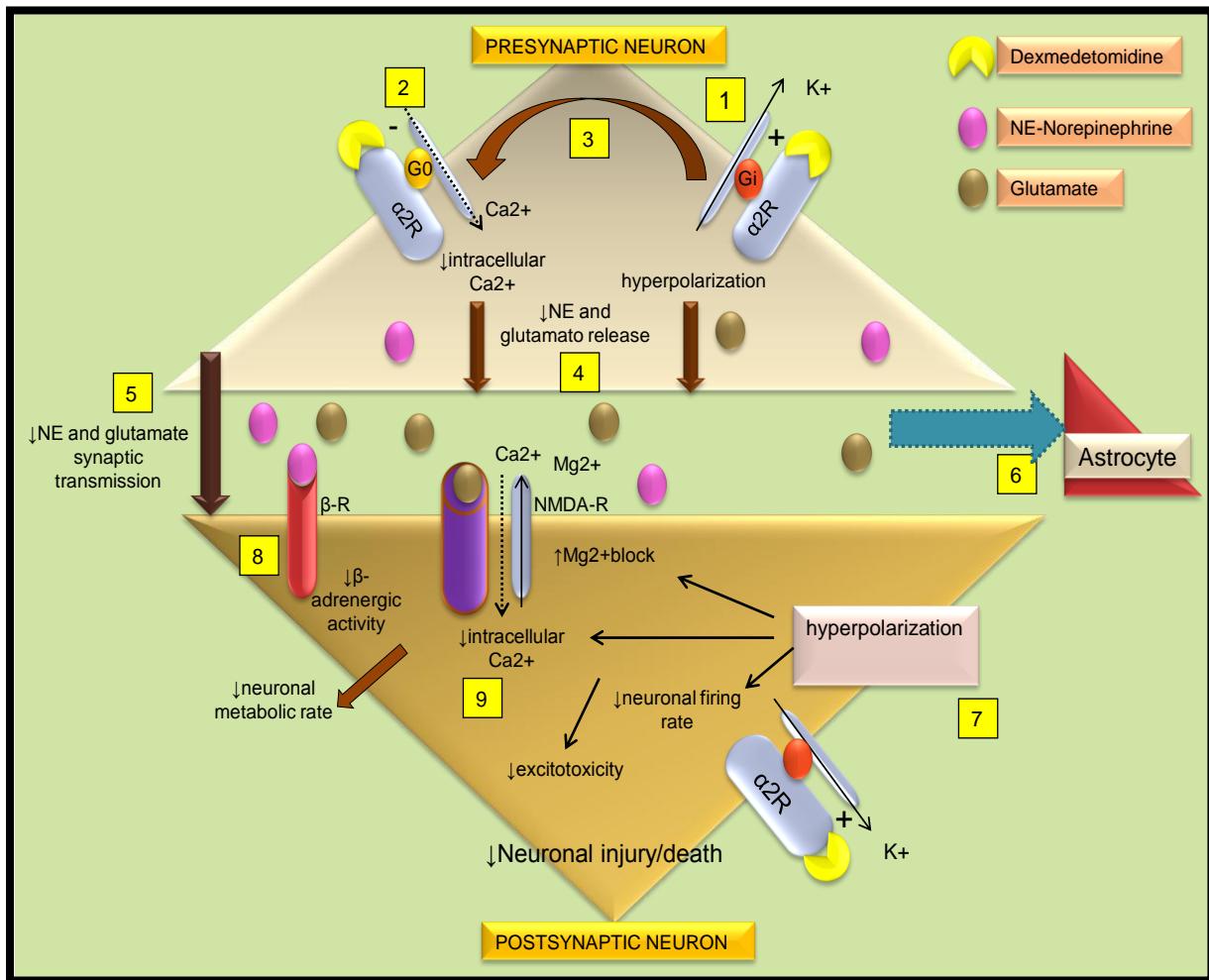
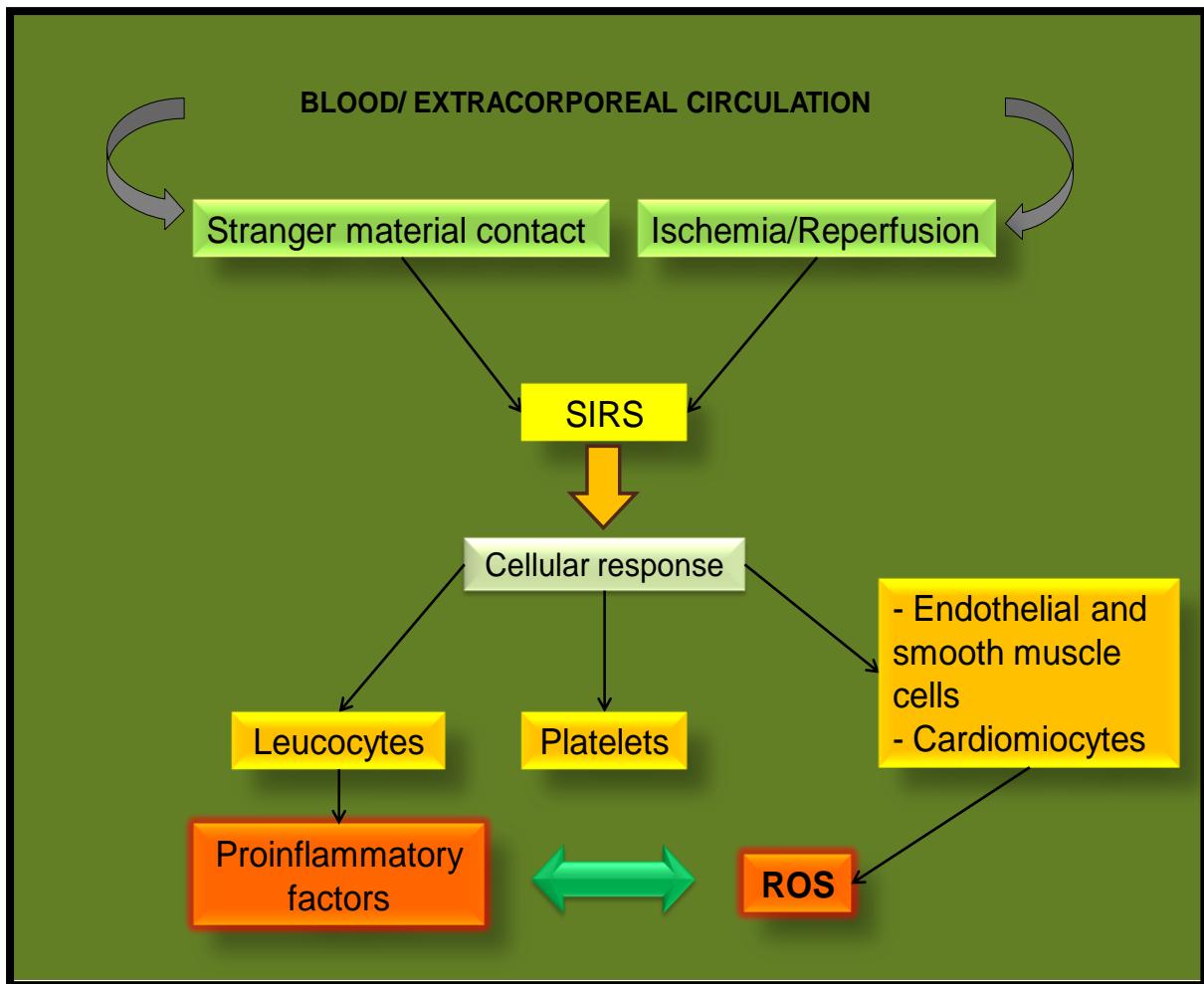


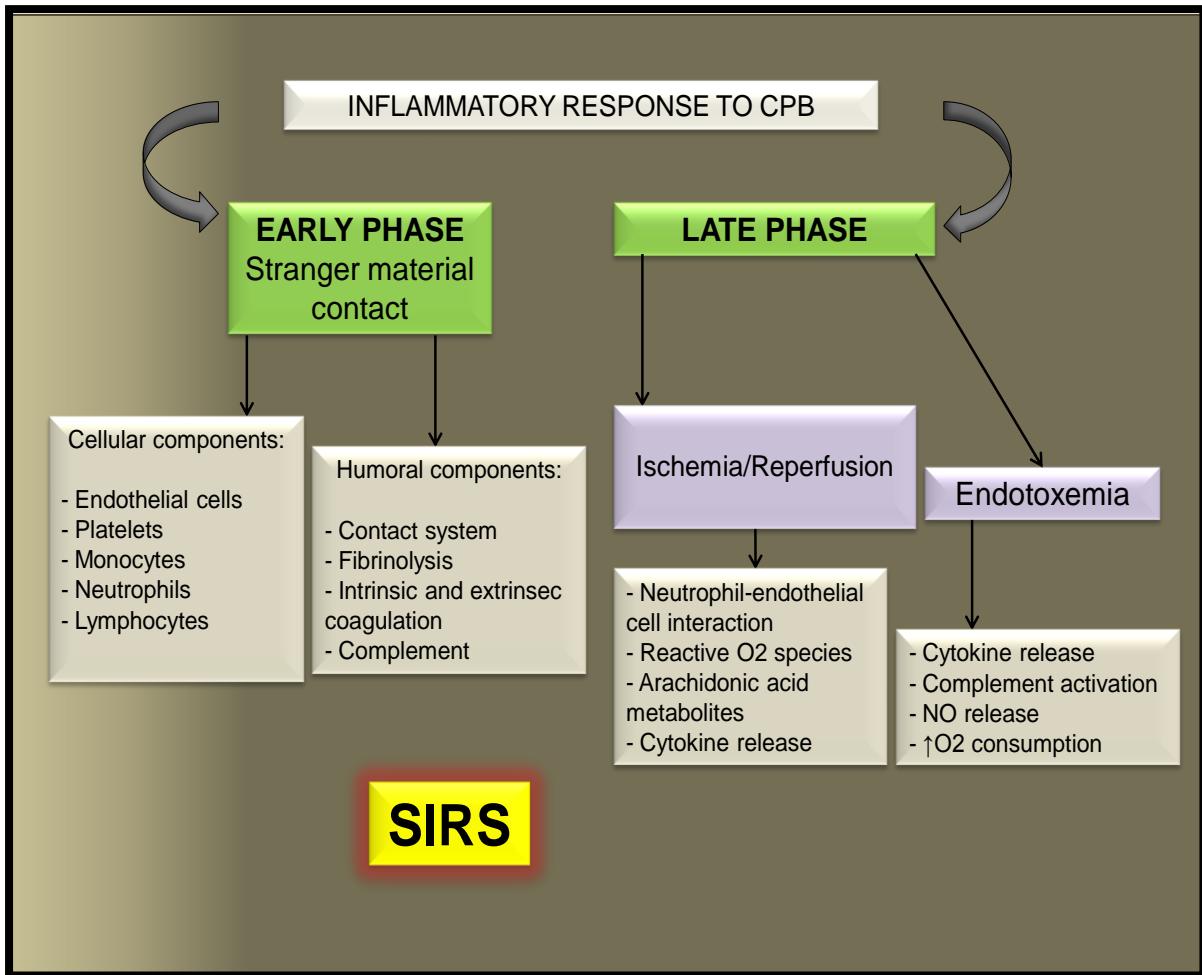
Figure 8. Neuroprotective mechanism(s) triggered by (α)-2-adrenoreceptors agonists. **Pre-synaptically:** 1- Activation of outward rectifying K⁺ channels causing hyperpolarization; 2- Inhibition of inward translocation of Ca²⁺ ions; 3- Hyperpolarization resulting from action-1 causes reduced Ca²⁺ entry; 4- Reduced intracellular Ca²⁺ (as a result of actions 2 and 3) causes diminished neurotransmitter release. **Synaptically:** 5- Due to action-4 as well as reduced receptor sensitivity; 6- Extrasynaptic scavenging of glutamate by astrocytes. **Post-synaptically:** 7- Hyperpolarization reduces the activation of NMDA receptors by enhancing Mg²⁺ block, and also causes reduced neuronal firing and reduced intracellular Ca²⁺ release; 8- Due to action 5 and 9. The reduced excitotoxic neuronal death due to combination of all actions but the main pathway is via a reduction in the free intracellular Ca²⁺. (Ma D, Rajakumaraswamy N, Maze M: α_2 -Adrenoreceptor agonists: shedding light on neuroprotection? Br Med Bull 2005; 71:77- 92 ¹⁰⁷) (Modified of Dexmedetomidine and neuroprotection Janke, EL, Samra, S. Seminars in Anesthesia, Perioperative Medicine and Pain 2006 25: 71-76 ¹⁵⁸).

Legends of schemes



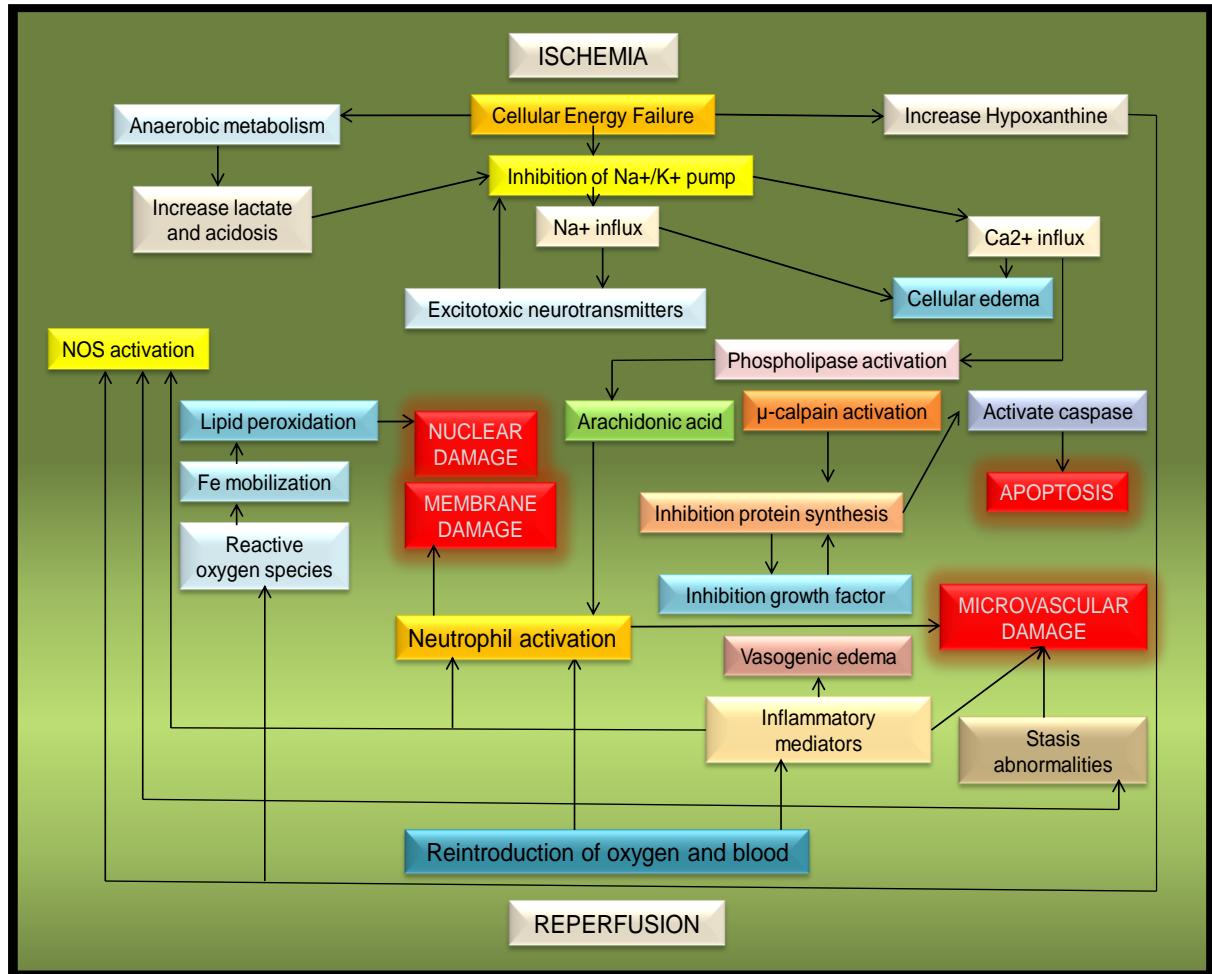
Scheme 1. Cardiopulmonary bypass and the extracorporeal circulation responses with the pathophysiologic changes resembling the systemic inflammatory response syndrome (SIRS). The contact of blood with xenosurfaces of the extracorporeal machine device, the ischemia/reperfusion and the hyperbaric oxygen triggered SIRS-like pathophysiological responses. The SIRS-like response is associated with overactivation of leukocytes, platelets (which can contribute to an increased coagulopathy), endothelial and cardiac cell. The secretion of pro-inflammatory factors by leukocytes and the increase tension and blood oxygenation stimulate the overproduction of reactive oxygen species (ROS), which feeds a vicious cycle of inflammation ↔ ROS production.

Legends of schemes



Scheme 2 - The inflammatory response to cardiopulmonary bypass is divided into 2 phases: "early" and "late" phases. The first phase is induced by the contact with xenosurfaces and the late phase is more related to oxygen reperfusion after ischemia and endotoxemia.

Legends of schemes



Scheme 3. Complex cascade of pathophysiologic phenomena associated with ischemia/reperfusion in CABG. Anaerobic metabolism carry to an increase on lactate and reduced pH with transmembrane pump impairment, which lead to a intracellular Ca^{2+} and Na^{+} increases, and consequently cellular edema. Increase on intracellular Ca^{2+} activated phospholipase A2 and calpain, with arachidonic acid degranulate and protein synthesis inhibition. Thus, caspase and neutrophil activation occur with cellular apoptosis. The neutrophils activation induce membrane lesions and more proinflammatory mediators liberation, including nitric oxid, through oxidonitrico sintase (NOS) activation and that lead to microvascular damage and endothelial impairment, in a vicious circle.

2.2 MANUSCRITO 2 - Dexmedetomidina reduz a resposta inflamatória após cirurgia miocárdica sob mini-circulação extracorpórea

Manuscrito 2

**DEXMEDETOMIDINE DECREASE THE INFLAMMATORY
RESPONSE TO MYOCARDIAL SURGERY UNDER MINI
CARDIOPULMONARY BYPASS**

NEUSA MARIA HEINZMANN BULOW, ELISÂNGELA COLPO,
EDUARDO FRANCISCO MAFASSIOLY CORREA, EMILY PANSERA
WACZUK, ROCHELLE SILVEIRA SCHLOSSER, ANELISE LAUDA, JOÃO
BATISTA TEIXEIRA DA ROCHA

Dexmedetomidine decrease the inflammatory response to myocardial surgery under mini cardiopulmonary bypass

Neusa Maria Heinzmann Bulow^{1,b}, Elisângela Colpo², Eduardo Francisco Mafassioly Correa^{3,b}, Emily Pansera Waczuk^{4,a}, Rochelle Silveira Schlosser^{5,b}, Anelise Lauda^{6,b}, João Batista Teixeira da Rocha^{7,a}

^a Departamento de Química, Programa de Pós-graduação em Ciências Biológicas: Bioquímica Toxicológica, Centro de Ciências Naturais e Exatas, Universidade Federal de Santa Maria, Cep 97105-900, Santa Maria, RS, Brazil.

^b Departamento de Cirurgia, Centro de Ciências da Saúde, Universidade Federal de Santa Maria, Cep 97105-900, Santa Maria, RS, Brazil.

*Corresponding author:

Neusa Maria Heinzmann Bulow and João Batista Teixeira da Rocha

UFSM – CCNE – Dep. de Química

Cep 97105-900, Santa Maria, RS, Brasil.

Tel: #55-55-3220-8140

Fax: #55-55-3220-8978

2.2.1 Abstract

Despite great technological advances in cardiac surgery, there is a high incidence of myocardial dysfunction and neurocognitive deficit or severe strokes in the postoperative period. Preventive measures are essential to reducing these adverse situations that are responsible for significant impairment on life quality. Surgery and cardiopulmonary bypass (CPB) with extracorporeal circulation produces important changes in the immune system, directly involved in the incidence of these complications. During this period occurs the release of proinflammatory cytokines and reduction of antiinflammatory cytokines, and an increase of reactive oxygen species (ROS). We hypothesize that the anesthetic choice could modify these inflammatory responses in patients undergoing coronary artery bypass grafting (CABG) surgery with mini-CPB. Methods: In a prospective, randomized and blinded study, we intended to demonstrate the influence of dexmedetomidine (TIVA-DEX group), an alpha-2-agonist anesthetic drug, as a component of a conventional total intravenous anesthesia (TIVA), on the behavior of this inflammatory response. Intraoperative dosage of cytokines, such as interleukin-1 (IL-1), interleukin-6 (IL-6), interleukin-10 (IL-10), gamma interferon (INF- γ), tumor necrosis factor (TNF- α), C-reactive protein (CRP), creatine phosphokinase (CPK), creatine phosphokinase-MB (CPK-MB), I troponin (cTnI), cortisol and glucose were performed. Lipid peroxidation was evaluated by the study of thiobarbituric acid reactive substances (TBARS) and the activity of delta-aminolevulinate dehydratase (d-ALA-D) was evaluated as an indicator of the production of reactive oxygen species. The blood collect samples times were before anesthesia (Time 1), at 90 minute after beginning of CPB (Time 2), 5 hours after beginning of CPB (Time 3) and at 24 hours after the end of surgery (Time 4). Results: Dexmedetomidine induce to a statistical significative reduction of IL-1, IL-6, TNF- α and INF- γ as compared to did not received dexmedetomidine group. The levels of IL-10 decreasing in both groups along the time, in a similar pattern. Do not occur difference between groups on d-ALA-D activity and TBARS were higher in dexmedetomidine group. Conclusion: Dexmedetomidine associated to TIVA was able to reduce plasma levels of IL-1, IL-6, TNF- α and INF- γ in patients under coronary artery bypass grafting surgery with mini-cardiopulmonary bypass, as compared to conventional TIVA patient group.

