#### UNIVERSIDADE FEDERAL DE SANTA MARIA CENTRO DE CIÊNCIAS DA SAÚDE PROGRAMA DE PÓS-GRADUAÇÃO EM FARMACOLOGIA

# AÇÃO DO ÓLEO ESSENCIAL DE Hyptis mutabilis EM Rhamdia quelen PARASITADOS COM Ichthyophthirius multifiliis E SEUS EFEITOS EM PEIXES SADIOS

DISSERTAÇÃO DE MESTRADO

Jessyka Arruda da Cunha

Santa Maria, RS, Brasil 2015

# AÇÃO DO ÓLEO ESSENCIAL de Hyptis mutabilis EM Rhamdia quelen PARASITADOS COM Ichthyophthirius multifiliis E SEUS EFEITOS EM PEIXES SADIOS

#### Jessyka Arruda da Cunha

Dissertação apresentada ao Curso de Mestrado do Programa de Pós-Graduação em Farmacologia, Área de Concentração em Farmacologia Aplicada à Produção Animal, da Universidade Federal de Santa Maria (UFSM, RS), como requisito parcial para obtenção do grau de **Mestre em Farmacologia** 

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E-mail: jessyka.arruda.c@hotmail.com

#### Universidade Federal de Santa Maria Centro de Ciências da Saúde Programa de Pós-Graduação em Farmacologia

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#### AÇÃO DO ÓLEO ESSENCIAL de Hyptis mutabilis EM Rhamdia quelen PARASITADOS COM Ichthyophthirius multifiliis E SEUS EFEITOS EM PEIXES SADIOS

elaborada por Jessyka Arruda da Cunha

Como requisito parcial para obtenção do grau de Mestre em Farmacologia

Comissão Examinadora:

Berta Maria Heinzmann, Dra. (UFSM)

(Orientadora)

Luciano de Oliveira Garcia, Dr. (FURG)

Mauro Alves da Cunha, Dr. (UFSM)

Santa Maria, 25 de Fevereiro de 2015.

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#### **RESUMO**

Dissertação de Mestrado Programa de Pós-Graduação em Farmacologia Universidade Federal de Santa Maria

#### AÇÃO DO ÓLEO ESSENCIAL de Hyptis mutabilis EM Rhamdia quelen PARASITADOS COM Ichthyophthirius multifiliis E SEUS EFEITOS EM PEIXES SADIOS

AUTOR: JESSYKA ARRUDA DA CUNHA ORIENTADORA: BERTA MARIA HEINZMANN Data e Local da Defesa: Santa Maria, 25 de Fevereiro de 2015.

O objetivo deste estudo foi avaliar a atividade do óleo essencial (OE) de folhas de Hyptis mutabilis, bem como o seu principal componente (-)-globulol contra o parasito de peixe Ichthyophthirius multifiliis (Ich) e os seus efeitos sobre parâmetros hematológicos, bioquímicos e imunológicos em jundiá (Rhamdia quelen). No primeiro experimento, peixes infectados naturalmente foram tratados com óleo essencial (0, 10 e 20 mg.L<sup>-1</sup> e 199,26 μL.L<sup>-1</sup> etanol) por quatro dias através de diferentes métodos de exposição (uma única aplicação no início do experimento, duas vezes com um intervalo de 48 h e banhos diários de 1h). A sobrevivência dos peixes e o número de parasitos (trofontes por peixe) foram avaliados após 48 e 96 h. Os banhos diários de 1h apresentaram a melhor sobrevivência, quando comparados com uma única aplicação e duas aplicações com intervalo de 48 h, e essa metodologia foi definida para o segundo experimento, em que animais infectados foram expostos a (-)globulol em 2,5 e 5 mg.L<sup>-1</sup>. As concentrações mais eficazes nos experimentos 1 e 2 foram escolhidas para experimento 3, no qual animais saudáveis foram submetidos a banhos diários de 1h com OE (20 mg.L<sup>-1</sup>) ou (-)-globulol (2,5 mg.L<sup>-1</sup>). Quatro dias mais tarde, os animais tiveram parâmetros hematológicos, bioquímicos, e imunológicos avaliados. Após a exposição de animais saudáveis ao OE, foi detectado um aumento significativo no hematócrito (28,42%) e de número total de eritrócitos (2,11 x 10<sup>6</sup> mL<sup>-1</sup>) e leucócitos (48,82 x 10<sup>3</sup> mL<sup>-1</sup>), enquanto a contagem total de trombócitos (20,50 x 10<sup>3</sup> µL<sup>-1</sup>) diminuiu em relação a estudos anteriores com jundiá. No entanto, a exposição ao (-)-globulol aumentou apenas o número de leucócitos (70,87 x 10<sup>3</sup> mL<sup>-1</sup>), relativo aos valores padrão já reportados a esta espécie. O colesterol total mostrou níveis elevados (226,66 mg.dL<sup>-1</sup>) em peixes expostos ao OE, enquanto que o grupo tratado com (-)-globulol apresentou maior valor para LDL (244,66 mg dL<sup>-1</sup>) e inferior para HDL (87,66 mg.dL<sup>-1</sup>) do que os grupos de controle e etanol. OE e (-)-globulol aumentaram a sobrevida dos peixes infectados com ich e alteraram alguns parâmetros hematológicos e bioquímicos, tais como albumina, colesterol total, HDL, LDL, hematócrito, eritrócitos, leucócitos, trombócitos e MCV. Nenhuma diferença significativa foi detectada em ensaio imune inespecífica entre os grupos tratados e controle, indicando que o OE e (-)-globulol não aumentaram a imunidade inata do peixe.