Keywords: Cytokines; Systemic Inflammatory Response Syndrome (SIRS); Total Intravenous Anesthesia (TIVA); Dexmedetomidine, Coronary Artery Bypass Grafting Surgery (CABG), Mini-Cardiopulmonary Bypass (Mini-CPB).

2.2.2 Introduction

Cardiac surgery induces a variety of metabolic, endocrine, and immune changes known as the "stress response", which may lead to prolonged in-hospital stay. Clinical manifestation include postoperative complications such as respiratory failure, wound infections (1), myocardial damage with contractile dysfunction, renal impairment, coagulopathy and neurologic dysfunction (2) with an increased mortality (3). Responses to cardiopulmonary bypass have been compared with the changes occurring in systemic inflammatory response syndrome (SIRS) (4). Mechanisms such the exposure of blood to nonphysiologic surfaces, surgical trauma, anesthesia, changes in body temperature, increased intestinal permeability by endotoxins, and ischemia/reperfusion injury (5) can be responsible to release into circulation of reactive oxygen species (ROS), proinflammatory cytokines, endothelins, platelet-activating factors, and endothelial and leukocyte adhesion molecules (6, 7).

Reactive oxygen species are recognized as critical mediators of cardiac and neurologic injury during ischemia/reperfusion. It is known that the cardiopulmonary bypass is potentially responsible for an activation of neutrophils, an important source of systemic primary reactive oxygen species, and damages related to activation and infiltration of neutrophils in reperfused tissues (4).

Although it has been shown that, compared with clinical management alone, conventional coronary artery bypass surgery with cardiopulmonary bypass prolongs life and reduces symptoms of patient, these benefits are accompanied by risks such as necessity of transfusions (30–90%), mortality (2–5%), stroke (2%), atrial fibrillation (30%), and neurocognitive dysfunction (50–60%) (8,9). These adverse clinical consequences have been attributed to the inflammatory responses to the cardiopulmonary bypass circuit, hypothermic cardiac arrest, aortic cannulation and cross-clamping (10, 11, 12). Nevertheless, there are considerable evidence that multiple pathways exist by which anesthetic agents have the potential to exert clinically important benefits (13, 14, 15, 16, 17, 18, 19, 20) and may be of interest to study potential benefit of specific anesthetic drugs, through its potential antiinflammatory mechanisms, that could reduce these postoperative complications and mortality.

Alpha(α)-2-adrenergic receptor agonists, such clonidine and dexmedetomidine, have been utilized in anesthesia to the sedative, analgesic, hemodynamic-stabilizing properties, and sympatholytic pharmacologic effects (21, 22). The stress response to surgery

can be attenuated by sympatholytic effects caused by postsynaptic activation of central (α)-2-adrenergic receptor, leading to reductions in blood pressure and heart rate (23). Recently, a few numbers of studies have investigated whether or not dexmedetomidine have antiinflammatory properties (24, 25, 26, 27, 28).

As corollary, it would be of great interest to have a safe and effective anesthetic drug with antiinflammatory properties to be used in a major surgery, especially in coronary artery bypass grafting surgery. In this regard, dexmedetomidine can be considered a promising candidate, because (α)-2-adrenergic agonists can modulate inflammatory response (29, 30, 31, 19). Here, we have hypothesized that dexmedetomidine in association with a conventional total intravenous anesthesia (infusion of propofol and sufentanil), could decrease the inflammatory response and oxidative stress associated with coronary artery bypass grafting surgery.

2.2.3 Materials and Methods

After institutional ethics review board approval and written informed patient consent, 30 clinical ASA II to III class patients, aged 42-72 yr, presenting for scheduled coronary artery bypass surgery under mini-cardiopulmonary bypass were assigned, according randomization, to the conventional total intravenous anesthesia (propofol/sufentanil) (TIVA; group 1-15 patients) or to TIVA with dexmedetomidine (propofol/sufentanil/dexmedetomidine) (TIVA-DEX; group 2- 15 patients) group. The surgery team, surgeon and perfusionist, was the same for all the patients that were recruited during two years (period of data collection).

Patient exclusion criteria included: severe ventricular dysfunction (left ventricle ejection fraction < 40%), reintervention surgery, need of blood products on the enter of cardiopulmonary bypass, preoperative history of liver or kidney dysfunction, immunological disease, preoperative intake of corticosteroids or anti-inflammatory drugs (except salicylic-acetyl acid) and history of a recent myocardial infarction (last two weeks).

Group 1 patients (TIVA): patients anesthetized with total intravenous anesthesia in target-controlled infusions (TCI infusion system, Diprifusor®; AstraZeneca, Wedwel, Germany) of propofol as hypnotic (an initial target blood concentration of 4 $\mu\text{g} \cdot \text{ml}^{-1}$) in induction and maintenance of anesthesia, based on bispectral (BIS) index evaluation (BIS values between 45 and 55). Associated with propofol, an infusion of sufentanil at a dose of 0.5 to 1 $\mu\text{g} \cdot \text{kg}^{-1}$ at induction and posterior maintenance of 0.5 to 1 $\mu\text{g} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ during surgery

time. The muscle relaxation for tracheal intubation was obtained with pancuronium (0.1 mg. kg⁻¹) at induction and additional 1/3 dose if necessary.

Group 2 patients (TIVA-DEX): patients anesthetized with total intravenous anesthesia in target-controlled infusions (TCI infusion system, Diprifusor®; AstraZeneca, Wedel, Germany) of propofol as hypnotic (an initial target blood concentration of 4 µg. ml⁻¹) in induction and maintenance of anesthesia, based on bispectral (BIS) index evaluation (BIS values between 45 and 55). Associated with propofol, an infusion of sufentanil at a dose of 0.5 to 1 µg. kg⁻¹ at induction and posterior maintenance of 0.5 to 1 µg. kg⁻¹. h⁻¹ during surgery time. All patients receive at anesthesia induction and during surgery, a continuous dexmedetomidine infusion, at 0.3 µg. kg⁻¹. h⁻¹ rate. The muscle relaxation for tracheal intubation was obtained with pancuronium (0.1 mg. kg⁻¹) at induction and additional 1/3 dose if necessary. Surgeons working in the operation room and the medical team on intensive care unit (ICU) were blinded to treatments protocols.

Systemic arterial blood pressure was measured via a radial artery catheterization. A Swan-Ganz catheter was inserted for central venous pressure (CVP), pulmonary capillary pressure (PCP) and cardiac index (CI) determinations. Hemodynamic parameters are intermittently monitored 24 hours after surgery, to intend maintenance of arterial blood pressure and cardiac index range at 20% basal levels. Cardioscopy, pulse oximetry, CO₂ exhaled levels, nasopharynx temperature and bispectral index were monitored. The surgery was conducted under mini-cardiopulmonary bypass and mild hypothermia (34-35 °C). Serials blood samples were collected to verify arterial gasometry, hemodilution and electrolytes.

For biomarkers determination, arterial blood was sampled at the radial performed catheterization before anesthesia induction (Time 1 or basal), 90 minutes after mini-cardiopulmonary bypass beginning (Time 2, during surgery), five hours after mini-CPB beginning (Time 3, within 2 to 3 hours after the end of surgery) and 24 hours after surgery end (Time 4).

FIGURE 1 ABOUT HERE

Citokynes (interleukin-1, interleukin-6, interleukin-10, TNF-α and INF-γ), were measured by chemical analysis (commercial kits: eBIOSCIENCE®, San Diego, USA). The plasma levels of C-reactive protein was measured by immunoassay (Dimension®, Siemens, Healthcare Diagnostics Inc., Newark, DE, USA), for cTnI determination was used a chemiluminescent method (IMMULITE®, Siemens, Healthcare Diagnostics Inc., Newark,

DE, USA), CPK and CPK-MB were evaluated by an enzymatic method (Dimension®, Siemens, Healthcare Diagnostics Inc., Newark, DE, USA), cortisol was determined using a chemiluminescent enzyme immunoassay (IMMULITE®, Siemens, Healthcare Diagnostics Inc., Newark, DE, USA) and glucose was determined by biochromatic method (Dimension®, Siemens, Healthcare Diagnostics Inc., Newark, DE, USA). TBARS (nmolMDA.ml^{-1} erythrocytes) were determinate based on a colorimetric method previously described (32) and δ -ALA-D activity were also performed at a colorimetric method (33).

Plasma samples were coded, and the investigators were blinded regarding treatment regimen. Similarly, all hemodynamic data were collected by trained observers who were not authors of this study and who were blinded to the anesthetic regimen used. They also recorded surgery duration, duration of mini-cardiopulmonary bypass, time for extubation, time in ICU and time for in-hospital stay. Possibly postoperative complications and necessity of inotropic support were investigated (considered the necessity of two or more inotropic drugs for hemodynamic stability) and the mini mental state examination (MMSE) was performed before and five days after surgery, considering the scholarly of patients.

All continuous data were expressed as mean \pm SD. Statistical analysis were performed by two-way ANOVA (2 anaesthetic procedures x 4 sampling time) with time factor treated as repeated measures. Values were considered to be statistical significant when P was < 0.05 .

2.2.4 Results

The characteristics of the two groups were similar in age, weight, height, comorbidities, mini-cardiopulmonary bypass time, total surgery time, time for extubation, time in intensive care unit (ICU), in-hospital stay time and mini mental state examination (Table 1).

TABLE 1 ABOUT HERE

Two-way ANOVA of mean arterial pressure (MAP), heart rate (HR) and arterial O_2 pressure (PO_2), indicated only a significant main effect of time of sampling ($P < 0.001$ for all cases), indicating that the anesthetic procedure for the groups were not significantly different (Table 2). Comparison in each group revealed a progressive statistical significant decrease in mean arterial pressure, as compared the basal time and another collected times (Figure 2) however, none patient necessitated special postoperative inotropic drug support.

Intravascular doses of dexmedetomidine induced dose-dependent decreases in systolic and diastolic blood pressure and in heart rate with important decreases in plasma norepinephrine levels (34). However, at high-bolus intravascular doses (50–75 µg), a transient initial hypertensive response may be seen, because an activation of peripheral vascular (α)-2-adrenergic receptors before the central sympatholytic effect on the vasomotor center occur (34). Here, we do not use a bolus dose of dexmedetomidine, and these patients were hemodynamic stable. Statistical analysis of the mean arterial pressure of both patient groups indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 2 and Table 2).

TABLE 2 AND FIGURE 2 ABOUT HERE

Statistical analysis of heart rate (HR) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthesia (TIVA and TIVA-DEX) indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 3).

FIGURE 3 ABOUT HERE

The hemodilution observed was important and similar between groups, with a significant difference intragroup with de time, as considered the basal time (Table 3). Statistical analysis of hematocrit (HT) of patients using two different anesthesia (TIVA and TIVA-DEX) indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 4). Statistical analysis of hemoglobin (HB) of patients also indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 5).

TABLE 3 ABOUT HERE

FIGURE 4 ABOUT HERE

FIGURE 5 ABOUT HERE

Plasma interleukin-1 (IL-1) statistical analysis of patients indicated a significant type of anesthesia versus sampling time ($p<0.0001$), and that the increase in IL-1, as a function of sample was lower in the patients anesthetized with TIVA-DEX than that with TIVA (Figure 6). A significant type of anesthesia versus sampling time ($p<0.0001$), indicating that the increase in interleukin-6 (IL-6), as a function of sample was also lower in the patients anesthetized with TIVA-DEX than that with TIVA (Figure 7).

FIGURE 6 ABOUT HERE

FIGURE 7 ABOUT HERE

Plasma interleukin-10 (IL-10) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthesia (TIVA and TIVA-DEX) indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 8) with a progressive reduction of IL-10 along time in both groups.

FIGURE 8 ABOUT HERE

Statistical analysis of plasma gamma interferon (INF- γ) of patients indicated a significant type of anesthesia versus sampling time ($p<0.0001$) and also indicating that the increase in INF- γ , as a function of sample was lower in the patients anesthetized with TIVA-DEX than that with TIVA (Figure 9). The same was observed with plasma tumor necrosis factor alpha (TNF- α) ($p<0.0001$) (Figure 10).

FIGURE 9 ABOUT HERE

FIGURE 10 ABOUT HERE

Erthrocytic thiobarbituric acid reactive substances (TBARS) of patients using two different anesthesia (TIVA and TIVA-DEX) indicated a significant interaction effect of type of anesthesia vs sampling time ($p<0.0001$) and revealed that the increase in TBARS after surgery was higher in TIVA-DEX than in TIVA patient group (Figure 11).

FIGURE 11 ABOUT HERE

Delta-amino levulinate dehydratase (δ -ALA-D) activity statistical analysis of patients using two different anesthesia (TIVA and TIVA-DEX) revealed no significant main or interaction effects (table 4).

TABLE 4 ABOUT HERE

Plasma C-reactive protein (CRP) of patients under coronary arterial bypass graft (CABG) surgery using two different anesthesia (TIVA and TIVA-DEX) indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 12) with a great increase in levels at 24 hour after surgery in both patient groups.

FIGURE 12 ABOUT HERE

Statistical analysis of plasma creatine phosphokinase (CPK) (Figure 13) and MB-creatine phosphokinase (MB-CPK) (Figure 14) of patients using two different anesthesia (TIVA and TIVA-DEX) presented only a significant main effect of sampling time ($p<0.0001$). Plasma I troponin (cTn-I) statistical analysis of patients indicated only a significant main effect of sampling time ($p<0.0001$) (Figure 15).

FIGURE 13 ABOUT HERE

FIGURE 14 ABOUT HERE

FIGURE 15 ABOUT HERE

Statistical analysis of cortisol (Figure 16) and glucose (Figure 17) of patients using two different anesthesia (TIVA and TIVA-DEX) indicated only a significant main effect of sampling time ($p<0.0001$).

FIGURE 16 ABOUT HERE

FIGURE 17 ABOUT HERE

Mini mental state examination (MMSE) statistical analysis of patients under coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthesia (TIVA and TIVA-DEX) indicated no significant main or interaction effects (all p values > 0.10) (Figure 18).

FIGURE 18 ABOUT HERE

2.2.5 Discussion

The main finding of the present study was that dexmedetomidine (as a component of total intravenous anesthesia-TIVA) modified the inflammatory response in coronary artery bypass grafting surgery under mini-cardiopulmonary bypass. Dexmedetomidine use was associated with less statistical significant increase in plasma IL-1, IL-6, TNF- α and INF- γ levels as compared to conventional TIVA patients group. In both groups of patient occurred a similar delayed postoperative decrease in IL-10.

Dexmedetomidine is a selective (α -2-adrenergic receptor agonist with a great ratio of (α -2 to (α -1 activity, of 1.620:1. Specific (α -2-adrenergic receptor subtypes mediate dexmedetomidine pharmacodynamic effects. Agonism at the (α -2A-adrenergic receptor

appears to promote sedation, hypnosis, analgesia, sympatholysis, neuroprotection (35) and inhibition of insulin secretion (36). Agonism at the (α)-2B-adrenergic receptor suppresses shivering centrally (37) induces analgesia at spinal cord and promote vasoconstriction in peripheral arteries. The (α)-2C-adrenergic receptor is associated with cognition, sensory processing, mood and regulation of epinephrine outflow from the adrenal medulla (38). Inhibition of norepinephrine release appears to be equally affected by all three (α)-2-adrenoreceptor subtypes (39). Dexmedetomidine also binds to imidazoline receptors and this activity may explain some of the non (α)-2-adrenergic receptor effects of this drug. Imidazoline-1 receptors are linked to G-proteins, modulate blood pressure regulation and have anti-arrhythmic effects (21). Imidazoline-2 receptors have been implicated in neuroprotection in a cerebral ischemia model in animals and in generation of memory (40).

Limited literature data have investigated the effects of dexmedetomidine and others (α)-2-adrenergic receptors agonists on cytokines (41) and TNF- α production by macrophages (30). Taniguchi et al. (19, 20) demonstrated that dexmedetomidine has an inhibitory effect on cytokine responses to endotoxemia. These findings suggest that one of the mechanisms of antiinflammatory effects of dexmedetomidine may be via modulation of cytokine production by macrophages and monocytes. Hofer et al. (2009) demonstrated that the dexmedetomidine infusion decreased proinflammatory cytokine production in sepsis (42). They have shown that preventive administration of clonidine or dexmedetomidine significantly improved survival after sepsis induction. This was accompanied by a reduction in the plasma proinflammatory mediators IL-1 β , IL- 6 and tumoral necrosis factor- α . Furthermore, Hofer et al. (2009) suggested that administration of a central acting (α)-2-adrenergic receptor agonist might be considered as a preventive therapeutic option in high-risk patients undergoing major surgery. In our study, patients were not in sepsis, but coronary artery bypass grafting surgery demanded a high immunological stress response, which resembles SIRS. Of particular therapeutic significance, here dexmedetomidine promoted IL-1, IL-6, INF- γ and TNF- α lesser increase, when compared to conventional TIVA group.

Tasdogan et al. (43) conducted a study to compare the effects of an intravenous infusion of propofol and dexmedetomidine, on inflammatory response and intra abdominal pressure in severe sepsis after abdominal surgery. Dexmedetomidine infusion decreased tumor necrosis factor-alpha, IL-1, and IL-6 levels and intra abdominal pressure more than did propofol infusion. In vitro studies with murine macrophages have also indicated an antiinflammatory effect of dexmedetomidine in experimental endotoxemia (44).

Bekker et al. (45) hypothesized that the intraoperative administration of dexmedetomidine would reduce the stress response and improve the quality of recovery in patients undergoing major spinal surgery. They compared a propofol/ fentanyl/ dexmedetomidine anesthesia group with propofol/ fentanyl/ placebo-saline anesthesia. Plasma cortisol levels increased in the post-anesthesia care unit in both groups; however, the increase was less accentuated in the dexmedetomidine group than in non-dexmedetomidine group (45). In contrast, C-reactive protein levels were similarly elevated in both groups after surgery, which is similar to our findings.

Erythrocyte TBARS levels were increased after CABG surgery, which indicates an increase in oxidative stress after cardiopulmonary bypass. However, TBARS levels were higher in dexmedetomidine-TIVA than in TIVA group. Literature has indicated that dexmedetomidine can decrease TBARS production in rodents after experimental surgery (46). Yagmurdur et al. (47), in human, related that dexmedetomidine significantly attenuated plasma hypoxanthine production in the ischemia and plasma malondialdehyde production in the reperfusion periods, after upper-extremity surgery requiring tourniquet application. Blood creatine phosphokinase and uric acid levels were significantly lower in the dexmedetomidine group as compared with those in the control group after reperfusion (47). Previously, in another investigation (48), our laboratory demonstrated that dexmedetomidine did not blunt blood TBARS increase, but protected δ -aminolevulinate dehydratase from inactivation caused by hyperoxygenation in total intravenous anesthesia, whereas the activity of enzyme decreased in patients anesthetized with remifentanil (48). Here δ -ala-D activity was not modified by anesthesia after CABG surgery in both groups.

In animal studies, dexmedetomidine inhibited cortisol synthesis at supratherapeutic concentrations but this has not been reported in short-term use in humans (49, 50). We have, before, in a human study, dexmedetomidine influence on cortisol levels of anesthetized patients also investigated (51). We measured cortisol concentrations before anesthetic induction, 5 minutes after intubation, and 30 minutes after surgical incision in patients undergoing gynecologic videolaparoscopic surgery, receiving dexmedetomidine or remifentanil. After intubation, there was a significant decrease in cortisol concentrations from baseline in both groups ($-4.3 \pm 1.4 \mu\text{g. dL}^{-1}$ and $-4.6 \pm 1.6 \mu\text{g. dL}^{-1}$, respectively) but only in the remifentanil group at 30 minutes after incision ($2.6 \pm 1.8 \mu\text{g. dL}^{-1}$ and $-7.1 \pm 2.1 \mu\text{g. dL}^{-1}$) (51) and dexmedetomidine did not suppress steroidogenesis. Here, at present study, we concluded the same. In the MENDS trial (52) cortisol concentrations were determined at baseline and two days after stopping dexmedetomidine infusion, and there was no statistically

significant difference in concentrations. At doses up to $1.5 \text{ } \mu\text{g. kg}^{-1} \cdot \text{h}^{-1}$, it does not appear that dexmedetomidine causes any clinically significant adrenal suppression (53).

Recently, Peng et al. (54) suggest that dexmedetomidine is a potent suppressor of lipopolysaccharide-induced inflammation in activated microglia and may be a potential therapeutic agent for the treatment of intensive care unit delirium. They investigated the effects of dexmedetomidine on the production of proinflammatory mediators in lipopolysaccharide-stimulated microglia. The concentrations of dexmedetomidine were chosen to correspond to 1, 10, and 100 times of clinically relevant concentration (i.e., 1, 10, and 100 ng. mL^{-1}). They measured the levels of proinflammatory mediators, such as inducible nitric oxide synthase or nitric oxide, prostaglandin E2, interleukin 1 β , and tumor necrosis factor alpha. Dexmedetomidine at 1 ng. mL^{-1} did not affect the production of proinflammatory mediators, but at 10 and 100 ng. mL^{-1} , dexmedetomidine significantly inhibited the release of nitric oxide, prostaglandin E2, interleukin 1 β , and tumor necrosis factor alpha. The dosage that we used here can be considered low to moderate, nevertheless affected significantly IL-1, IL-6, TNF- α and INF- γ levels.

Chen et al. (55) in a human model, demonstrated that cognitive deficit of patients, undergone laparoscopic cholecystectomy, assessed using the mini mental state examination (MMSE), for the dexmedetomidine and control groups one week after surgery (dexmedetomidine group, 27.6 ± 1.2 ; control group, 25.7 ± 1.5) were significantly different ($P=0.005$), with better scores on dexmedetomidine group. It suggested a dexmedetomidine neuroprotective effect in human patients. Here, we cannot significative differences on mini mental state examination scores between groups demonstrate at five days after surgery evaluation.

Intravascular doses of dexmedetomidine induced dose-dependent decrease in systolic and diastolic blood pressure and in heart rate with important decrease in plasma norepinephrine levels. However, at high-bolus intravascular doses ($50\text{--}75 \text{ } \mu\text{g}$), a transient initial hypertensive response may be seen, because an activation of peripheral vascular (α -2B-adrenergic receptors before the central sympatholytic effect on the vasomotor center occur (34). Because that, we do not use a bolus dose of dexmedetomidine at anesthesia induction.

Laringoscopy and endotracheal intubation also provoke marked sympathetic and sympathoadrenal response that increase the risk of perioperative myocardial ischemia and infarction. The perioperative use of dexmedetomidine may improve endocardial perfusion and decreasing heart rate with attenuation of stress response (56). Dexmedetomidine appear to increase the hemodynamic stability by altering the stress-induced sympathoadrenal responses

to intubation, during surgery and emergence from anesthesia (57) and this reflect on a better outcome. There are increasing evidence that the decrease in central nervous system sympathetic outflow by dexmedetomidine, in a dose-dependent manner, has organ protective effects against ischemic and hypoxic injury, including cardio-, neuro-, and reno-protection (58).

Dexmedetomidine possesses analgesic properties and many other advantageous influences that may make it a useful and safe adjunct in several clinical applications. When used as an adjunct to general anesthesia, dexmedetomidine can reduce both the minimum alveolar concentration requirement of inhalation agents and provide opiate-sparing properties up to 90% (59). We based our anesthesia on BIS index control. It was not the aim of the study to verify the total consume of propofol or sufentanil, although it was apparently a reduction in these drugs consumptions when dexmedetomidine was used simultaneously. However, the titration use of propofol did not induce an augmented release of stress hormones in response to cardiopulmonary bypass in the presence of potent narcotic, as previous demonstrated (60).

Althoug there were significant intergroup differences in plasma IL-1, IL-6, TNF- α and INF- γ levels, clinical outcome and in-hospital stay did not differ among groups at this study. Further larger studies are merited to determine the long-term relevance of the changes in biochemical markers observed in the related studies. One of the limitations of the present study was the small number of patients enrolled, however, the similarity on comorbidities, smoky habits, time of cardiopulmonary bypass and the same surgical team, established to confirm these results. The dose of dexmedetomidine utilized was considered a low dose, and other results about the antiinflammatory response could be obtained with higher doses.

2.2.6 Conclusions

It would be valuable to have a safe and effective means of preventing inflammatory response to a major surgery, like coronary artery bypass grafting surgery, and its complications, with the beneficial actions of anesthetic drugs. We believe that dexmedetomidine can be considered particularly promising. In this study we demonstrated the effect of dexmedetomidine on promote a lesser increase in IL-1, IL-6, TNF- α and INF- γ levels in patients under coronary artery bypass grafting surgery with mini cardiopulmonary bypass, as compared to patients group that not received dexmedetomidine. Others approach will require and additional research to further clarify both safety and efficacy to

dexmedetomidine use on these patients group. This can reflect on a reduction of postoperative complications, with a better clinical outcome.

Acknowledgments

Supported by FAPERGS (Fundação de Amparo a Pesquisa do Estado do Rio Grande do Sul), CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior), CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico), FINEP (Rede Instituto Brasileiro de Neurociência (IBN-Net) # 01.06.0842-00), FAPERGS-PRONEX-CNPQ and INCT-EN (Instituto Nacional de Ciência e Tecnologia em Excitotoxicidade e Neuroproteção).

2.2.7 References

1. Sander M, von Heymann C, von Dossow V, Spaethe C, Konertz, Uday Jain WF, Spies CD. Increased interleukin-6 after cardiac surgery predicts infection. *Anesth Analg* 2006; 102: 1623- 9
2. Murkin JM. Panvascular inflammation and mechanisms of injury in perioperative cns outcomes. *Sem Cardioth Vasc Anesth* 2010; 14: 190- 195
3. Plomondon ME, Cleveland JC Jr, Ludwig ST, et al. Off-pump coronary artery bypass is associated with improved risk-adjusted outcomes. *Ann Thorac Surg* 2001; 72: 114- 119
4. Wan S, Le Clerc JL, Vincent JL. Inflammatory response to cardiopulmonary bypass: mechanisms involved and possible therapeutic strategies. *Chest* 1997; 112: 676- 92
5. Elahi MM, Khan JS, Matata BM. Deleterious effects of cardiopulmonary bypass in coronary artery surgery and scientific interpretation of off-pump's logic. *Acute Cardiac Care* 2006; 8: 196- 209
6. Matata BM, Sosnowski AW, Galinanes M. Off-pump bypass graft operation significantly reduces oxidative stress and inflammation. *Ann Thorac Surg* 2000; 69: 785- 791
7. Matata BM, Galinanes M. Cardiopulmonary bypass exacerbates oxidative stress but does not increase proinflammatory cytokine releasein patients with diabetes compared with patients without diabetes: Regulatory effects of exogenous nitric oxide. *J Thorac Cardiovasc Surg* 2000; 120: 1- 11
8. Stover EP, Siegel LC, Parks R, Levin J, Body SC, Maddi R, D'Ambra MN, Mangano DT, Spies BD. Variability in transfusion practice for coronary artery bypass surgery persists

- despite national consensus guidelines: A 24-institution study. Institutions of the Multicenter Study of Perioperative Ischemia Research Group. *Anesthesiology* 1998; 88: 327- 339
9. Stamou SC, Hill PC, Dangas G, Pfister AJ, Boyce SW, Dullum MK, Bafi AS, Corso PJ. Stroke after coronary artery bypass: Incidence, predictors, and clinical outcome. *Stroke* 2001; 32: 1508- 13
 10. Mathew JP, Parks R, Savino JS, Friedman AS, Koch C, Mangano DT, Browner WS. Atrial fibrillation following coronary artery bypass graft surgery: Predictors, outcomes, and resource utilization. Multi Center Study of Perioperative Ischemia Research Group. *JAMA* 1996; 276: 300- 6
 11. Rose EA. Off-pump coronary-artery bypass surgery. *N Engl J Med* 2003; 348: 379- 80 12
 12. Ascione R, Caputo M, Angelini GD. Off-pump coronary artery bypass grafting: not a flash in the pan. *Ann Thorac Surg* 2003; 75: 306- 13
 13. Aantaa, R., Jalonens, J. Perioperative use of alpha2-adrenoceptor agonists and the cardiac patient. *Eur J Anaesthesiol* 2006; 23: 361- 372,
 14. Corcoran TB, Engel A, Sakamoto H, O'shea A, O'callaghan-Enright S, Shorten GD. The effects of propofol on neutrophil function, lipid peroxidation and inflammatory response during elective coronary artery bypass grafting in patients with impaired ventricular function. *Br J Anaesth* 2006; 97: 825- 831
 15. Memis D, Hekim, Glu S, Vatan I, Yandim T, Yuksel M, Sut N. Effects of midazolam and dexmedetomidine on inflammatory responses and gastric intramucosal pH to sepsis, in critically ill patients. *Br J Anaesth* 2007; 98: 550- 2
 16. Murphy PG, Myers DS, Davies MJ, Webster NR, Jones JG. The antioxidant potential of propofol (2,6 -diisopropylphenol). *Br J Anaesth* 1992; 68: 613- 8
 17. Nader ND, Ignatowski TA, Kurek CJ, Knight PR, Spengler RN. Clonidine suppresses plasma and cerebrospinal fluid concentrations of TNF-alpha during the perioperative period. *Anesth Analg*; 93: 363- 9
 18. Navapurkar V, Skepper J, Jones J, Menon D. Propofol preserves the viability of isolated rat hepatocyte suspensions under an oxidant stress. *Anesth Analg*; 87: 1152- 7
 19. Taniguchi T, Kidani Y, Kanakura H, Takemoto Y, Yamamoto K. Effects of dexmedetomidine on mortality rate and inflammatory responses to endotoxin-induced shock in rats. *Crit Care Med*; 32: 1322- 6
 20. Taniguchi T, Kurita A, Kobayashi K, Yamamoto K, Inaba H. Dose- and time-related effects of dexmedetomidine on mortality and inflammatory responses to endotoxin-induced shock in rats. *J Anesth*; 22: 221- 8

21. Khan ZP, Ferguson CN, Jones RM. Alpha-2 and imidazoline receptor agonists. Their pharmacology and therapeutic role. *Anesthesia* 1999; 54: 146- 65
22. Carollo DS, Nossaman BD, Ramadhyani U. Dexmedetomidine: A review of clinical applications. *Curr Opin Anaesthesiol* 2008; 21: 457- 61
23. Candiotti KA, Bergese SD, Bokesch PM, Feldman MA, Wisemandle W, Bekker AY. Monitored anesthesia care with dexmedetomidine: A prospective, randomized, double-blind, multicenter trial. *Anesth Analg*. 2010; 110: 47- 56
24. Koca U, Olguner ÇG, Ergür BU, Altekin E, Taşdögen A, Duru S, Girgin P, Gündüz K, Cilaker Micili S, Güzelda S, Akkuş M. The effects of dexmedetomidine on secondary acute lung and kidney injuries in the rat model of intra-abdominal sepsis. *The Scientific World Journal* 2013; art. no. 292687
25. Can M, Gul S, Bektas S, Hancı V, Acikgos S. Effects of dexmedetomidine or methylprednisolone on inflammatory responses in spinal cord injury *Acta Anaesth Scand* 2009; 53: 1068- 1072
26. Gu J, Chen J, Xia P, Tao G, Zhao H, Ma D. Dexmedetomidine attenuates remote lung injury induced by renal ischemia-reperfusion in mice Authors: *Acta Anaesth Scand* 2011; 55: 1272-1278
27. Wu X, Song X, Li N, Zhan L, Meng Q, Xia Z. Protective effects of dexmedetomidine on blunt chest trauma-induced pulmonary contusion in rats. *Journal of Trauma and Acute Care Surgery* 2013; 74: 524–530
28. Sukegawa S, Inoue M, Higuchi H, Tomoyasu Y, Maeda S, Miyawaki T. Locally injected dexmedetomidine inhibits carrageenin-induced inflammatory reactions in injected region. ASA annual meeting 2011; A1590
29. Lai YC, Tsai PS, Huang CJ. Effects of dexmedetomidine on regulating endotoxin-induced up-regulation of inflammatory molecules in murine macrophages. *J Surg Res* 2009; 154: 212- 219
30. Szelenyi J, Kiss JP, Vizi ES. Differential involvement of sympathetic nervous system and immune system in the modulation of TNF-alpha production by alpha2- and beta-adrenoceptors in mice. *J Neuroimmunol* 2000; 103: 34- 40
31. Sleigh J. All hands on dex. *Anaesthesia* 2012; 67: 1193–1197
32. Puntel RL, Roos DH, Grotto D, Garcia SC, Nogueira CW, Rocha JB. Antioxidant properties of Krebs cycle intermediates against malonate pro-oxidant activity in vitro: a comparative study using the colorimetric method and HPLC analysis to determine malondialdehyde in rat brain homogenates. *Life Sci* 2007; 81: 51-62

33. Berlin A, Schaller KH. European standardized method for determination of delta-aminolevulinic-acid dehydratase activity in blood. *Z. Klin. Chem. Klin. Biochem* 1974; 12: 389– 390
34. Talke P, Richardson CA, Scheinin M, et al. Postoperative pharmacokinetics and sympatholytic effects of dexmedetomidine. *Anesth Analg* 1997; 85: 1136- 1142
35. Ma D, Hossain M, Rajakumaraswamy N, et al. Dexmedetomidine produces its neuroprotective effect via the alpha 2A-adrenoceptor subtype. *Eur J Pharmacol* 2004; 502: 87- 97
36. Fagerholm V, Scheinin M, Haaparanta M. Alpha2A-adrenoceptor antagonism increases insulin secretion and synergistically augments the insulinotropic effect of glibenclamide in mice. *Br J Pharmacol* 2008; 154: 1287- 1296
37. Takada K, Clark DJ, Davies MF, et al. Meperidine exerts agonist activity at the alpha (2B)-adrenoceptor subtype. *Anesthesiology* 2002; 96: 1420- 1426
38. Fagerholm V, Rokka J, Nyman L, et al. Autoradiographic characterization of alpha (2C)-adrenoceptors in the human striatum. *Synapse* 2008; 62: 508- 515
39. Moura E, Afonso J, Hein L, Vieira-Coelho MA. Alpha2-adrenoceptor subtypes involved in the regulation of catecholamine release from the adrenal medulla of mice. *Br J Pharmacol* 2006; 149: 1049-58
40. Takamatsu I, Iwase A, Ozaki M, et al. Dexmedetomidine reduces long-term potentiation in mouse hippocampus. *Anesthesiology* 2008; 108: 94- 102
41. Straub RH, Herrmann M, Berkmler G, et al. Neuronal regulation of interleukin 6 secretion in murine spleen: adrenergic and opioidergic control. *J Neurochem* 1997; 68: 1633- 1639
42. Hofer S, Steppan J, Wagner T, Funke B, Lichtenstern C, Martin E, Graf BM, Bierhaus A, Weigand MA. Central sympatholytics prolong survival in experimental sepsis. *Crit Care* 2009; 13: R11
43. Tasdogan M, Memis D, Sut N, Yuksel M. Results of a pilot study on the effects of propofol and dexmedetomidine on inflammatory responses and intraabdominal pressure in severe sepsis. *J Clin Anesth* 2009; 21: 394-400
44. Lai YC, Tsai PS, Huang CJ. Effects of dexmedetomidine on regulating endotoxin-induced up-regulation of inflammatory molecules in murine macrophages. *J Surg Res* 2009; 154: 212- 219

45. Bekker A, Haile M, Kline R; Didehvar S, Babu R, Martiniuk F, Urban M. The Effect of Intraoperative Infusion of Dexmedetomidine on the Quality of Recovery After Major Spinal Surgery. *J Neurosurg Anesthesiol* 2013; 25: 16-24
46. Arslan M, Çomu FM, Kuçuk A, Ozturk L, Yaylak F. Dexmedetomidine protects against lipid peroxidation and erythrocyte deformability alterations in experimental hepatic ischemia reperfusion injury. *Libyan J Med* 2012; 7: 18185
47. Yagmurdur H, Ozcan N, Dokumaci F, Kilinc K, Yilmaz F, Basar H. Dexmedetomidine Reduces the Ischemia-Reperfusion Injury Markers During Upper Extremity Surgery With Tourniquet. *Journal of Hand Surgery* 2008; 33: 941- 947
48. Rocha JBT, Bulow NMH, Correa EFM, Scholze C, Nogueira CW, Barbosa NBV. Dexmedetomidine protects blood d-aminolevulinate dehydratase from inactivation caused by hyperoxygenation in total intravenous anesthesia. *Human and Experimental Toxicology* 2010; 30: 289- 295
49. Maze M, Virtanen R, Daunt D, Banks SJ, Stover EP, Feldman D. Effects of dexmedetomidine, a novel imidazole sedative-anesthetic agent, on adrenal steroidogenesis: in-vivo and in-vitro studies. *Anesth Analg* 1991; 73: 204- 8 94
50. Venn RM, Bryant A, Hall GM, Grounds RM. Effects of dexmedetomidine on adrenal cortical function, and the cardiovascular, endocrine, and inflammatory responses in post-operative patients needing sedation in the intensive care unit. *Br J Anaesth* 2001; 86: 650-6
51. Bulow NMH, Barbosa NBV, Rocha JBT. Opioid consumption in total anesthesia is reduced with dexmedetomidine: a comparative study with remifentanil in gynecologic videolaparoscopic surgery. *J Clin Anesth* 2007; 19: 280- 5.
52. Pandharipande PP, Pun BT, Herr DL, et al. Effects of sedation with dexmedetomidine vs lorazepam on acute brain dysfunction in mechanically ventilated patients: the MENDS randomized controlled trial. *JAMA* 2007; 298: 2644- 53
53. Gerlach AT, Murphy CV, Dasta JF. An updated focused review of dexmedetomidine in adults. *The Annals of Pharmacotherapy* 2009; 43: 2064-2074
54. Peng M, Wang Y-L, Wang C-Y, Chen C. Dexmedetomidine attenuates lipopolysaccharide-induced proinflammatory response in primary microglia. *Journ Surg Res* 2013; 179: 219-225
55. Chen J, Yan J, Han X. Dexmedetomidine may benefit cognitive function after laparoscopic cholecystectomy in elderly patients. *Exp Therap Med* 2013; 5: 489-494
56. Sulaiman S, Karthekeyan RB, Vakamundi M, Sundair AS, Ravullapalli H, Gandhan R. The effects of dexmedetomidine on attenuation of stress response to endotracheal intubation

- in patients undergoing elective off pump coronary artery bypass grafting. Ann Cardiac Anaesth 2012; 15:1
57. Scheinin B, Lindgren L, Randell T, Scheinin H, Scheinin M. Dexmedetomidine attenuates sympathoadrenal responses to tracheal intubation and reduces the need for thiopentone and preoperative fentanyl. Br J Anaesth 1992; 68: 126- 31
58. Hall JE, Uhrich TD, Barney JA, Arain SR, Ebert TJ. Sedative, amnestic, and analgesic properties of small-dose dexmedetomidine infusions. Anesth Analg 2000; 90: 699- 705
59. Aho M, Erkola O, Kallio A, Scheinin H, Korttila K. Dexmedetomidine infusion for maintenance of anesthesia in patients undergoing abdominal hysterectomy. Anesth Analg 1992; 75: 940- 6
60. Bauer M, Wilhelm W, Kraemer T, Kreuer S, Brandt A, Adams HA, Hoff G, Larsen R. Impact of bispectral index monitoring on stress response and propofol consumption in patients undergoing coronary artery bypass surgery. Anesthesiology 2004; 101: 1096-1104

Legend of figures

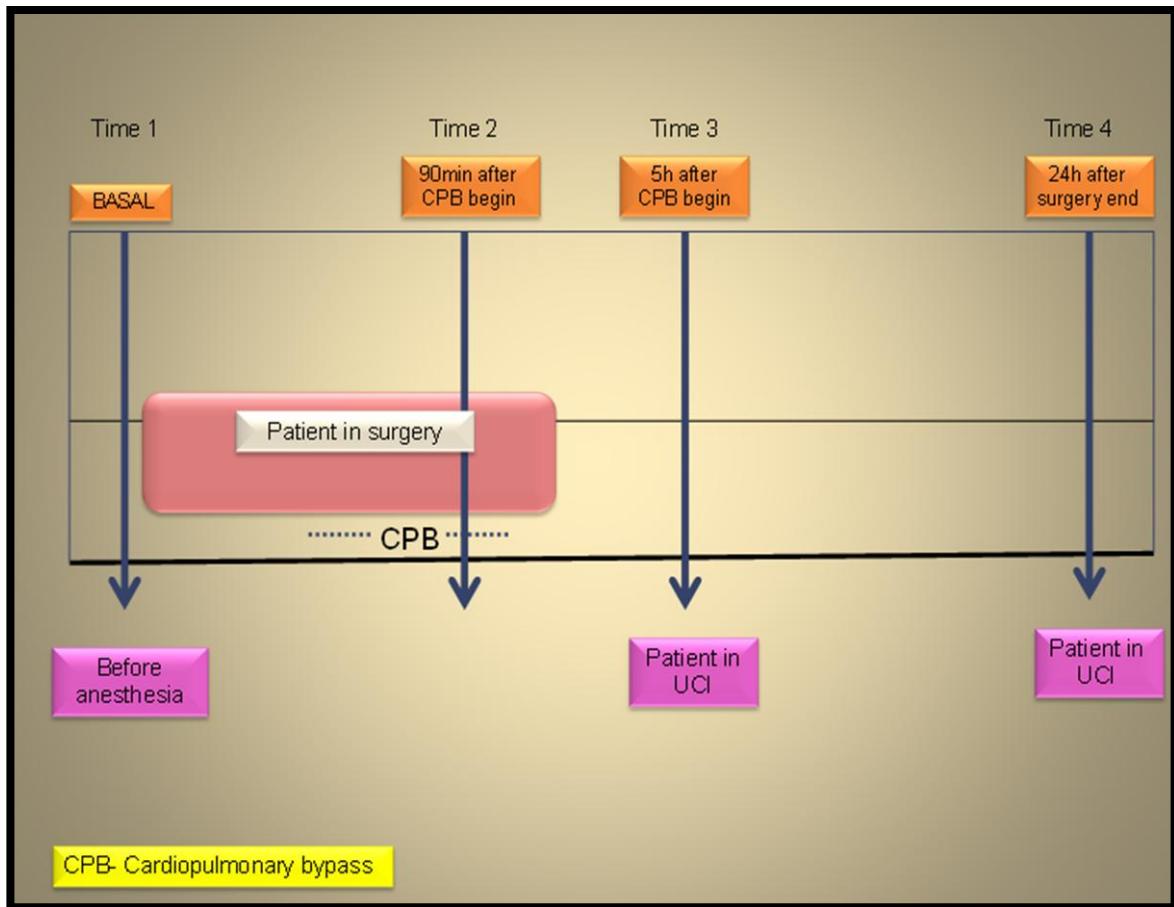


Figure 1. Sampling Protocol. Arterial blood at radial access was collected at four times. The first sample was before the anesthesia induction, considered the basal time (Time 1). Second sample was performed at 90 minute after cardiopulmonary bypass (CPB) beginning (Time 2). The third sample was at 5 hours after CPB beginning (Time 3) and the fourth sample was performed at 24 hours after surgery end (Time 4).