**Palavras-chave:** Aquicultura Orgânica. Controle Biológico. (-)-globulol. Doença Parasitária. Peixe.

#### **ABSTRACT**

Master Dissertation
Post-Graduate Course in Pharmacology
Universidade Federal de Santa Maria

## ACTION OF Hyptis mutabilis ESSENTIAL OIL IN Rhamdia quele) PARASITIZED WITH Ichthyophthirius multifiliis AND ITS EFFECT IN HEALTHY FISH

AUTHOR: JESSYKA ARRUDA DA CUNHA ADVISOR: BERTA MARIA HEINZMANN

Date and place of the defense: Santa Maria, February 25th, 2014.

The aim of this study was to evaluate the activity of leaves essential oil (EO) of Hyptis mutabilis, as well as its major constituent (-)-globulol against the fish parasite Ichthyophthirius multifiliis (Ich) and their effects on hematological, biochemical and immunological parameters in silver catfish (*Rhamdia quelen*). In the first experiment lasting four days, naturally infected fish were treated with EO (0, 10 and 20 mg.L<sup>-1</sup> and 199.26 µL.L<sup>-1</sup> <sup>1</sup> ethanol) by means of different exposure methods (a single application at the beginning of the experiment, twice with a 48 h interval, and 1 h daily baths). Fish mortality and the number of parasites (trophonts per fish) were assessed after 48 and 96 h. The 1 h daily baths provided the best survival when compared to a single application and two applications with a 48 h interval, and this methodology was set for the second experiment, in which infected animals were exposed to (-)-globulol at 2.5 and 5 mg.L<sup>-1</sup>. The most effective concentrations in the experiments 1 and 2 were chosen for experiment 3, in which healthy animals were subjected to 1 h daily baths with EO (20 mg.L<sup>-1</sup>) or (-)-globulol (2.5 mg.L<sup>-1</sup>). Four days later, the animals had hematological, biochemical, and immunological parameters assessed. After exposure of healthy animals to EO, significant increase was detected in hematocrit (28.42 %), erythrocytes (2.11 10<sup>6</sup> µL<sup>-1</sup>) and leukocytes (48.82 10<sup>3</sup> µl<sup>-1</sup>), while the total count of thrombocytes (20.50 10<sup>3</sup> µL<sup>-1</sup>) decreased in comparison to previous studies with *Rhamdia* quelen. However exposure to (-)-globulol increased only the leukocyte number (70.87 10<sup>3</sup> µl<sup>-</sup> 1) related to the default values already reported to this species. Total cholesterol showed elevated levels (226.66 mg dL<sup>-1</sup>) in fish exposed to EO, while the group treated with (-)globulol presented higher LDL (244.66 mg dL<sup>-1</sup>) and lower HDL (87.66 mg dL<sup>-1</sup>) values than the control and ethanol groups. EO and (-)-globulol increased survival of fish infected with ich and altered some hematological and biochemical parameters, such as such as albumin, total cholesterol, HDL, LDL, hematocrit, erythrocytes, leucocytes, thrombocytes and MCV. No significant differences were detected in nonspecific immune assay between the treated groups and control, indicating that the EO and (-)-globulol did not increase innate immunity of fish.

Key-words: Organic Aquaculture. Biological Control. (-)-globulol. Parasitic Disease. Fish.

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#### INTRODUÇÃO

Na piscicultura, atualmente o Brasil é o país que apresenta o maior potencial de produção de pescado devido ao clima propício, bem como em decorrência da rede hidrográfica que contém aproximadamente 12% da água doce disponível no planeta (CONROY, 1975; GUEDES, 1980). Dentro da ordem Siluriforme e da família Heptapteridae, encontra-se a espécie *Rhamdia quelen*, popularmente conhecida como jundiá (SWARÇA *et al.*, 2000). Esta espécie animal possui distribuição neotropical e tem despertado interesse crescente devido as suas características de cultivo (BALDISSEROTTO, 2009). Além disso, possui hábitos noturnos podendo ser facilmente manipulada em âmbito laboratorial (SILFVERGRIP, 1996; FRACALOSSI *et al.*, 2002).

Entretanto, os jundiás possuem susceptibilidade ao parasito *Ichthyophthirius multifiliis* (Ich) o qual ocasiona moratalidade de até 100% dos animais em um curto período de tempo (GARCIA, *et al.*, 2007,). Embora para o Brasil não tenham sido encontradas estatísticas informando sobre o montante do prejuízo provocado por este protozoário ciliado, o mesmo causa perdas consideráveis na piscicultura mundial, sendo responsável pelo decréscimo de até 140 mil dólares por ano na economia Europa (DICKERSON, 2012). Além disso, o Ich é facilmente estimulado à proliferação na temperatura ambiente, uma vez que a 20°C o ciclo de vida deste parasito é completado em sete dias (FAO, 2011; DICKERSON, 2012; MATTHEWS, 2005).

A utilização de óleos essenciais que são fonte de compostos farmacologicamente ativos, vem sendo uma alternativa promissora frente aos demais tratamentos já descritos por possuírem vantagens em relação à toxicidade e rentabilidade (CARNEIRO *et al.*, 2005; PICÓN-CAMACHO *et al.*, 2012). A espécie vegetal *Hyptis mutabilis* (Rich.) Briq. possui na sua constituição uma ampla gama de metabolitos secundários, com destaque para os terpenoides, incluindo monoterpenoides e sesquiterpenoides que compõe o óleo essencial presente tanto em folhas quanto em inflorescências (XIMENES *et al.*, 2013).Portanto, este estudo teve como objetivo avaliar a atividade antiparasitária *in vivo* do óleo essencial (OE) de folhas de *Hyptis mutabilis* e de seu constituinte majoritário (-)-globulol na tentativa de controle do parasito *Ichthyophthirius multifiliis* em juvenis de jundiá, através da aplicação de três metodologias distintas. Além disso, foram avaliados parâmetros hematológicos,

bioquímicos e imunológicos de animais saudáveis, submetidos à metodologia que promoveu a maior sobrevivência dos peixes parasitados nos desenhos experimentais  $1\ e\ 2$ .

#### 1 OBJETIVOS

#### 1.1 Objetivo geral

Avaliar o potencial do óleo essencial (OE) de *Hyptis mutabilis* para o controle de ictiofitiríase na aquicultura.

#### 1.2 Objetivos específicos

Avaliar a atividade antiparasitária *in vivo* do OE de folhas de *Hyptis mutabilis* frente ao *Ichthyophthirius multifiliis* em jundiás;

Avaliar a sobrevivência dos animais expostos ao OE ou ao seu constituinte majoritário, (-)-globulol, em jundiás parasitados com *Ichthyophthirius multifiliis*;

Verificar o potencial de inibição da atividade hemolítica de *A. hydrophila* do OE de *Hyptis mutabilis* e do (-)-globulol utilizando eritrócitos de peixes sadios;

Verificar os efeitos da adição do OE de *Hyptis mutabilis* e do (-)-globulol na água sobre parâmetros bioquímicos e hematológicos em jundiás sadios.

#### 2 REVISÃO BIBLIOGRÁFICA

#### 2.1 Piscicultura no Brasil

O Brasil abriga cerca de 25.000 espécies de peixes de água doce, e em consequência dessa variabilidade de espécies, há uma grande variedade de formas, habitats e características distintas dos animais (CONROY, VASQUEZ, 1975; GUEDES, 1980). O país produz cerca de 2 milhões de toneladas de pescado, sendo 40% cultivados. Dentre as espécies cultivadas destaca-se a tilápia, que apresentou um aumento de 105% na produção em apenas sete anos (2003-2009) (BRASIL, 2014).

Cada região brasileira produz e cultiva determinadas espécies de peixes, sendo comum a criação de tambaqui e pirarucu na região norte, enquanto que as regiões nordeste e sudeste cultivam principalmente tilápia e camarão-marinho. Já a região sul apresenta cultivos mais diversificados, com carpas, tilápias, jundiás, ostras e mexilhões. Por outro lado, cultivos de tambaqui, pacu e pintado são frequenes na região centro-oeste (BRASIL, 2014).