Legend of figures

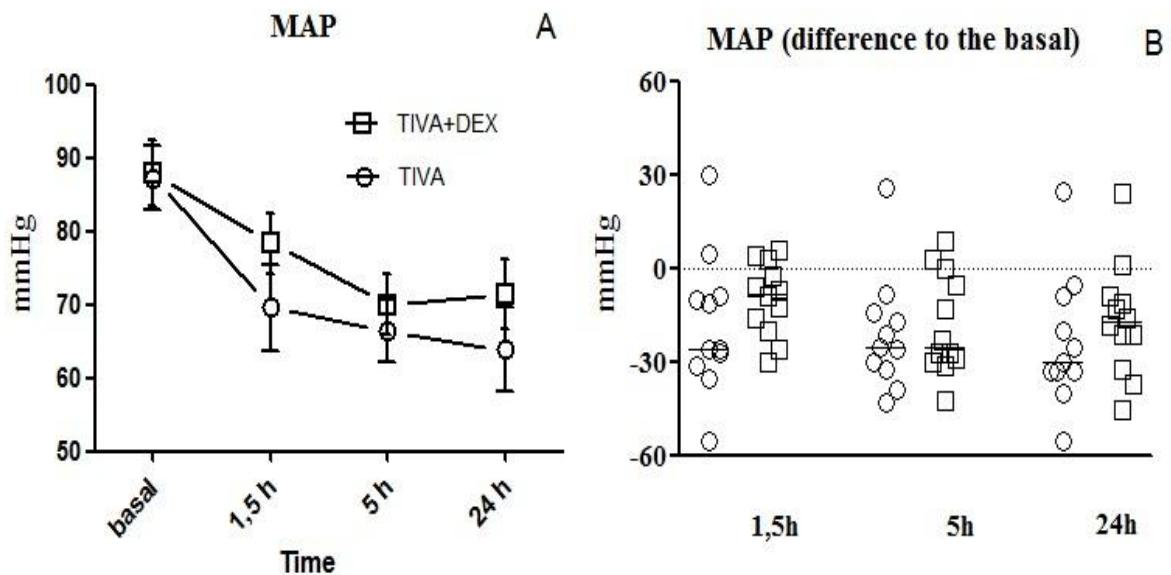


Figure 2. Mean arterial pressure (MAP) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$) (two-way).

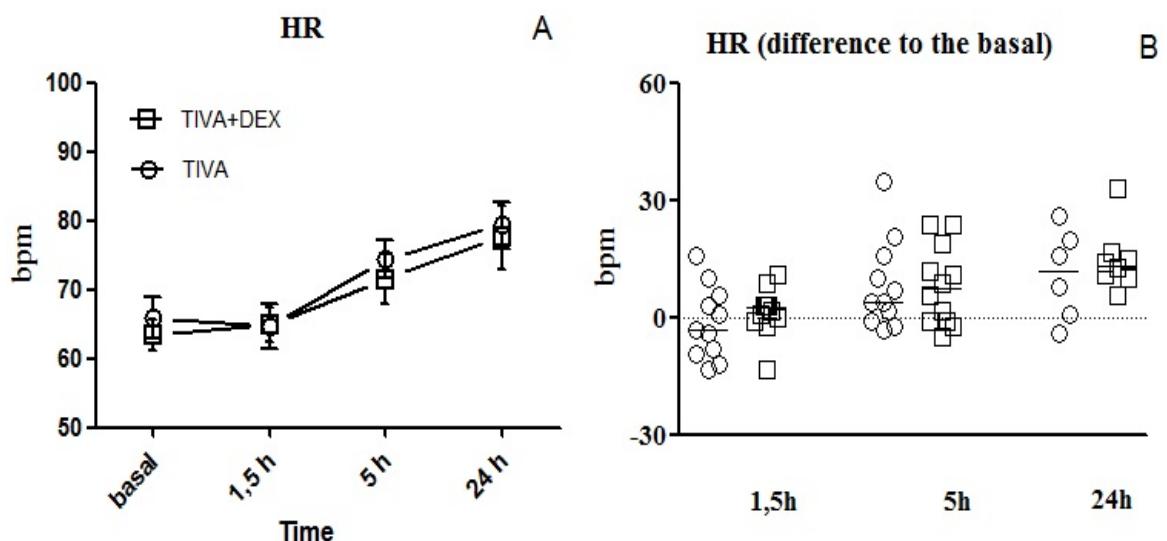


Figure 3. Statistical analysis of heart rate (HR) in patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX) indicated only a significant main effect of sampling time ($p<0.0001$).

Legend of figures

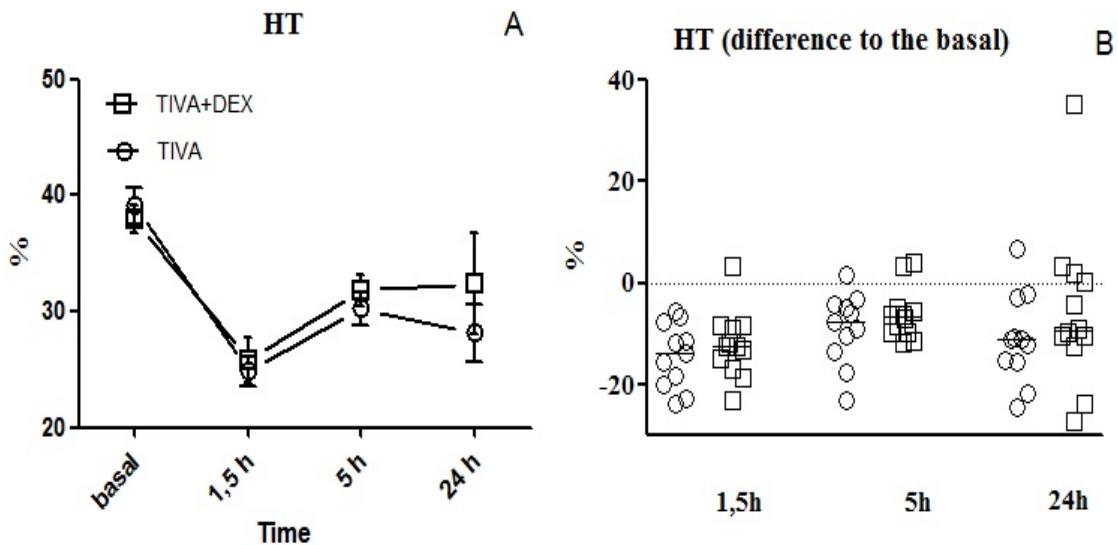


Figure 4. Hematocrit (HT) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$).

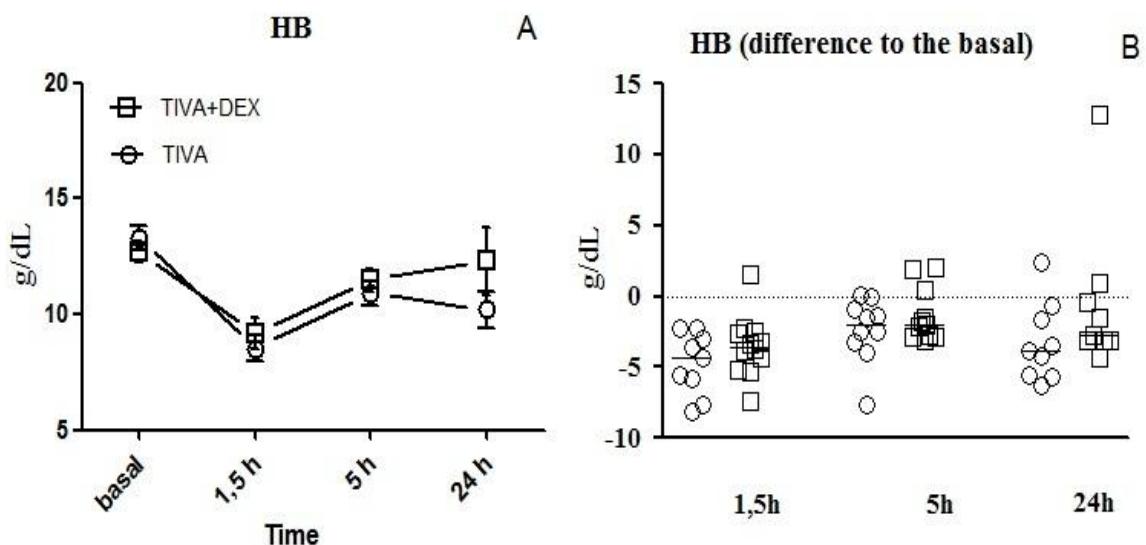


Figure 5. Hemoglobin (HB) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$).

Legend of figures

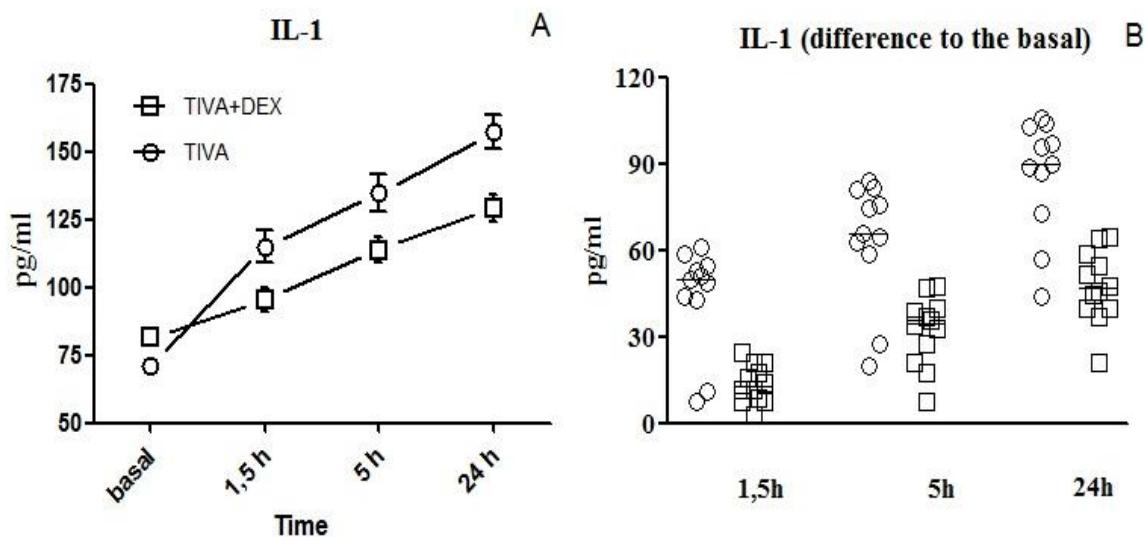


Figure 6. Plasma interleukin-1 (IL-1) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated a significant type of anesthesia versus sampling time ($p<0.0001$) and indicating that the increase in IL-1, as a function of sample was lower in the patients anesthetized with TIVA-DEX than that with TIVA.

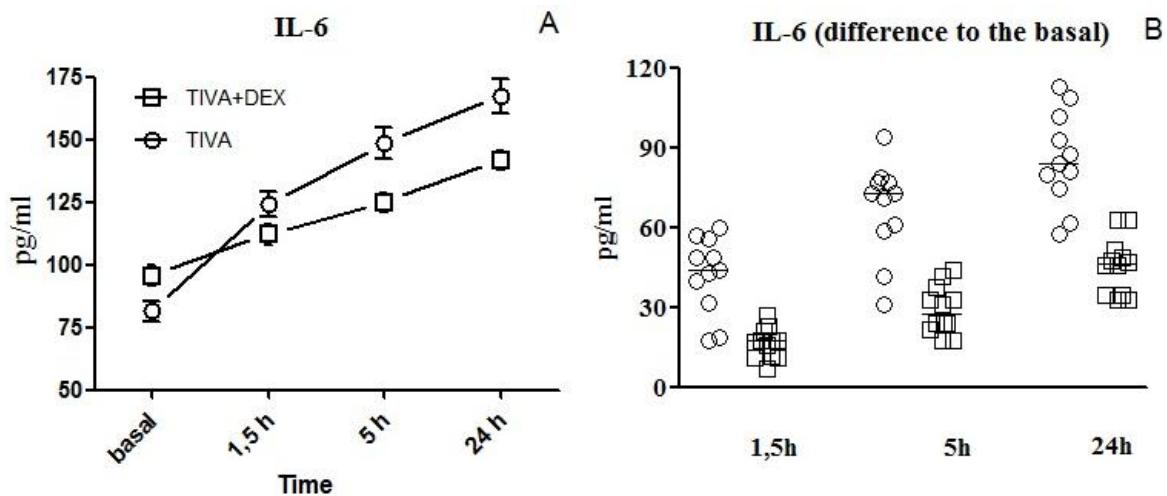


Figure 7. Plasma interleukin-6 (IL-6) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated a significant type of anesthesia versus sampling time ($p<0.0001$) and indicating also that the increase in IL-6, as a function of sample was lower in the patients anesthetized with TIVA-DEX than that with TIVA.

Legend of figures

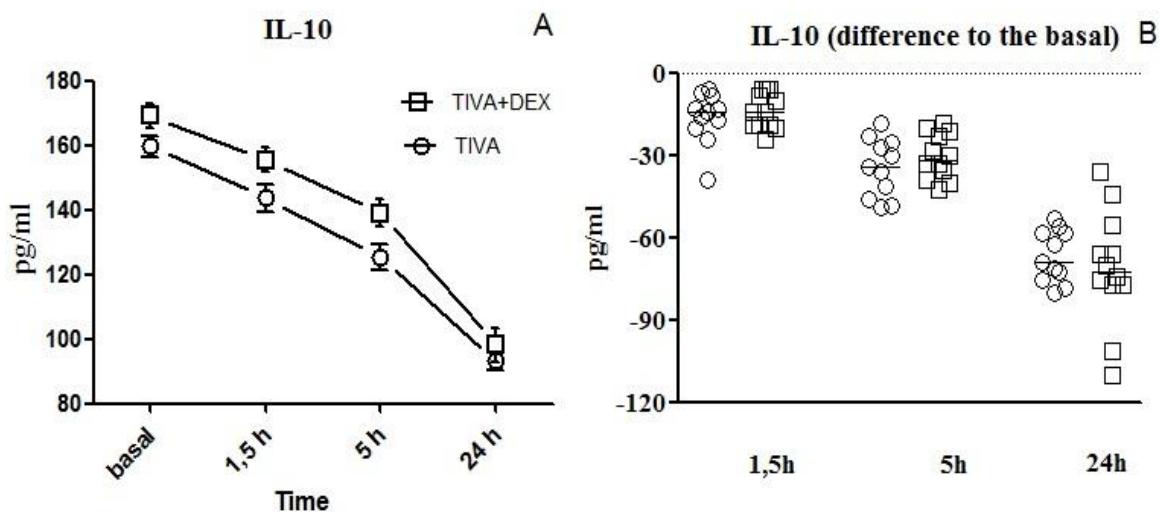


Figure 8. Plasma interleukin-10 (IL-10) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$) with a progressive decrease of IL-10 in both groups along time.

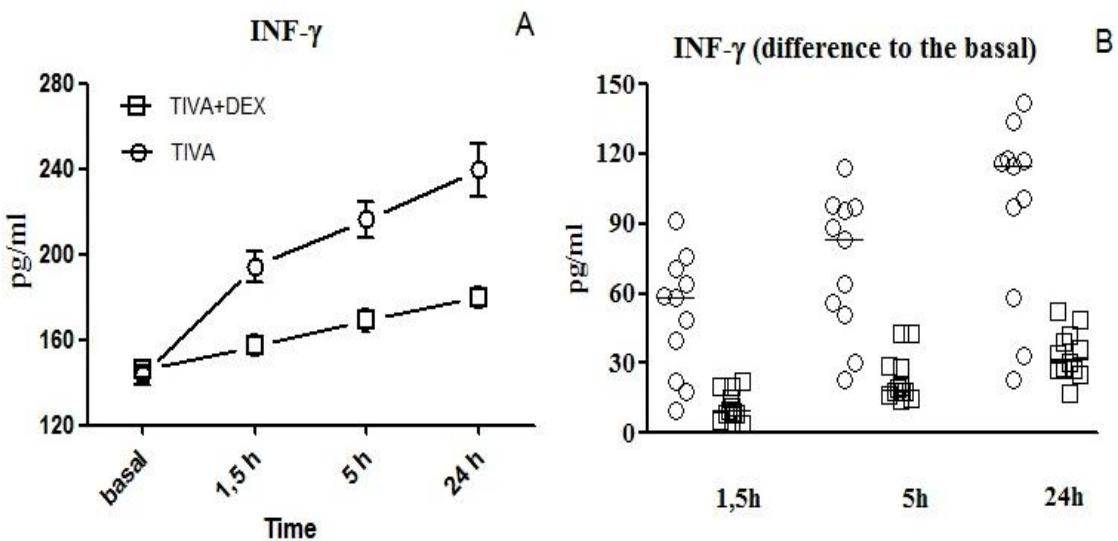


Figure 9. Plasma gamma interferon (INF- γ) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated a significant type of anesthesia versus sampling time ($p<0.0001$) and indicating that the increase in INF- γ , as a function of sample was lower in the patients anesthetized with TIVA-DEX than that with TIVA.

Legend of figures

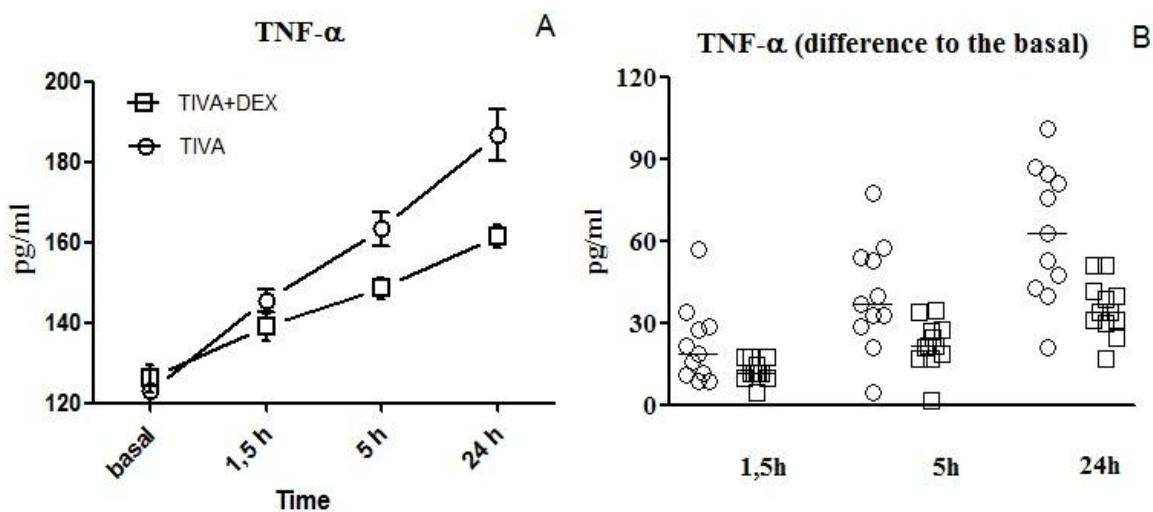


Figure 10. Plasma tumoral necrosis factor-alpha (TNF- α) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated a significant type of anesthesia versus sampling time ($p<0.0001$), indicating that the increase in TNF- α , as a function of sample was lower in the patients anesthetized with TIVA-DEX than that with TIVA.