Segundo os dados disponibilizados pela Agência Nacional de Águas (ANA) em 2010, o Rio Grande do Sul liderava a produção brasileira de pescado, com cerca de 55 mil toneladas. Entretanto, segundo a Emater, em 2013, o volume de pescado somou 17 mil toneladas. Desse total, 9,4 mil foram vendidas, o que gerou um faturamento de R\$ 43,9 milhões ao estado, sendo a espécie *Rhamdia quelen* responsável por R\$ 636,3 mil desse faturamento (CIGANA, 2014). Dentre os municípios do estado, Vacaria vem sendo um dos maiores produtores de jundiá no RS em sistemas extensivos e semi-intensivos (MANSKE, 2014).

Em contrapartida, a perda exponencial no ramo da aquicultura em decorrência de doenças infecciosas é um aspecto relevante e preocupante. Este fato pode ser atribuído à falta de saneamento básico nas unidades de produção, como a falta de renovação constante ou periódica de água, manejo inadequado, contaminação da água e do solo com herbicidas, além da falta de medidas de prevenção/tratamento frente a parasitoses (KREUTZ *et al.*, 2010). De maneira geral, esses fatores alteram direta e indiretamente a resposta imunológica dos peixes, impedindo a supressão à agressão de micro-organismos patogênicos, com destaque para o protozoário ciliado *Ichthyophthirius multifiliis* e a bactéria Gram-negativa *Aeromonas* 

*hydrophila*, contribuindo para um aumento da mortalidade e redução na produtividade (CONROY, VASQUEZ, 1975; KREUTZ *et al.*, 2010).

#### 2.2 Hyptis mutabilis

A utilização da fitoterapia, que compreende o tratamento de patologias e demais distúrbios através de produtos naturais obtidos de plantas, tem sua referência mais antiga datada há sessenta mil anos (REZENDE, COCCO, 2002). Já na Declaração de Alma-Ata, de 1978, a Organização Mundial da Saúde (OMS) reconheceu que 80% da população dos países em desenvolvimento valia-se de práticas tradicionais nos seus cuidados básicos de saúde e 85% usava extratos vegetais ou preparações a partir dos mesmos (OMS, 1978). Cerca de 50% dos medicamentos aprovados entre 1981 e 2006, pela Food and Drug Administration (FDA), são direta ou indiretamente derivados de produtos naturais, comprovando assim que as plantas medicinais beneficiaram, e continuam beneficiando a humanidade (FERREIRA, PINTO, 2010).

Dentre as espécies medicinais nativas do Brasil destaca-se o gênero *Hyptis* (Lamiaceae), que é formado por aproximadamente 400 espécies distribuídas desde o sul dos Estados Unidos até a Argentina. Esse gênero é composto por ervas, sub-arbustos, arbustos ou árvores pequenas, com caules geralmente quadrangulares e folhas contendo substâncias aromáticas (BORDIGNON, 1990).

No sul do Brasil, pode ser encontrada uma das principais representantes do gênero *Hyptis*, denominada *Hyptis mutabilis* (A. Rich. Briq.). Essa espécie é uma erva anual das Américas, sendo conhecida popularmente como "alfavação". Na medicina popular é utilizada para o tratamento de gastrite, dor de cabeça, como expectorante e sedativo, bem como devido aos seus efeitos antimicrobiano, anti-úlcera gástrica, larvicida, antidepressivo, anti-inflamatório e antinociceptivo. Do ponto de vista fitoquímico, essa espécie vegetal possui na sua composição química terpenoides, flavonoides, lactonas, lignanas, alcaloides e óleos essenciais (OE).

Os OEs extraídos desta espécie vegetal possuem como constituintes majoritários terpenoides, que são formados pela condensação de unidades de isopreno (C5). Esses constituintes constituem um dos maiores grupos dentre os metabólitos secundários vegetais,

sendo utilizados tanto na medicina popular quanto na terapêutica por possuírem propriedades sedativas, tranquilizantes e anticonvulsivantes (PASSOS *et al*, 2009). Dentre estes, os derivados monoterpenoides têm demonstrado atividades sobre o SNC, incluindo sedativa, antinociceptiva e antidepressiva (PERGENTINO DE SOUZA *et al.*, 2007). Já os sesquiterpenoides apresentam funções antimicrobianas descritas, além de serem comumente utilizados na indústria farmacêutica como componentes de fragrâncias e cosméticos, produtos de limpeza e aditivos de alimentos (SEO, *et al.*, 2008).