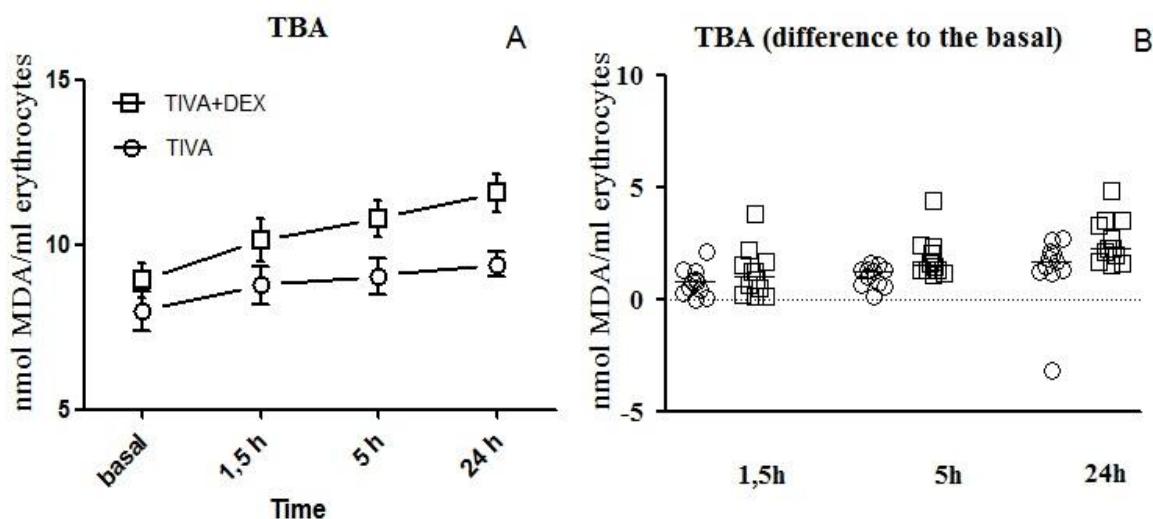


Figure 11. Erythrocytic thiobarbituric acid reactive substances (TBARS) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated a significant interaction effect of type of anesthesia vs sampling time ($p<0.0001$), indicating that the increase in TBARS after surgery was higher in TIVA-DEX than in TIVA patient group.

Legend of figures

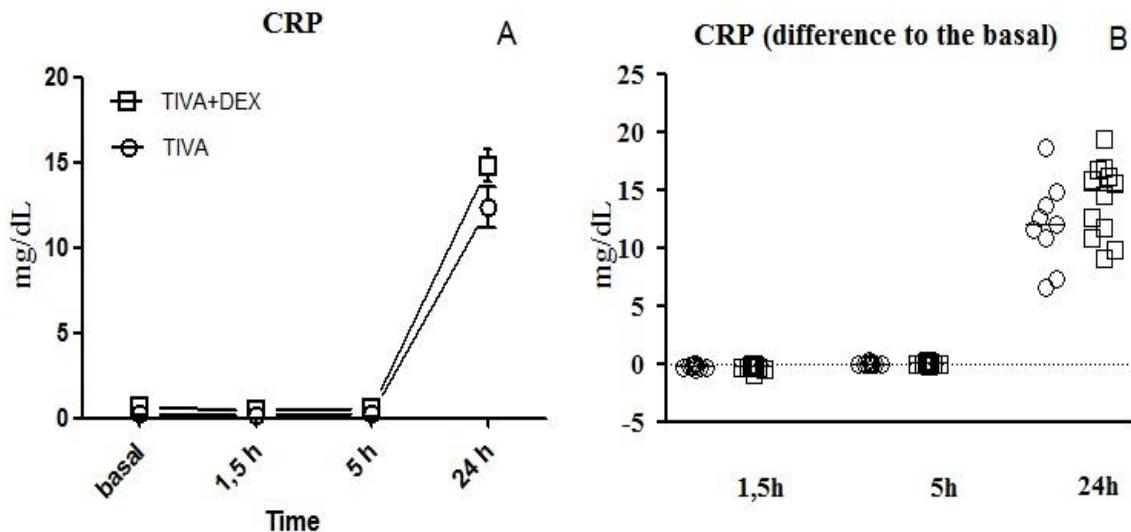


Figure 12. Plasma C-reactive protein (CRP) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthesia (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$) with a significant increase in CRP at 24 hours after end of surgery in both patients groups.

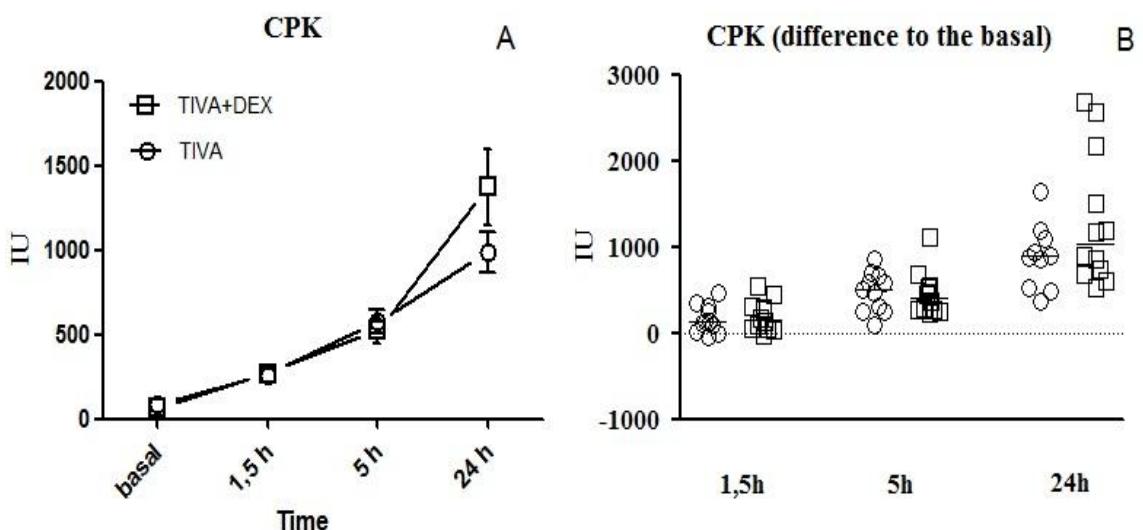


Figure 13. Plasma creatine phosphokinase (CPK) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthesia (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$) with a similar increase of CPK levels along time in both groups.

Legend of figures

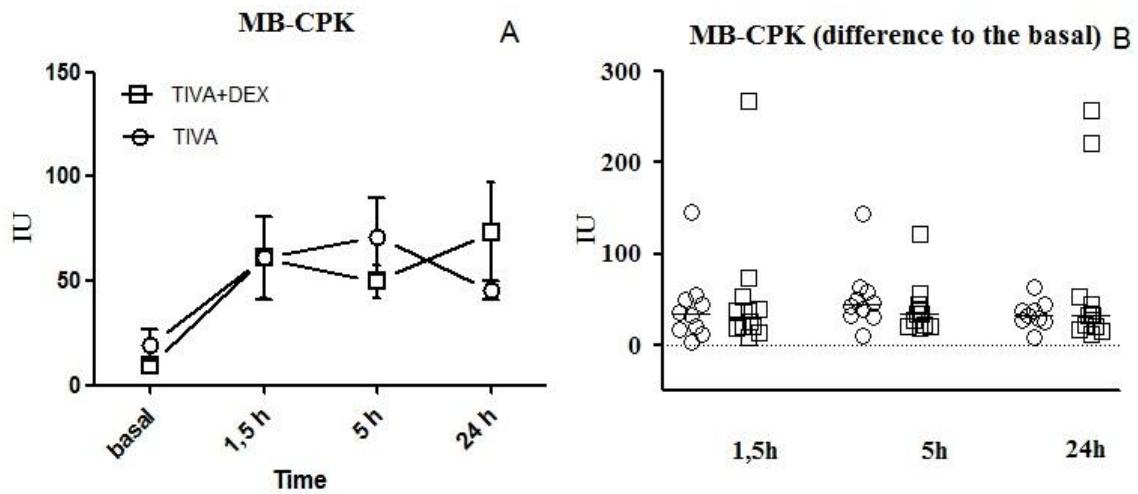


Figure 14. Plasma MB-creatine phosphokinase (MB-CPK) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$).

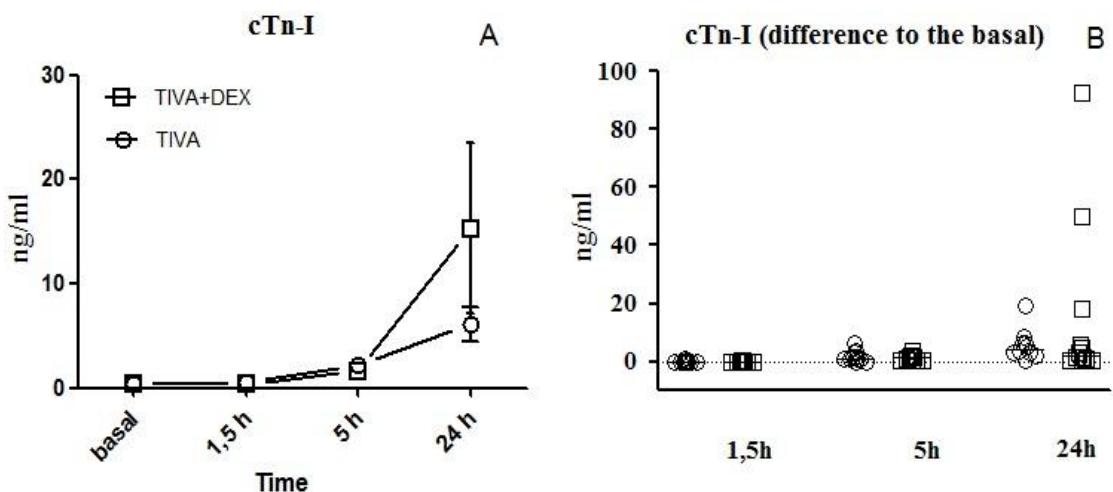


Figure 15. Plasma I troponin (cTn-I) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$), with a significant increase in cTn-I at 24 hours after end of surgery in both patient groups.

Legend of figures

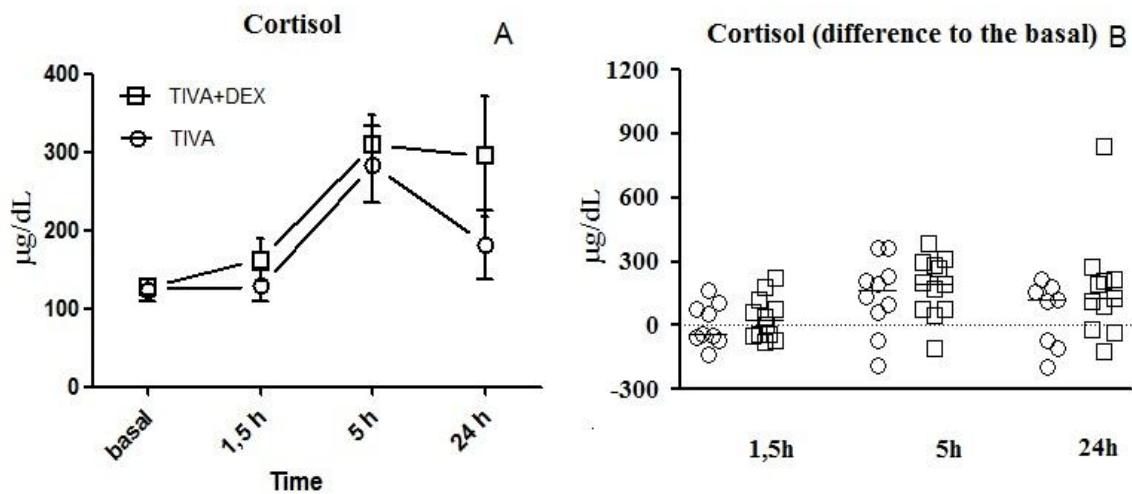


Figure 16. Plasmatic cortisol of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$). Similarly, in both groups cortisol increase at 5 hours after CPB beginning and 24 hours after end of surgery.

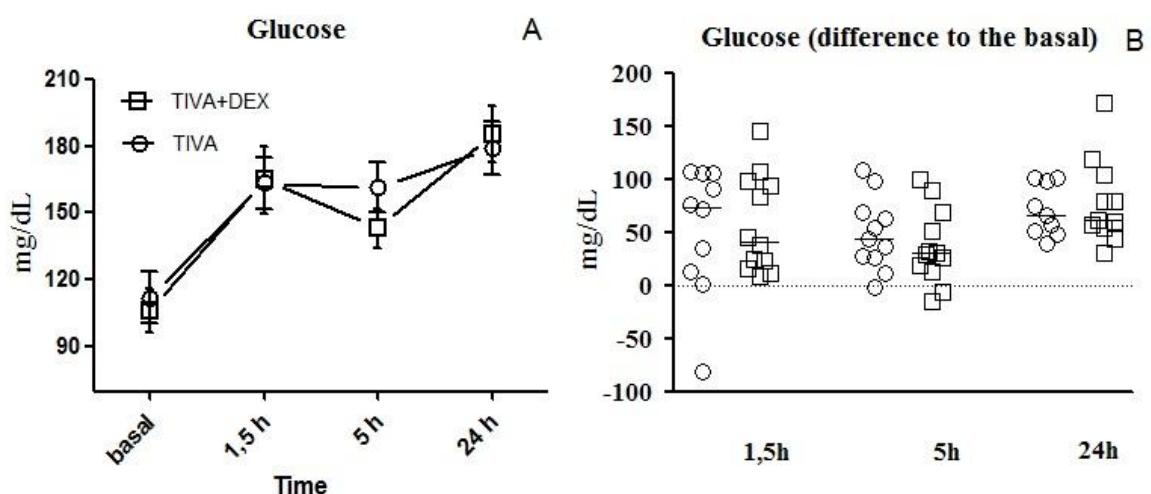


Figure 17. Plasmatic glucose of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated only a significant main effect of sampling time ($p<0.0001$) with increasing levels of glucose in both groups.

Legend of figures

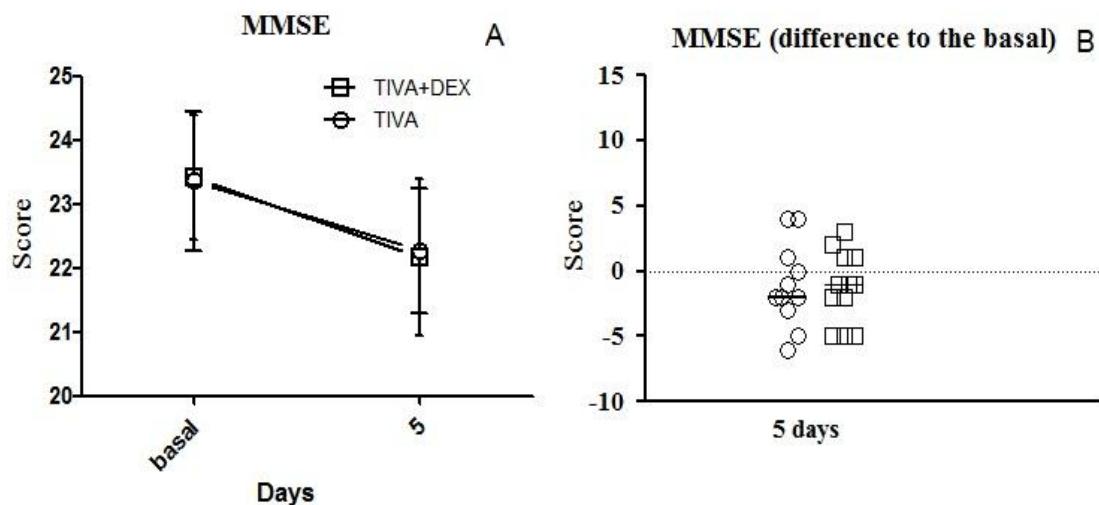


Figure 18. Mini mental state examination (MMSE) of patients submitted to coronary arterial bypass graft (CABG) surgery under mini-cardiopulmonary bypass, using two different anesthetics (TIVA and TIVA-DEX). Statistical analysis indicated no significant main or interaction effects (all p values > 0.10).

Legend of tables

Table 1. Patients anthropometric characteristics and surgery related parameters. (CPB: cardiopulmonary bypass)

	TIVA group (n: 11)	TIVA-DEX group (n: 12)
Age (years)	65 ± 8	60 ± 6
Body weight (kg)	74 ± 13	77 ± 15
Height (cm)	164 ± 8	165 ± 10
Gender (female-male)	4- 7	4- 8
Diabetes mellitus	4	6
Tabaco	4	9
Hypertensive disease	10	12
CPB time (min)	94 ± 22	89 ± 25
Surgery time (min)	299 ± 34	324 ± 35
Time for extubation (hour)	14 ± 3	14 ± 4
Intensive care unit time (days)	4 ± 1	4 ± 1
In-hospital stay (days)	7 ± 1	8 ± 2
Mini mental state examination scores (before surgery)	23 ± 3	23 ± 3
Mini mental state examination scores (5 days after surgery)	22 ± 3	22 ± 4

Legend of tables

Table 2. Perioperative haemodynamics parameters. HR: heart rate; MAP: mean arterial pressure; CI: cardiac index; CVP: central venous pressure; PO₂: arterial oxygen pressure; PCP: pulmonary capillary pressure.

GROUPS	TIVA group(n: 11)				TIVA-DEX group (n: 12)			
	Time 1	Time 2	Time 3	Time 4	Time 1	Time 2	Time 3	Time 4
HR (bpm)	65± 9	64±10	74±9	79±8	63±7	64±8	71±12	77±13
MAP (mmHg)	87±14	69±19	66±13	63±19	87±15	78±14	70±14	71±16
CI (L.min ⁻¹)	-	6.0±3.0	4.1±1.4	5.3±1.4	-	5.5±2.5	5.0±1.4	5.6±1.1
CVP (mmHg)	9.0±0.0	8.5±3.3	8.6±4.0	10.3±2.7	10.0±0.0	10.2±2.6	8.2±3.9	10.8±3.3
PO ₂ (mmHg)	164±42	213±54	146±39	81±32	164±32	166±55	124±38	85±16
PCP (mmHg)	-	16.5±6.6	14.4±7.5	16.2±5.6	-	16.2±6.2	14.1±4.3	19.1±6.3

Table 3. Hemodilution of patients in collected times. HT: hematocrit; Hb: hemoglobin.

	TIVA group (n: 11)				TIVA-DEX group (n: 12)			
Time	Time 1	Time 2	Time 3	Time 4	Time 1	Time 2	Time 3	Time 4
HT(%)	39.1±5.1	24.8±4.2	30.2±3.6	28.1±8.2	37.9±4.1	25.7±6.6	31.7±9.1	32.3±1.5
Hb(g/dL)	13.3±1.8	8.5±1.6	10.9±1.6	10.1±2.3	12.6±1.5	9.1±2.2	11.4±1.6	12.3±4.4

Legend of tables

Table 4. TBARS and δ-ALA-D activity in collected times. TIVA: total intravenous anesthesia; DTT: dithiothreitol reagent.

	Baseline	After CBP		
		1,5 h	5 h	24 h
δ-ALA-D activity (nmol PBG/h/mg protein)				
TIVA	429.5 ± 341.9	294.5 ± 234.9	367.0 ± 263.1	308.2 ± 249.7
TIVA-DEX	322.7 ± 274.7	278.0 ± 177.7	373.5 ± 237.8	349.0 ± 453. 6
δ-ALA-D activity with DTT (nmol PBG/h/mg protein)				
TIVA	425.0 ± 378.7	299.0 ± 211.5	439.9 ± 342.8	291.6 ± 265.1
TIVA-DEX	349.1 ± 203.0	327.2 ± 167.9	391.9 ± 227.9	370.2 ± 401.8
TBARS levels (nmolMDA/ml erythrocytes)				
TIVA	8.0 ± 2.0	8.8 ± 1.9	9.1 ± 1.8	9.4 ± 1.2
TIVA-DEX	8.9 ± 1.8	10.1 ± 2.2	10.7 ± 2.0	11.6 ± 2.1

3 DISCUSSÃO

O uso da dexmedetomidina como coadjuvante em anestesia ainda tem sido tímido e incipiente. Sua administração pode ter efeitos benéficos importantes em relação à proteção cerebral peri-operatória (MA e cols., 2005) e na atenuação da resposta inflamatória ao estresse induzido pela cirurgia (NADER e cols., 2001; VENN e cols., 2001; TANIGUCHI e cols., 2004; MEMIS e cols., 2007; TANIGUCHI e cols., 2008), sendo interessante explorar melhor os seus efeitos farmacológicos. A dexmedetomidina é um agonista (α)-2-adrenérgico, com grande seletividade para os receptores (α)-2/ (α)-1, na proporção de 1.620:1 e diferentes receptores (α)-2-adrenérgicos são responsáveis por seus efeitos clínicos específicos (TAKADA e cols., 2002; MA e cols., 2004; MOURA e cols., 2006; FAGERHOLM e cols., 2008; FAGERHOLM e cols., 2008). Também se liga a receptores imidazolínicos que modulam a pressão arterial sistêmica, possuem efeitos anti-arrítmicos (KHAN e cols., 1999) e têm sido implicados na sua capacidade de neuroproteção e geração de memória em modelos de isquemia cerebral em animais (TAKAMATSU e cols., 2008).

O principal resultado deste estudo clínico foi o de que a dexmedetomidina como componente da anestesia intravenosa total (AIVT) foi capaz de modular a resposta inflamatória em cirurgias de revascularização miocárdica (CRM) sob mini-circulação extracorpórea (mini-CEC). Os pacientes apresentaram menores níveis plasmáticos das citocinas pró-inflamatórias (IL-1, IL-6, TNF- α e INF- γ), se comparados ao grupo de pacientes que receberam AIVT convencional. Em ambos os grupos estudados, de maneira semelhante, houve diminuição progressiva pós-operatória da citocina anti-inflamatória IL-10.

Vários autores publicaram resultados sobre o efeito da dexmedetomidina e outros agonistas (α)-2-adrenérgicos sobre as citocinas (STRAUB e cols., 1997; Taniguchi e cols., 2004; HOFER e cols., 2009) e sobre a produção de TNF- α pelos macrófagos (SZELENYI e cols., 2000; TANIGUCHI e cols., 2004). Taniguchi e colaboradores (TANIGUCHI e cols., 2004) demonstraram que a dexmedetomidina possui efeito inibidor sobre a liberação de citocinas na endotoxemia em ratos, provavelmente através da modulação da produção de citocinas pelos macrófagos e monócitos. Hofer e colaboradores (HOFER e cols., 2009) demonstraram que a administração prévia de clonidina ou dexmedetomidina em sepsis induzida foi capaz de promover significativamente a sobrevida, sendo acompanhada pela redução de IL-1 β , IL- 6 e TNF- α . Em nosso estudo, os pacientes não se apresentavam

sépticos, mas a cirurgia de revascularização miocárdica com circulação extracorpórea demanda uma grande resposta inflamatória, semelhante à SRIS. A dexmedetomidina não conseguiu anular o aumento das citocinas pró-inflamatórias, porém demonstrou grande capacidade em sua diminuição, se comparada à AIVT convencional.

Resultados semelhantes foram obtidos por Tasdogan e colaboradores (TASDOGAN e cols., 2009) que compararam os efeitos de uma infusão de propofol e de dexmedetomidina na resposta inflamatória e pressão abdominal em pacientes com quadro séptico após cirurgia abdominal. A dexmedetomidina foi capaz de reduzir significativamente os níveis de TNF- α , IL-1 e IL-6 e a pressão intra-abdominal se comparado ao grupo que recebeu infusão de propofol. Bekker e colaboradores (BEKKER e cols., 2012), investigaram se a administração intra-operatória de dexmedetomidina poderia reduzir a resposta ao estresse e promover a qualidade na recuperação dos pacientes após cirurgia espinhal de grande porte. Eles compararam um grupo de pacientes anestesiados com propofol/fentanil/dexmedetomidina com outro recebendo propofol/fentanil/placebo-salina. A dexmedetomidina reduziu os níveis de cortisol e IL-10 se comparada ao grupo controle, mas não afetou IL-6 e IL-8. Clinicamente, os pacientes que receberam dexmedetomidina mostraram uma maior qualidade de recuperação.

Sukegawa e colaboradores (SUKEGAWA e cols., 2011) descreveram o efeito inibitório da dexmedetomidina sobre as reações inflamatórias, como o edema, o acúmulo de células inflamatórias, a produção de TNF- α e ciclooxygenase-2 (COX-2) induzidos pela injeção de carragenina em pata de camundongo. Outro estudo em modelo animal, também demonstrou a capacidade anti-inflamatória da dexmedetomidina em altas doses na inibição da resposta inflamatória por macrófagos induzidos por endotoxinas (LAI e cols., 2009). Peng e colegas (PENG e cols., 2013) relataram que a dexmedetomidina mostrou-se um potente inibidor da inflamação induzida por lipopolissacarídeos em micróglio ativada, podendo ser um agente terapêutico potencial no tratamento de *delirium* em unidades de terapia intensiva. As concentrações de dexmedetomidina utilizadas foram 1, 10 e 100 vezes as concentrações clinicamente relevantes (i.e., 1, 10, e 100 ng. mL $^{-1}$). Foram medidos a óxido nítrico sintase (iNOS), o óxido nítrico (NO), a prostaglandina E2, a IL-1 β e o TNF- α . A dexmedetomidina na dosagem de 1 ng. mL $^{-1}$ não afetou a produção de mediadores pró- inflamatórios, mas a 10 e 100 ng. mL $^{-1}$, a dexmedetomidina inibiu significativamente a liberação de óxido nítrico, prostaglandina E2, IL-1 β e TNF- α . A dose de infusão de dexmedetomidina que utilizamos em nosso estudo pode ser considerada de baixa a moderada, e mesmo assim foi capaz de reduzir

os níveis plasmáticos de IL-1, IL-6, TNF- α e INF- γ nos pacientes, se comparados ao grupo que não recebeu dexmedetomidina.

Arslan e colaboradores (2012), em modelo animal de isquemia hepática, demonstraram que a dexmedetomidina, administrada antes da indução da isquemia, protegeu contra a peroxidação lipídica (ARSLAN e cols., 2012). Yagmurdur e colaboradores (YAGMURDUR e cols., 2008) avaliaram o efeito da dexmedetomidina em lesões por isquemia/reperfusão devido ao uso de torniquete durante cirurgia em membro superior pela determinação dos níveis sanguíneos de malondialdeído e hipoxantina. A dexmedetomidina atenuou significativamente os níveis plasmáticos de hipoxantina na isquemia e a produção de malondialdeído no período de reperfusão. Eles sugerem que a dexmedetomidina pode ter a vantagem de inibir a peroxidação lipídica em caso de uso antecipado ao período de isquemia/reperfusão, como no caso descrito. Em cirurgia de revascularização miocárdica sob CEC, ocorre aumento progressivo dos níveis plasmáticos de proteína C reativa (PCR) e de TBARS (MELEK e cols., 2012) que denota claramente a ocorrência de inflamação e estresse oxidativo. Esta resposta inflamatória relacionada à CEC tem o potencial para produzir manifestações clínicas, bioquímicas e radiológicas de disfunções orgânicas (DHALLA e cols., 2000; MAULIK e YOSHIDA, 2000; RAJAS e BERG, 2007) que são o resultado do desequilíbrio entre a formação de espécies reativas de oxigênio e a capacidade antioxidante endógena. O uso da dexmedetomidina em cirurgia de revascularização miocárdica em nosso estudo, não conseguiu modificar os níveis de TBARS e da atividade da δ -ALA-D dos pacientes. Anteriormente, porém, em estudo de nosso grupo de pesquisa, observamos em pacientes submetidas à videolaparoscopia pélvica cirúrgica, o potencial efeito da dexmedetomidina (ROCHA e cols., 2010) na proteção da enzima delta-aminolevulinato dehidratase (δ -ALA-D) do sangue de ser inativada pela hiperoxigenação em anestesia intravenosa total.

A profundidade anestésica em nosso estudo foi monitorizada pelo uso do índice biespectral (BIS) e não foi o nosso objetivo verificar o consumo total de propofol e sufentanil, embora, aparentemente, houvesse uma redução nas doses necessárias dos mesmos quando associados ao uso da dexmedetomidina, para manter os níveis do BIS entre 45-55. A titulação do uso de concentrações menores do propofol através da monitorização pelo BIS, em estudos anteriores, associado a narcóticos potentes não alterou por si só os níveis de mediadores de estresse oxidativo nos pacientes estudados (BAUER e cols., 2004), o que poderia se supor, uma vez que o propofol tem efeito antioxidantante.

Em ambos os grupos estudados, de maneira similar, ocorreu aumento importante da PCR 24 horas após o término da cirurgia. Não houve diferenças entre os grupos quanto aos níveis de CPK-MB, cTnI e cortisol plasmático. Em estudos em animais, a dexmedetomidina inibiu a síntese de cortisol se usada em concentrações supraterapêuticas, mas este efeito supressor não tem sido relatado em usos de curta duração em humanos (MAZE e cols., 1991; VENN e cols., 2001; BULOW e cols., 2007), com doses de até $1.5 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ (GERLACH e cols., 2009) e por até dois dias (PANDHARIPANDE e cols., 2007). No presente estudo, reforçamos estas conclusões prévias de que a dexmedetomidina não leva à supressão da esteroidogênese.

Embora houvesse significativa diferença nas concentrações plasmáticas de citocinas pró-inflamatórias entre os grupos estudados, não houve correlação das mesmas com diferenças na evolução clínica dos pacientes. Chen e colaboradores (CHEN e cols., 2013), usando o mini-exame do estado mental (MEEM), demonstraram em pacientes submetidos à videocolangiocolite, que os déficits cognitivos para o grupo recebendo dexmedetomidina se comparado ao grupo controle uma semana após a cirurgia foram significativamente diferentes ($P=0.005$), com melhores escores no grupo da dexmedetomidina, sugerindo um efeito neuroprotetor em humanos. Não conseguimos demonstrar diferenças clínicas significativas entre os grupos estudados pela avaliação do MEEM, sendo os escores para os pacientes de ambos os grupos semelhantes se comparados os valores antes da cirurgia e cinco dias após a mesma. Há aumento de evidências de que a dexmedetomidina possui efeito protetor sobre os órgãos expostos à isquemia, promovendo cardio, neuro e renoproteção (HALL e cols., 2000). Além disso, a dexmedetomidina possui propriedades analgésicas que a tornam uma droga vantajosa em várias situações clínicas. Quando usada associada a outros anestésicos, pode reduzir tanto a concentração alveolar mínima dos anestésicos inalatórios, bem como de opióides, em até 90% das suas doses habituais (AHO e cols., 1992).

As doses intravasculares de dexmedetomidina induzem a redução dose-dependente da pressão arterial sistólica e diastólica e da frequência cardíaca, com importante diminuição nos níveis plasmáticos de norepinefrina. Contudo, doses elevadas, em *bolus* (50–75 µg), podem levar a uma hipertensão transitória inicial, devido à ativação dos receptores (α)-2-adrenérgicos periféricos antes que ocorram os efeitos simpatolíticos centrais sobre o centro vasomotor (TALKE e cols., 1997). Evitamos o uso da dose inicial em *bolus*, justamente por este motivo, e o grupo de pacientes manteve-se hemodinamicamente estável, semelhante ao grupo sem dexmedetomidina, observando-se apenas uma redução da frequência cardíaca e pressão arterial média com o passar do tempo, como já era esperado. A laringoscopia e intubação oro-

traqueal provocam resposta simpática marcante que aumenta o risco de isquemia miocárdica perioperatória e infarto do miocárdio. O uso da dexmedetomidina melhora a perfusão endocárdica e diminui a frequência cardíaca com atenuação da resposta clínica ao estresse ao qual o paciente é submetido (SULAIMAN e cols., 2012). A dexmedetomidina promove a estabilidade hemodinâmica pela modulação da resposta simpático-adrenal ao estresse da intubação e extubação oro-traqueal (SCHEININ e cols., 1992) o que se reflete em melhor evolução clínica. Estudos futuros, com maior número de pacientes, são desejáveis para melhor avaliar a influência da dexmedetomidina e a relevância da redução dos biomarcadores inflamatórios, induzida por ela, sobre a evolução clínica dos pacientes.

4 CONCLUSÕES

Buscam-se meios efetivos para alterar a resposta inflamatória em cirurgias de grande porte, especialmente as cirurgias de revascularização miocárdica usando circulação extracorpórea, objetivando reduzir as complicações peri-operatórias. Acreditamos que o uso da dexmedetomidina possa ser especialmente promissor. Neste estudo demonstramos a capacidade da dexmedetomidina de reduzir significativamente os níveis plasmáticos de citocinas pró-inflamatórias, como IL-1, IL-6, TNF- α , INF- γ , em pacientes submetidos a cirurgia de revascularização miocárdica sob mini-circulação extracorpórea, se comparados aos pacientes que não a receberam.

MANUSCRITO 1

A busca por resultados de um novo estado da arte no uso da dexmedetomidina em anestesia na literatura reforça a idéia de sua potencialidade ainda pouco explorada, na proteção dos órgãos durante períodos de agressão tecidual, como acontece no perioperatório. O seu uso ainda não se tornou rotineiro. Acreditamos ser a dexmedetomidina uma droga de múltiplas possibilidades, cada vez mais confirmadas através de estudos em animais e humanos, podendo promover evolução clínica mais adequada àqueles pacientes submetidos à cirurgia.

MANUSCRITO 2

Neste estudo demonstramos a capacidade da dexmedetomidina em reduzir os níveis plasmáticos de citocinas pró-inflamatórias, como IL-1, IL-6, TNF- α , INF- γ , em pacientes submetidos à cirurgia de revascularização miocárdica sob mini-circulação extracorpórea, se comparados aos pacientes que não receberam dexmedetomidina. Reforçamos o potencial benéfico da dexmedetomidina como anestésico com efeitos moduladores sobre a resposta inflamatória.

REFERÊNCIAS

- AANTAA, R., JALONEN, J. Perioperative use of alpha2-adrenoceptor agonists and the cardiac patient. **Eur J Anaesthesiol.** 23, 361- 372, 2006.
- AHO, M., ERKOLA, O., KALLIO, A., SCHEININ, H., KORTTILA, K. Dexmedetomidine infusion for maintenance of anesthesia in patients undergoing abdominal hysterectomy. **Anesth Analg.** 75, 940- 6, 1992.
- AOKI, H., MIZOBE, T., NOZUCHI, S., HIRAMATSU, N. *In vivo* and *in vitro* studies of the inhibitory effect of propofol on human platelet aggregation. **Anesthesiology.** 88, 362-70, 1998.
- ARCANGELI, A., D'ALO, C., GASPARI, R. Dexmedetomidine use in general anaesthesia. **Curr Drug Targets.** 10, 687- 95, 2009.
- ARSLAN, M., ÇOMU, F.M., KUÇUK, A., OZTURK, L., YAYLAK, F. Dexmedetomidine protects against lipid peroxidation and erythrocyte deformability alterations in experimental hepatic ischemia reperfusion injury. **Libyan J Med.** 7, 18185, 2012.
- ASCIONE, R., LLOYD, C.T., UNDERWOOD, M.J., LOTTO, A.A., PITIS, A.A., ANGELINI, G.D. Inflammatory response after coronary revascularization with or without cardiopulmonary bypass. **Ann Thorac Surg.** 69, 1198- 204, 2000.
- ASIMAKOPOULOS, G., GOURLA, Y. T. A review of anti-inflammatory strategies in cardiac surgery. **Perfusion.** 18, 7- 12, 2003.
- BAUER, M., WILHELM, W., KRAEMER, T., KREUER, S., BRANDT, A., ADAMS, H.A., HOFF, G., LARSEN, R. Impact of Bispectral Index monitoring on stress response and propofol consumption in patients undergoing coronary artery bypass surgery. **Anesthesiology.** 101, 1096-1104, 2004.
- BARAK, M., KATZ, Y. Microbubbles: pathophysiology and clinical implications. **Chest.** 128, 2918- 32, 2005.
- BEKKER, A., HAILE, M., KLINE, R., DIDEHVAR, S., BABU, R., MARTINIUK, F., URBAN, M. The effect of intraoperative infusion of dexmedetomidine on the quality of recovery after major spinal surgery. **J Neurosurg Anesthesiol.** 2012.

BERLIN, A., SCHALLER, K.H. European standardized method for determination of delta-aminolevulinic-acid dehydratase activity in blood. **Z Klin Chem Klin Biochem.** 12, 389-390, 1974.

BICAL, O.M., FROMES, Y., GAILLARD, D., FISCHER, M., PONZIO, O., DELEUZE, P., GERHARDT, M.F., TRIVIN, F. Comparison of the inflammatory response between miniaturized and standard CPB circuits in aortic valve surgery. **Eur Jour of Cardio-thoracic Surg.** 29, 699- 702, 2006.

BOLLI, R., PATEL, B.S., JEROUDI, M.O. Demonstration of free radical generation in "stunned" myocardium of intact dogs with the use of the spin trap alpha-phenyl N-tert-butyl nitrone. **J Clin Invest.** 82, 476- 85, 1988.

BRUDA, N.L., HURLBERT, B.J., HILL, G.E. Aprotinin reduces nitric oxide production in vitro and in vivo in a dose-dependent manner. **Clin Sci.** 94, 505- 509, 1998.

BULOW, N.M.H., BARBOSA, N.B.V., ROCHA, J.B.T. Opioid consumption in total anesthesia is reduced with dexmedetomidine: a comparative study with remifentanil in gynecologic videolaparoscopic surgery. **J Clin Anesth.** 19, 280- 5, 2007.

BUROW, B.K., JOHNSON, M.E., PACKER, D.L. Metabolic acidosis associated with propofol in the absence of other causative factors. **Anesthesiology.** 101, 239- 41, 2004.

BUTLER, J., ROCKER, G.M., WESTABY, S. Inflammatory response to cardiopulmonary bypass. **Ann Thorac Surg.** 55, 552- 9, 1993.

CANTACUZENE, J. Nouvelles recherches sur le monde de destruction des vibrios dans l'organisme. **Ann Inst Pasteur.** 12, 273- 300, 1898.

CARTIER, R., BOUCHOUT, O., EL-HAMAMSY, I. Influence of sex and age on long-term survival in systematic off-pump coronary artery bypass surgery. **Eur J Cardiothorac Surg.** 34, 826- 32, 2008.

CASEY, L.C. Role of the cytokines in the pathogenesis of cardiopulmonary-induced multisystem organ failure. **Ann Thorac Surg.** 56, S92- 6, 1993.

CERQUEIRA, N.F., YOSHIDA, W.B. Óxido nítrico. Revisão. **Acta Cir Bras.** 17, 417- 42, 2002.

CHANAY, M.A., DURAZO-ARVIZU, R.A., NIKOLOV, M.P., BLAKEMAN, B.P., BAKHOS, M. Methylprednisolone does not benefit patients undergoing coronary artery bypass grafting and early tracheal extubation. **Thorac Cardiovasc Surg.** 121, 561- 569, 2001.

CHEN, J., YAN, J., HAN, X. Dexmedetomidine may benefit cognitive function after laparoscopic cholecystectomy in elderly patients. **Experimental and Therapeutic Medicine.** 5, 489- 494, 2013.

CHRISTEN, S., FINCKH, B., LYKKEFELDT, J., GESSLER, P., FRESE-SCHAPER, M., NIELSEN, P., SCHMID, E.R., SCHMITT, B. Oxidative stress precedes peak systemic inflammatory response in pediatric patients undergoing cardiopulmonary bypass operation. **Free Radic Biol Med.** 38, 1323- 1332, 2005.

CORCORAN, T.B., ENGEL, A., SAKAMOTO, H., O'SHEA, A., O'CALLAGHAN-ENRIGHT, S., SHORTEN, G.D. The effects of propofol on neutrophil function, lipid peroxidation and inflammatory response during elective coronary artery bypass grafting in patients with impaired ventricular function. **Br J Anaesth.** 97, 825- 831, 2006.

CROZIER, T.A., MULLER, J.E., QUITTKA,T. D. Effect of anaesthesia on the cytokine responses to abdominal surgery. **Br J Anaesth.** 72, 280- 5, 1994.

DE LA CRUZ, J.P., CARMONA, J.A., PAEZ, M.V., BLANCO, E., SANCHEZ, D., DE LA CUESTA, F. Propofol inhibits in vitro platelet aggregation in humanwhole blood. **Anesth Analg.** 84, 919- 21, 1997.

DE MOURA, H.V., POMERANTZEFF, P.M.A., GOMES, W.J. Síndrome da resposta inflamatória sistêmica na circulação extra- corpórea: papel das interleucinas. **Rev Bras Cir Cardiovasc.** 16, 376- 387, 2001.

DE SOUZA, M.H.L., ELIAS, D.O. Fundamentos da circulação extracorpórea. 2^a ed., 2006.

DE VROEGE, R., STOOKER, W., VAN OEVEREN, W., BAKKER, E.W., HUYBREGTS, R. A.J.M., VAN KLARENBOSCH, J., VAN KAMP, G. J., HACK, C. E., EIJSMAN, L., WILDEVUUR, C. The impact of heparin-coated circuits upon metabolism in vital organs: Effect upon cerebral and renal function during and after cardiopulmonary bypass. **ASAIO J.** 51, 103-109, 2005.

DHALLA, N.S., ELMOSELHI, A.B., HATA, T., MAKINO, N. Status of myocardial antioxidants in ischemia-reperfusion injury. **Cardiovasc Res.** 47, 446- 56, 2000.

ELAHI, M.M., COURTNEY, J.M., MATATA, B.M. The interaction between reactive oxygen species and proinflammatory cytokines in human blood during extracorporeal circulation. **Filtration.** 1, 89- 94, 2005.

ELAHI, M.M., KHAN, J.S., MATATA, B.M. Deleterious effects of cardiopulmonary bypass in coronary artery surgery and scientific interpretation of off-pump's logic. **Acute Cardiac Care.** 8,196- 209, 2006.

ELAHI, M.M., MATATA, B.M. 2006. Is there a role for free radicals in the systemic inflammatory reaction? **J Cardiothorac Ren Res.** 1,131- 133, 2006.

ELAHI, M.M., MATATA, B.M. Free radicals in blood: evolving concepts in the mechanism of ischemic heart disease. **Arch Biochem Biophys.** 450, 78- 88, 2006.

ELAHI, M.M., M., MATATA, B.M. Significance of Oxidants and Inflammatory Mediators in Blood of Patients Undergoing Cardiac Surgery. **Journ Cardioth Vasc Anesth.** 22, 455- 467, 2008.

ELAHI, M.M., MATATA, B.M., HAKIM, N.S. Quiescent interplay between inducible nitric oxide synthase and tumor necrosis factor-alpha: Influence on transplant graft vasculopathy in renal allograft dysfunction. **Exp Clin Transplant.** 4, 445- 450, 2006.

EPPINGER, M.J., WARD, P.A., BOLLING, S.F., DEEB, G.M. Regulatory effects of interleukine- 0 on long ischemia-reperfusion injury. **J Thorac Cardiovasc Surg.** 112, 1301- 6, 1996.

FAGERHOLM V., ROKKA J., NYMAN L., SALLINEN, J., TIIHONEN, J., TUPALA, E., HAAPARANTA, M., HIETALA, J. Autoradiographic characterization of alpha (2C)-adrenoceptors in the human striatum. **Synapse.** 62, 508- 515, 2008.

FAGERHOLM V., SCHEININ M., HAAPARANTA M. Alpha2A-adrenoceptor antagonism increases insulin secretion and synergistically augments the insulinotropic effect of glibenclamide in mice. **Br J Pharmacol.** 154, 1287- 1296, 2008.

FLIERL, M.A., RITTIRSCH, D., NADEAU, B.A., CHEN, A.J., SARMA, J.V., ZETOUNE, F.S., MCGUIRE, S.R., LIST, R.P., DAY, D.E., HOESEL, L.M., GAO, H., VAN ROOIJEN, N., HUBER-LANG, M.S., NEUBIG, R.R., WARD, P.A. Phagocyte-derived catecholamines enhance acute inflammatory injury. **Nature.** 449, 721- 5, 2007.

FRANCISCHETTI, I., MORENO, J.B., SCHOLZ,M., YOSHIDA, W.B. Leukocytes and the inflammatory response in ischemia-reperfusion injury. **Rev Bras Cir Cardiovasc.** 25, 575-584, 2010.

FRANGOGIANNIS, N.G., LINDSEY, M.L., MICHAEL, L.H. Resident cardiac mast cells degranulate and release performed TNF-alpha, initiating the cytokine cascade in experimental canine myocardial ischemia/reperfusion. **Circulation.** 98, 699- 710, 1998.

FRANGOGIANNIS, N.G., YOUNK, K.A., ROSEN, R.D. Cytokines and the microcirculation in ischemia and reperfusion. **J Mol Cell Cardiol.** 30, 2567- 76, 1998.

FRANKE, A., LANTE,W., FACKELDEY, V., BECKER, H.P., KURIG, E., ZOLLER, L.G.,WEINHOLD, C., MARKEWITZ, A. Pro-inflammatory cytokines after different kinds of cardiothoracic surgical procedures: is what we see what we know? **Eur J Cardiothorac Surg.** 28, 569- 75, 2005.

FRANKE, A., LANTE, W., FACKELDEY, V., BECKER, H.P., THODE, C. KUHLMANN, W.D., MARKEWITZ, A. Proinflammatory and antiinflammatory cytokines after cardiac operation: Different cellular sources at different times. **Ann Thorac Surg.** 74, 363- 370, 2002.

FROMES, Y., GAILLARD, D., PONZIO, O., CHAUFFERT, M., GERHARDT, M.F., DELEUZE, P., BICAL, O.M. Reduction of the inflammatory response following coronary bypass grafting with total minimal extracorporeal circulation. **Eur J Cardiothorac Surg.** 22, 527- 33, 2002.

GALLETTI, P.M. AND BRECHER, G.A. Hear-Lung Bypass. Principles And Techniques Of Extracorporeal Circulation. Grune And Stratton, New York, 1962.

GALLEY HF, LOWE PR, CARMICHAEL RL, WEBSTER NR. Genotype and interleukin-10 responses after cardiopulmonary bypass. **Br J Anaesth.** 91: 424- 6, 2003.

GERLACH, A.T., MURPHY, C.V., DASTA, J.F. An Updated Focused Review of Dexmedetomidine in Adults. **The Annals of Pharmacotherapy.** Volume 43, 2009.

GETS, J., MONROY, F.P. Effects of alpha- and beta-adrenergic agonists on Toxoplasma gondii infection in murine macrophages. **J Parasitol.** 91, 93- 5, 2005.

GIBBON, J.H. JR. Application of a mechanical heart and lung apparatus to cardiac surgery. **Minn Med.** 37, 171- 85, 1954.

GIBBON, J.H.JR. The Development Of The Heart-Lung Apparatus. **Rev Surg.** 27, 231, 1970.

GIBBON, M.H. Recollections Of The Early Development Of The Heart-Lung Machine. Citado Por Litwak, R.S. – The Growth Of Cardiac Surgery. Historical Notes. In Cardiac Surgery . **Cardiovasc Clin.** 3, 5- 50, 1971.

GOGOS CA, DROSOU E, BASSARIS HP, SKOUTELIS A. Pro- versus anti-inflammatory cytokine profile in patients with severe sepsis: a marker for prognosis and future therapeutic options. **J Infect Dis.** 181: 176- 80, 2000.

GOMES, O.M., CONCEIÇÃO, D.S. Aparelho coração pulmão artificial. **Circulação extracorpórea.** 2a. Edição. Belo horizonte, 1985.

GONENC, A., HACISEVKI, A., BAKKALOGLU, B., SOYAGIR, A., TORUN, M., KARAGOZ, H., SIMSEK, B. Oxidative stress is decreased in off-pump versus on-pump coronary artery surgery. **J Biochem Mol Biol.** 39, 377- 382, 2006.

GOUDEAU, J.J., CLERMONT, G., GUILLERY, O., LEMAIRE-EWING, S., MUSAT, A., VERNET, M., VERGELY, C., GUIGUET, M., ROCHELLE, L., GIRARD, C. In high-risk patients, combination of antiinflammatory procedures during cardiopulmonary bypass can reduce incidences of inflammation and oxidative stress. **J Cardiovasc Pharmacol.** 49, 39- 45, 2007.

HALL ,J.E., UHRICH, T.D., BARNEY, J.A., ARAIN, S.R., EBERT, T.J. Sedative, amnestic, and analgesic properties of small-dose dexmedetomidine infusions. **Anesth Analg.** 90, 699- 705, 2000.

HANNAN, E.L., WU, C., RYAN, T.J., BENNETT, E., CULLIFORD, A.T., GOLD, J.P., HARTMAN, A., ISOM, O.W., JONES, R.H., MCNEIL, B., ROSE, E.A., SUBRAMANIAN,V.A. Do hospital and surgeons with higher coronary artery bypass graft surgery volumes still have lower risk-adjusted mortality rates? **Circulation.** 108, 795- 801, 2003.

HELLER, A., HELLER, S., BLECKEN, S., URBASCHEK, R., KOCH, T. Effects of intravenous anesthetics on bacterial elimination in human blood in vitro. **Acta Anaesthesiol Scand.** 42, 518- 26, 2008.

HEYER, E.J., LEE,K.S., MANSPEIZER, H.E., MONGERO, L., SPANIER, T.B., CALISTE, X., ESRIG, B., SMITH, C. Heparin-bonded cardiopulmonary bypass circuits reduce cognitive dysfunction. **J Cardiothorac Vasc Anesth.** 16, 37- 42, 2002.

HERR, D.L., SUM-PING, S.T., ENGLAND, M. ICU sedation after coronary artery bypass graft surgery: dexmedetomidine-based versus propofol-based sedation regimens. **J Cardiothorac Vasc Anesth.** 17, 576- 584, 2003.

HILL, G.E., DIEGO, R.P., STAMMERS, A.H., HUFFMAN, S.M., POHOREKI, R. Aprotinin enhances the endogenous release of interleukin-10 after cardiac operations. **Ann Thorac Surg.** 65, 66- 69, 1998.

HOFER, S., STEPPAN, J., WAGNER, T., FUNKE, B., LICHTENSTERN, C., MARTIN, E., GRAF, B.M., BIERHAUS, A., WEIGAND, M.A. Central sympatholytics prolong survival in experimental sepsis. **Critical Care.** 13, R11, 2009.

HUBER, A.R., KUNKEL, S.L., TODD, R.H., WEISS, S.J. Regulation of transendothelial neutrophil migration by endogenous interleukin-8. **Science.** 254, 99–102, 1991.

JALONEN, J., HYNNEN, M., KUITUNEN, A., HEIKKILÄ, H., PERTTILA, J., SALMENPERA, M., VALTONEN, M., AANTAA, R., KALLIO, A. Dexmedetomidine as anesthetic adjunct in coronary artery bypass grafting. **Anesthesiology.** 86, 331-345, 1997.

JANKE, E.L., SAMRA, S. Seminars in Anesthesia, **Perioperative Medicine and Pain** 25, 71- 76, 2006.

JESSEN, M.E. Heparin-coated circuits should be used for cardiopulmonary bypass. **Anesth Analg.** 103, 1365- 1369, 2006.

JOHNSON, D., THOMSON, D., HURST, T., PRASAD, K., WILSON, T., MURPHY, F., SAXENA, A., MAYERS, I. Neutrophil-mediated acute lung injury after extracorporeal perfusion. **J Thorac Cardiovasc Surg.** 107, 1193- 202, 1994.

JUNQUEIRA, L.C., CARNEIRO, C. **Histologia Básica.** 10^a ed. Rio de Janeiro. Guanabara Koogan, 223- 37, 2004.

KAHRAMAN, S., DEMIRYUREK, A.T. Propofol is a peroxynitrite scavenger. **Anesth Analg.** 84, 1127- 9, 1997.

KAM, P.C., CARDONE, D. Propofol infusion syndrome. **Anaesthesia.** 62, 690- 701, 2007.

KANG, W.-S., KIM, S.-Y., SON, J.-C., KIM, J.-D., MUHAMMAD, H.D., KIM, S.-H., YOON, T.-G., KIM, T.-Y. The effect of dexmedetomidine on the adjuvant propofol

requirement and intraoperative hemodynamics during remifentanil-based anesthesia. **Korean J Anesthesiol.** 62, 113-118, 2012.

KAPOOR, M.C., RAMACHANDRAN, T.R. Inflammatory response to cardiac surgery and strategies to overcome it. **Ann Card Anaesth.** 7,113- 128, 2004.

KAWAHITO, K., KOBAYASHI, E., OHMORI, M., HARADA, K., KITOH, Y., FUJIMURA, A., FUSE, K. Enhanced responsiveness of circulatory neutrophils after cardiopulmonary bypass: Increased aggregability and superoxide producing capacity. **Artif Organs.** 24,37- 42, 2000.

KEVIN, L.G., NOVALIJA, E., STOWE, D.F. Reactive Oxygen Species as Mediators of Cardiac Injury and Protection: The Relevance to Anesthesia Practice. **Anesth Analg.** 101, 1275- 87, 2005.

KHAN, Z.P., FERGUSON, C.N., JONES, R.M. Alpha-2 and imidazoline receptor agonists.Their pharmacology and therapeutic role. **Anesthesia.** 54, 146- 65, 1999.

KOTANI, N., HASHIMOTO, H., SESSLER, D.I., YASUDA, T., EBINA, T., MURAOKA, M., MATSUKI, A. Expression of genes for proinflammatory cytokines in alveolar macrophages during propofol and isoflurane anesthesia. **Anesth Analg.** 89, 1250- 6, 1999.

KRUMHOLZ, W., ENDRASS, J., HEMPELMANN, G. Propofol inhibits phagocytosis and killing of *Staphylococcus aureus* and *Escherichia coli* by polymorphonuclear leukocytes in vitro. **Can J Anaesth.** 41, 446- 9, 1994.

KUTAY, V., NOYAN, T., OZCAN, S., MELEK, Y., EKIM, H., YAKUT, C. Biocompatibility of heparincoated cardiopulmonary bypass circuits in coronary patients with left ventricular dysfunction is superior to PMEA-coated circuits. **J Card Surg.** 21, 572- 577, 2006.

KUYPERS, F.A. Red cell membrane damage. **J Heart Valve Dis.** 7, 387- 95, 1998.

LAI, Y.C., TSAI, P.S., HUANG, C.J. Effects of dexmedetomidine on regulating endotoxin-induced up-regulation of inflammatory molecules in murine macrophages. **J Surg Res.** 154, 212- 219, 2009.

LAUTH, C.I., SMITH, P.L., ARNOLD, J.V., Et Al. Influence of oxygenator type on the incidence and extent of microembolic retinal ischemia during cardiopulmonary bypass. **J Thorac Cardiovasc Surg.** 99, 61- 69, 1990.

LEVI, M., CROMHEECKE, M.E., DE JONGE, E., PRINS, M.H., DE MOL, B.J.M., BRIËT, E., BÜLLER, H.R. Pharmacological strategies to decrease excessive blood loss in cardiac surgery: A metaanalysis of clinically relevant endpoints. **Lancet.** 354, 1940- 1947, 1999.

LEVY, J.H., TANAKA, K.A. Inflammatory response to cardiopulmonary bypass. **Ann Thorac Surg.** 75, 715- 720, 2003.

LIAKOPoulos, O.J., SCHMITTO, J.D., KAZMAIER, S., BRAUER, A., QUINTEL, M., SCHOENDUBE, F.A., DORGE, H. Cardiopulmonary and systemic effects of methylprednisolone in patients undergoing cardiac surgery. **Ann Thorac Surg.** 84, 110- 118, 2007.

LI, S., PRICE, R., PHIROZ, D., SWAN, K., CRANE, T.A. Systemic inflammatory response during cardiopulmonary bypass and strategies. **J Extra Corpor Technol.** 37, 180- 188, 2005.

LIOLIOS, A., GUERIT, J.M., SCHOLTES, J.L., RAFTOPOULOS, C., HANTSON, P. Propofol infusion syndrome associated with short-term large-dose infusion during surgical anesthesia in an adult. **Anesth Analg.** 100, 1804- 6, 2005.

LYONS A, KELLY JL, RODRICK ML, ET AL. Major injury induces increased production of interleukin-10 by cells of the immune system with a negative impact on resistance to infection. **An Surg.** 226: 450- 8, 1997.

MADDEN, N.J., DEMARSICO, A.J., SCHOCKER, L.A., Et Al. On-pump vs. off-pump coronary artery bypass surgery at a non-academic community hospital: Have biocompatibility improvements eliminated the superiority of off-pump surgery? **Int J Artif Organs.** 30, 338- 344, 2007.

MA, D., HOSSAIN, M., RAJAKUMARASWAMY, N., ARSHAD, M., SANDERS, R.D., FRANKS, N.P., MAZE, M. Dexmedetomidine produces its neuroprotective effect via the α 2A-adrenoceptor subtype. **Eur J Pharmacol.** 502, 87- 97, 2004.

MA, D., RAJAKUMARASWAMY, N., MAZE, M. α 2-Adrenoreceptor agonists: shedding light on neuroprotection? **Br Med Bull.** 71, 77- 92, 2005.

MAIER, C., STEINBERG, G.K., SUN, G.H. Neuroprotection by the alpha2-adrenoreceptor agonist dexmedetomidine in a focal model of cerebral ischemia. **Anesthesiology.** 79, 306- 312, 1993.

MAQSOOD, M., ELAHI, M., BASHIR, M., MATATA, B. M. Significance of Oxidants and Inflammatory Mediators in Blood of Patients Undergoing Cardiac Surgery. *Journ Cardioth Vasc Anesth.* 22, 455- 467, 2008.

MARCZIN, N., EL-HABASHI, N., ROYSTON, D. Free radicals and cardiac arrhythmias following coronary surgery: actors of the drama or bystanders of the spectacle. *Acta Anaesthesiol Scand.* 47, 639- 4, 2003.

MATATA, B.M., GALINANES, M. Peroxynitrite is an essential component of cytokines production mechanism in human monocytes through modulation of nuclear factor-kappa B DNA binding capacity. *J Biol Chem.* 277, 2330- 2335, 2002.

MATATA, B.M., GALINANES, M. Cardiopulmonary bypass exacerbates oxidative stress but does not increase proinflammatory cytokine release in patients with diabetes compared with patients without diabetes: Regulatory effects of exogenous nitric oxide. *J Thorac Cardiovasc Surg.* 120,1-11, 2000.

MATATA, B.M., SOSNOWSKI, A.W., GALINANES, M. Off-pump bypass graft operation significantly reduces oxidative stress and inflammation. *Ann Thorac Surg.* 69, 785- 791, 2000.

MATSUMOTO, M., ZORNOW, M.H., RABIN, B.C., MAZE, M. The alpha 2-adrenergic agonist, dexmedetomidine, selectively attenuates ischemia-induced increases in striatal norepinephrine concentrations. *Brain Res.* 627, 325- 9, 1993.

MAULIK N, YOSHIDA T. Oxidative stress developed during open heart surgery induces apoptosis: reduction of apoptotic cell death by ebselen, a glutathione peroxidase mimic. *J Cardiovasc Pharmacol.* 36, 601- 8, 2000.

MAULIK, N., YOSHIDA, T., DAS, D.K. Oxidative stress developed during the reperfusion of ischemic myocardium induces apoptosis. *Free Radic Biol Med.* 24, 869- 75, 1998.

MAZE, M., VIRTANEN, R., DAUNT, D., BANKS, S.J., STOVER, E.P., FELDMAN, D. Effects of dexmedetomidine, a novel imidazole sedative-anesthetic agent, on adrenal steroidogenesis: in-vivo and in-vitro studies. *Anesth Analg.* 73, 204- 8, 1991.

MELEK, F.E., BARONCINI, L.A.V., REPKA, J.C.D., NASCIMENTO, C.S., BERTOLIM, D. Pré-coma. *Rev Bras Cir Cardiovasc.* 27, 61- 5, 2012.

MEMIS, D., HEKIMO_GLU S, VATAN I, YANDIM, T., YUKSEL, M., SUT, N. Effects of midazolam and dexmedetomidine on inflammatory responses and gastric intramucosal pH to sepsis, in critically ill patients. **Br J Anaesth.** 98, 550- 2, 2007.

MERZ, T.M., REGLI, B., ROTHEN, H.U., FELLEITER, P. Propofol infusion syndrome-- a fatal case at a low infusion rate. **Anesth Analg.** 103, 1050, 2006.

MILES, B.A., LAFUSE, W.P., ZWILLING, B.S. Binding of -adrenergic receptors stimulates the anti-mycobacterial activity of murine peritoneal macrophages. **J Neuroimmunol.** 71, 19-24, 1996.

MITCHELL, R.N., BEVILACQUA M.P. Endothelial-leukocyte adhesion molecules. **Ann Rev Immunol** 1993;11:767-804, 2006

MOURA, E., AFONSO, J., HEIN, L., VIEIRA-COELHO, M.A. Alpha2-adrenoceptor subtypes involved in the regulation of catecholamine release from the adrenal medulla of mice. **Br J Pharmacol.** 149, 1049- 58, 2006.

MURPHY, P.G., MYERS, D.S., DAVIES, M.J., WEBSTER, N.R., JONES, J.G. The antioxidant potential of propofol (2,6 -diisopropylphenol). **Br J Anaesth.** 68, 613- 8, 1992.

NADER, N.D., IGNATOWSKI, T.A., KUREK, C.J., KNIGHT, P.R., SPENGLER, R.N. Clonidine suppresses plasma and cerebrospinal fluid concentrations of TNF-alpha during the perioperative period. **Anesth Analg.** 93, 363- 9, 2001.

NANCE, D.M., SANDERS, V.M. Autonomic innervation and regulation of the immune system (1987–2007). **Brain Behav Immun.** 21, 736- 45, 2007.

NAVAPURKAR, V., SKEPPER, J., JONES, J., MENON, D. Propofol preserves the viability of isolated rat hepatocyte suspensions under an oxidant stress. **Anesth Analg.** 87, 1152- 7, 1998.

NIEMAN, G., SEARLES, B., CARNEY, D., MCCANN, U., SCHILLER, H., LUTZ, C. Systemic inflammation induced by cardiopulmonary bypass: a review of pathogenesis and treatment. **J Extra Corpor Technol.** 31, 202- 10, 1999.

OPAL, S.M. The host response to endotoxin, antilipopolysaccharide strategies, and management of severe sepsis. **Int J Med Microbiol.** 297, 365- 377, 2007.

OYAMA, J., SHIMOKAWA, H., MOMII, H., CHENG, X., FUKUYAMA, N., ARAI, Y., EGASHIRA, K., NAKAZAWA, H., TAKESHITA, A. Role of nitric oxide and peroxynitrite in the cytokine-induced sustained myocardial dysfunction in dogs in vivo. **J Clin Invest.** 101, 2207- 2214, 1998.

PANDHARIPANDE, P.P., PUN, B.T., HERR, D.L., MAZE, M., GIRARD, T.D., MILLER, R.R., SHINTANI, A.K., THOMPSON, J.L., JACKSON, J.C., DEPPEN, S.A., STILES, R.A., DITTUS, R.S., BERNARD, G.R., ELY, E.W. Effect of sedation with dexmedetomidine vs lorazepam on acute brain dysfunction in mechanically ventilated patients: the MENDS randomized controlled trial. **JAMA.** 298, 2644- 2653, 2007.

PANDHARIPANDE, P.P., SANDERS, R.D., GIRARD, T., Et Al. Comparison of sedation with dexmedetomidine versus lorazepam in septic ICU patients. **Critical Care.** 12, 275, 2008.

PEMBERTON, M.G., ANDERSON, G., VETVICKA, V., JUSTUS, D.E., ROSS, G.D. Microvascular effects of complement blockade with soluble recombinant CR1 on ischemia/reperfusion injury of skeletal muscle. **J Immunol.** 150, 5104- 5113, 1993.

PENG,M.,WANG,Y.-L.,WANG,C.-Y.,CHEN,C. Dexmedetomidine attenuates lipopolysaccharide-induced proinflammatory response in primary microglia. **Journ Surg Res.** 179, E219- E225, 2013.

PERRY, T.E., MUEHLSCHLEGEL, J.D., LIU, K.Y., FOX, A.A., COLLARD, C.D., BODY, S.C., SHERNAN, S.K. Preoperative C-reactive Protein Predicts Long-term Mortality and Hospital Length of Stay after Primary, Nonemergent Coronary Artery Bypass Grafting. **Anesthesiology.** 112, 607- 13, 2010.

PERTHEL, M., KSEIBI, S., SAGEBIEL, F., ALKEN, A., LAAS, J. Comparison of conventional extracorporeal circulation and minimal extracorporeal circulation with respect to microbubbles and microembolic signals. **Perfusion.** 20, 329- 333, 2005.

PIEGAS, L.P., BITTAR, O.J.N.V., HADDAD, N. Cirurgia de revascularização miocárdica. Resultados do Sistema Único de Saúde. **Arq Bras Cardiol.** 93, 555- 60, 2009.

PLOMONDON, M.E., CLEVELAND, J.C. JR., LUDWIG, S.T., GRUNWALD, G.K., KIEFE, C.I., GROVER, F.L., SHROYER, A.L. Off-pump coronary artery bypass is associated with improved risk-adjusted outcomes. **Ann Thorac Surg.** 72, 114- 119, 2001.

PUNTEL, R.L., ROOS, D.H., GROTTO, D., GARCIA, S.C., NOGUEIRA, C.W., ROCHA, J.B. Antioxidant properties of Krebs cycle intermediates against malonate pro-oxidant activity

in vitro: a comparative study using the colorimetric method and HPLC analysis to determine malondialdehyde in rat brain homogenates. **Life Sci.** 81, 51- 62, 2007.

PRYOR, W.A. Free radicals in autoxidation and in aging. Free Radicals in Molecular Biology, Aging and Disease. New York, Raven Press Publishers. 13- 41, 1984.

RAJAS, S.G., BERG, G.A. Impact of off-pump coronary artery bypass surgery on systemic inflammation: Current best available evidence. **J Card Surg.** 22, 445- 455, 2007.

REMADI, J.P., MARTICHO, P., BUTOI, I., RAKOTOARIVELO, Z. , TROJETTE, F., BENAMAR, A., BELOUCIF,S. , FOURE, D., HENRI, J. Clinical Experience With the Mini-Extracorporeal Circulation System: An Evolution or a Revolution? **Poulain Ann Thorac Surg.** 77, 2172- 6, 2004.

REMADI, J.P., RAKOTOARIVELO, Z., MARTICHO, P., BENAMAR, A. Prospective randomized study comparing coronary artery bypass grafting with the new mini-extracorporeal circulation Jostra System or with a standard cardiopulmonary bypass. **Am Heart J.** 151, 198, 2006.

RICHTER, J., NG-SIKORSKI, J., OLSSON, I., ANDERSON, T. Tumor necrosis factor-induced degranulation in adherent human neutrophils is dependent on CD11b/CD18-integrin-triggered oscillations of cytosolic free Ca²⁺. **Proc Natl Acad Sci USA.** 87, 9472- 6, 1990.

ROCHA, J.B.T., BULOW, N.M.H., CORREA, E.F.M., SCHOLZE, C., NOGUEIRA, C.W., BARBOSA, N.B.V. Dexmedetomidine protects blood d-aminolevulinate dehydratase from inactivation caused by hyperoxygenation in total intravenous anesthesia. **Hum Experim Toxicol.** 30, 289- 295, 2010.

RUBENS, F.D., NATHAN, H., LABOW, R., WILLIAMS, K.S., WOZNY, D., KARSH, J., RUEL, M., MESANA, T. Effects of methylprednisolone and a biocompatible copolymer circuit on blood activation during cardiopulmonary bypass. **Ann Thorac Surg.** 79, 655- 665, 2005.

SALENGROS, J.C., VELGHE-LENELLE, C.E., BOLLENS, R., ENGELMAN, E., BARVAIS, L. Lactic acidosis during propofol-remifentanil anesthesia in an adult. **Anesthesiology.** 101, 241- 3, 2004.

SABLOTZKI A, WELTERS I, LEHMANN N, ET AL. Plasma levels of immunoinhibitory cytokines interleukin-10 and transforming growth factor-beta in patients undergoing coronary artery bypass grafting. **Eur J Cardiothorac Surg.** 11: 763- 8, 1997.

SANDER, M., VON HEYMANN, C., VON DOSSOW, V., SPAETHE, C., KONERTZ, W.F., JAIN, U., SPIES, C.D. Increased Interleukin-6 After Cardiac Surgery Predicts Infection. **Anesth Analg.** 102, 1623- 9, 2006.

SANDERS, R.D., HUSSELL, T., MAZE, M. Sedation & Immunomodulation. **Anesthesiology Clin.** 29, 687- 706, 2011.

SANO, T., MORITA, S., MASUDA, M., TOMITA, Y., NISHIDA, T., TATEWAKI H., YASUI, H. Cardiopulmonary bypass, steroid administration, and surgical injury synergistically impair memory T cell function and antigen presentation. **Interact Cardiovasc Thorac Surg.** 2, 598- 602, 2003.

SATO, H., ZHAO, Z.Q., JORDAN, J.E., TODD, J.C., RILEY, R.D., TAFT, C.S., HAMMON JR, J.W., LI, P., MA, X.L., VINTEN-JOHANSEN, J. Basal nitric oxide expresses endogenous cardioprotection during reperfusion by inhibition of neutrophils- mediated damage after surgical revascularization. **J Thorac Cardiovasc Surg.** 113, 399- 409, 1997.

SAVARIS, N., POLANCZYK, C., CLAUSELL, N. Cytokines and Troponin-I in Cardiac Dysfunction After Coronary Artery Grafting with Cardiopulmonary Bypass. **Arq Bras Cardiol.** 77, 114- 9, 2001.

SCHEININ, B., LINDGREN, L., RANDELL, T., SCHEININ, H., SCHEININ, M. Dexmedetomidine attenuates sympathoadrenal responses to tracheal intubation and reduces the need for thiopentone and preoperative fentanyl. **Br J Anaesth.** 68, 126- 31, 1992.

SCHOLZ, J., TONNER, P.H. Alpha 2-adrenoreceptor agonists in anaesthesia: a new paradigm. **Curr Opin Anaesthesiol.** 13, 437- 442, 2000.

SEDRAKYAN, A., TREASURE, T., ELEFTERIADES, J.A. Effects of aprotinin on clinical outcomes in coronary artery bypass graft surgery: A systematic review and meta-analysis of randomized clinical trials. **J Thorac Cardiovasc Surg.** 128, 442- 448, 2004.

SHAPPELL, S.B., TOMAN, C., ANDERSON, D.C., TAYLOR, A.A., ENTMAN, M.L., SMITH, C.W. MAC-1(CD11b/CD18) mediates adherence dependent hydrogen peroxide production by human, and canine neutrophils. **J Immunol.** 144, 2702- 11, 1990.

SHROYER, L., GROVER, F.L., HATTLER, B., COLLINS, J.F., MCDONALD, G.O., KOZORA, E., LUCKE, J.C., BALTZ, J.H., NOVITZKY, D. On-Pump versus Off-Pump Coronary-Artery Bypass Surgery A. for the Veterans Affairs Randomized On/Off Bypass (ROOBY) Study Group. **N Engl J Med.** 361, 1827- 37, 2009.

SIMCHON, S., JAN, K.M., CHIEN, S. Influence of reduced red cell deformability on regional blood flow. **Am J Physiol.** 253, 898-903, 1987.

SIVIOTTI, M.L. Oxidant stress and haemolysis of the human erythrocyte. **Toxicol Rev.** 23, 169- 88, 2004.

SMITH, I.M., KENNEDY, L.R., REGNE'-KARLSSON, M.H., JOHNSON, V.L., BURMEISTER, L.F. Adrenergic mechanisms in infection. III. Alpha- and beta-receptor blocking agents in treatment. **Am J Clin Nutr.** 30, 1285- 8, 1977.

SPENGLER, R.N., ALLEN, R.M., REMICK, D.G., STRIETER R.M., KUNKEL, S.L. Stimulation of alpha-adrenergic receptor augments the production of macrophage-derived tumor necrosis factor. **J Immunol.** 145, 1430- 4, 1990.

SPIES, C.D., DUBISZ, N., NEUMANN, T., BLUM, S., MULLER, C., ROMMELSPACHER, H., BRUMMER, G., SPECHT, M., SANFT, C., HANNEMANN, L., STRIEBEL, H.W., SCHAFFARTZIK, W. Therapy of alcohol withdrawal syndrome in intensive care unit patients following trauma: results of a prospective, randomized trial. **Crit Care Med.** 24, 414- 422, 1996.

STAHL, G.L., XU, Y., HAO, L., MILLER, M., BURAS, J.A., FUNG, M., ZHAO, H. Role for the alternative complement pathway in ischemia/reperfusion injury. **Am J Pathol.** 162, 449- 455, 2003.

STEINBERG, J.B., KAPELANSKI, D.P., OLSON, J.D., WEILER, J.M. Cytokine and complement levels in patients undergoing cardiopulmonary bypass. **J Thorac Cardiovasc Surg.** 106, 1008- 16, 1993.

STERNBERG, E.M. Neural regulation of innate immunity: a coordinated nonspecific host response to pathogens. **Nat Rev Immunol.** 6, 318- 28, 2006.

STRAUB, R.H., HERRMANN, M., BERKMILLER, G., FRAUENHOLZ, T., LANG, B., SCHÖLMERICH, J., FALK, W. Neuronal regulation of interleukin 6 secretion in murine spleen: adrenergic and opioidergic control. **J Neurochem.** 68, 1633- 9, 1997.

SUD, R., SPENGLER, R.N., NADER, N.D., IGNATOWSKI, T.A. Antinociception occurs with a reversal in alpha 2-adrenoceptor regulation of TNF production by peripheral monocytes/ macrophages from pro- to anti-inflammatory. **Eur J Pharmacol.** 588, 217- 31, 2008.

SUKEGAWA, S., INOUE, M., HIGUCHI, H., TOMOYASU, Y., MAEDA, S., MIYAWAKI, T. Locally Injected Dexmedetomidine Inhibits Carrageenin-induced Inflammatory Reactions in Injected Region. **ASA meet.** A1590, 2011.

SULAIMAN, S., KARTHEKEYAN, R.B., VAKAMUNDI, M., SUNDAIR, A.S., RAVULLAPALLI, H., GANDHAN, R. The effects of dexmedetomidine on attenuation of stress response to endotracheal intubation in patients undergoing elective off pump coronary artery bypass grafting. **Ann Card Anaesth.** 15,1, 2012.

SULEMANJI, D.S., DONMEZ, A., ALDEMIR, D., SEZGIN, A., TURKOGLU, S. Dexmedetomidine during coronary artery bypass grafting surgery: is it neuroprotective? A preliminary study. **Acta Anaesth Scand.** 51, 1093- 1098, 2007.

SZELENYI, J., KISS, J.P., VIZI, E.S. Differential involvement of sympathetic nervous system and immune system in the modulation of TNF-alpha production by alpha2- and beta-adrenoceptors in mice. **J Neuroimmunol.** 103, 34- 40, 2000.

TAKAI, H., EISHI, K., YAMACHIKI, S., HAZAMA, S., ARIYOSHI, T., NISHI, K. Demonstration and operative influence of low prime volume closed pump. **Asian Cardiovasc Thorac Ann.** 13, 65- 69, 2005.

TAKADA, K., CLARK, D.J., DAVIES, M.F., TONNER, P.H., KRAUSE, T.K., BERTACCINI, E., MAZE, M. Meperidine exerts agonist activity at the alpha (2B)-adrenoceptor subtype. **Anesthesiology.** 96, 1420- 1426, 2002.

TAKAMATSU, I., IWASE, A., OZAKI, M., KAZAMA, T., WADA, K., SEKIGUCHI, M. Dexmedetomidine reduces long-term potentiation in mouse hippocampus. **Anesthesiology.** 108, 94- 102, 2008.

TALKE, P., RICHARDSON, C.A., SCHEININ, M., FISCHER, D.M. Postoperative pharmacokinetics and sympatholytic effects of dexmedetomidine. **Anesth Analg.** 85, 1136- 1142, 1997.

TANG, M., GU, Y.J., WANG, W.J., XU, Y.P., CHEN, C.Z. Effect of cardiopulmonary bypass on leukocyte activation: changes in membrane-bound elastase on neutrophils. **Perfusion.** 19, 93- 9, 2004.

TANIGUCHI, T., KIDANI, Y., KANAKURA, H., TAKEMOTO, Y., YAMAMOTO, K. Effects of dexmedetomidine on mortality rate and inflammatory responses to endotoxin-induced shock in rats. **Crit Care Med.** 32, 1322- 6, 2004.

TANIGUCHI, T., KURITA, A., KOBAYASHI, K., YAMAMOTO, K., INABA, H. Dose- and time-related effects of dexmedetomidine on mortality and inflammatory responses to endotoxin-induced shock in rats. **J Anesth.** 22, 221- 8, 2008.

TAN, J., MCCLUNG, W.G., BRASH, J.L. Nonfouling biomaterials based on polyethylene oxide-containing amphiphilic triblock copolymers as surface-modifying additives: Protein adsorption on PEO-copolymer/ polyurethane blends. **J Biomed Mater Res.** 85, 873-880, 2007.

TASDOGAN, M., MEMIS, D., SUT, N., YUKSEL, M. Results of a pilot study on the effects of propofol and dexmedetomidine on inflammatory responses and intra-abdominal pressure in severe sepsis. **J Clin Anesth.** 21, 394- 400, 2009.

THEROND, P., BONNEFONT-ROUSSELOT, D., DAVID-SPRAUL, A., CONTI, M., LEGRAND, A. Biomarkers of oxidative stress: an analytical approach. **Curr Opin Clin Nutr Metab Care.** 3, 373- 84, 2000.

TORRE-AMIONE, G., KAPADIA, S., LEE, J., BIES, R.D., LEBOVITZ, R., MANN, B.L. Expression and functional significance of tumor necrosis factor receptors in human myocardium. **Circulation.** 92, 1487- 93, 1995.

TRACEY, K.J. Physiology and immunology of the cholinergic anti-inflammatory pathway. **J Clin Invest.** 117, 289- 96, 2007.

VAN DER LINDEN, P.J., HARDY, J.F., DAPER, A., TRENCHANT, A., DE HERT, S.G. Cardiac surgery with cardiopulmonary bypass: Does aprotinin affect outcome? **Br J Anaesth.** 99, 646- 652, 2007.

VAN-DISSEL JT, VAN-LANGEVELDE P, WESTENDORP RG, ET AL. Antiinflammatory cytokine profile and mortality in febrile patients. **Lancet.** 351: 950- 3, 1998.

VENN, R.M., BRYANT, A., HALL, G.M., GROUNDS, R.M. Effects of dexmedetomidine on adrenal cortical function, and the cardiovascular, endocrine, and inflammatory responses in post-operative patients needing sedation in the intensive care unit. **Br J Anaesth.** 86, 650- 6, 2001.

WADA, K., MONTALTO, M.C., STAHL, G.L. Inhibition of complement C5 reduces local and remote organ injury after intestinal ischemia/reperfusion in the rat. **Gastroenterology.** 120, 126- 133, 2001.

WALSH, M.C., BOURCIER, T., TAKAHASHI, K., SHI, L., BUSHE, M.N., ROTHER, R.P., SOLOMON, S.D., EZEKOWITZ, R.A.B., ATAHL, G.L. Mannose-binding lectin is a regulator of inflammation that accompanies myocardial ischemia and reperfusion injury. **J Immunol.** 175, 541- 546, 2005.

WAN, S., LECLERC, J.L., HUYNH, C.H., SCHMARTZ, D., DESMET, M., YIM, A.P.C., VINCENT, J.L. Does steroid pretreatment increase endotoxin release during clinical cardiopulmonary bypass? **J Thorac Cardiovasc Surg.** 117, 1004- 1008, 1999.

WARREN, O.J., SMITH, A.J., ALEXIOU, C., ROGERS, P.L.B., JAWAD, N., VINCENT, C., DARZI, A.W., ATHANASIOU. T. The inflammatory response to cardiopulmonary bypass: part 1 – mechanisms of pathogenesis. **J Cardioth Vasc Anesth.** 23, 223- 231, 2009.

WARREN, O.J., WATRET, A.L., DE WIT, K.L., ALEXIOU, C., VINCENT, C., DARZI, A.W., ATHANASIOU, T. The inflammatory response to cardiopulmonary bypass: part 2- anti-inflammatory therapeutic strategies. **J Cardioth Vasc Anesth.** 23, 384- 393, 2009.

WARREN, O., ALEXIOU, C., MASSEY, R., LEFT, D., PURKAYASTHA, S., KINROSS, J., DARZI, A., ATHANASIOU, T. The effects of various leukocyte filtration strategies in cardiac surgery. **Eur J Cardiothorac Surg.** 31, 665- 676, 2007.

WEATHERBY, K.E., ZWILLING, B.S., LAFUSE, W.P. Resistance of macrophages to *Mycobacterium avium* is induced by alpha2-adrenergic stimulation. **Infect Immun.** 71, 22- 9, 2003.

WEINERT, C.R., KETHIREDDY, S., ROY, S. Opioids and infections in the intensive care unit should clinicians and patients be concerned? **J Neuroimmune Pharmacol.** 3, 218-29, 2008.

WEISMAN, H.F., BARTOW, T., LEPOO, M.K., MARSH, H.C. JR., CARSON, G.R., CONCINO, M.F., BOYLE, M.P., ROUX, K.H., WEISFELDT, M.L., FEARON, D.T. Soluble human complement receptor type 1: In vivo inhibitor of complement suppressing post-ischemic myocardial inflammation and necrosis. **Science.** 249, 146- 151, 1990.

WEST, J.P., DYKSTRA, L.A., LYSLE, D.T. Immunomodulatory effects of morphine withdrawal in the rat are time dependent and reversible by clonidine. **Psychopharmacology.** 146, 320- 7, 1999.

WIJEYSUNDERA, D.N., NAIK, J.S., BEATTIE, W.S. Alpha2-adrenergic agonists to prevent perioperative cardiovascular complications: a metaanalysis. **Am J Med.** 114, 742- 752, 2003.

XIA, Z., HUANG, Z., ANSLEY, D.M. Large-Dose Propofol During Cardiopulmonary Bypass Decreases Biochemical Markers of Myocardial Injury in Coronary Surgery Patients: A Comparison with Isoflurane. **Anesth Analg.** 103, 527 -532, 2006.

YANG, C.L., TSAI, P.S., HUANG, C.J. Effects of dexmedetomidine on regulating pulmonary inflammation in a rat model of ventilator-induced lung injury. **Acta Anaesthesiol Taiwan.** 46, 151- 9, 2008.

YAGMURDUR, H., OZCAN, N., DOKUMACI, F., KILINC, K., YILMAZ, F., BASAR, H. Dexmedetomidine Reduces the Ischemia-Reperfusion Injury Markers During Upper Extremity Surgery With Tourniquet. **J Hand Surg.** 33, 941- 947, 2008.

YOUNG, Y., MENON, D., TISAVIPAT. N. Propofol neuroprotection in a rat model of ischaemia reperfusion injury. **Eur J Anaesthesiol.** 14, 320- 6, 1997.

ZHAO, H., MONTALTO, M.C., PFEIFFER, K.J., HAO, L., STAHL, G.L. Murine model of gastrointestinal ischemia associated with complement-dependent injury. **J Appl Physiol.** 93, 338- 345, 2002.

ZINCHUK, V.V. Erythrocyte deformability: physiological aspects. **Usp Fiziol Nauk.** 32, 66- 78, 2001.