O constituinte majoritário dos OE dessa espécie varia de acordo com vários fatores, entre eles o local de coleta e as características genéticas da planta, caracterizando a ocorrência de quimiotipos (FACHINETTO, TEDESCO, 2009; XIMENES *et al.*, 2013), bem como de acordo com o órgão vegetal usado para sua extração. A espécie *H. mutabilis* coletada em Santa Maria, RS (Figura 1), forneceu como constituinte majoritário o (-)-globulol, quando o OE foi proveniente de folhas e germacreno D quando o OE foi obtido de inflorescências (SILVA *et al.*, 2013).



Figura 1 – Foto da espécie vegetal Hyptis mutabilis

Fonte: Acervo da Dra. Lenise Lima

#### 2.3 Ichthyophthirius multifiliis

Uma das maiores preocupações na aquicultura é a infecção de espécies de peixes de água doce com o ectoparasito ciliado *Ichthyophthirius multifiliis* Fouquet, 1876. Esse

protozoário é responsável pela patologia ictiofitiríase, conhecida popularmente como "doença dos pontos brancos" devido aos sinais clínicos evidenciados na superfície corpórea dos animais parasitados (BUCHMANN *et al.*, 2001; DICKERSON, 2012; MATTHEWS, 2005; BUCHMANN *et al.*, 1999; PICÓN-CAMACHO *et al.*, 2012; MARTINS *et al.*, 2002).

Há relatos na literatura que mencionam que os peixes em algum momento de sua vida irão ter contato com esse parasito, pelo fato do mesmo estar muito difundido nos ambientes de criação ou aquários (MATTHEWS, 2005; BUCHMANN *et al.*, 1999). No entanto, o contato não é o principal problema, uma vez que peixes sadios com sistema imunológico imunosuprimido são capazes de combater este parasito e impedir que a infecção ocorra (BUCHMANN *et al.*, 2001). Porém, quando os animais passam por situações que possam debilitar seu sistema imunológico como estresse, transporte, alimentação inadequada ou trocas bruscas de temperatura, eles se tornam suscetíveis a infecções oportunistas como esta parasitose (DICKERSON, 2012; MATTHEWS, 2005; POST, 1987; STOSKOPF, 1993; FAIRFIELD, 2000; FAO, 2010; FAO, 2011; CEPAL/ FAO/ IICA, 2011; INFOPESCA, 2012).

O "ich" possui um ciclo de vida bastante complexo, incluindo três estágios que compreendem as fases de teronte, trofonte e tomonte, sendo que em todas elas o parasito apresenta-se sob a forma ciliada. A primeira fase do parasito é denominada teronte, sendo o estágio infectante. Após o protozoário anexar-se à epiderme ou às brânquias do animal, ele se transforma em trofozoíto, sendo que nesta etapa o mesmo aloca-se na superfície do animal, até o momento sadio, e começa a alimentar-se das células do hospedeiro, desencadeando uma irritação cutânea. O parasito perfura e penetra a pele do hospedeiro e se aloja entre a derme e a epiderme. O peixe, por sua vez, em resposta a perfuração/penetração do parasito, desencadeia a restauração da camada perfurada e assim o parasito encistado fica alojado entre estas camadas e inacessível a substâncias externas. Após o amadurecimento do protozoário na pele do peixe, o parasito passa a ser conhecido como trofonte (trofozoíto maduro). Nesta fase o parasito deixa o corpo do animal e fica alocado geralmente no fundo dos aquários e criadouros ou até mesmo aderido à superfície de plantas para completar seu ciclo de vida na água, nesta etapa ocorreo processo de divisão e formação de células-filhas, denominadas tomontes, os quais são móveis. Neste estágio o parasito sobrevive por aproximadamente 48h, quando diferencia-se em teronte e o ciclo se repete (Figura 2) (DICKERSON, 2006,2012; MATTHEWS, 2005; LYNN, 2008; GHIRALDELLI et al., 2007; POST, 1987; STOSKOPF,

1993; FAIRFIELD, 2000; FAO, 2010; FAO, 2011; CEPAL/ FAO/ IICA, 2011; INFOPESCA, 2012).

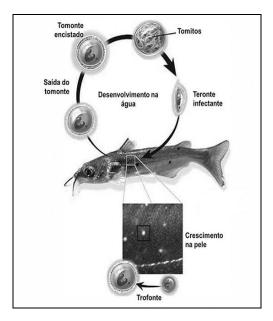


Figura 2 – Representação ilustrativa do ciclo do parasito *Ichthyophthirius multifiliis*Fonte: Panorama da aquicultura. <a href="http://www.panoramadaaquicultura.com.br/novosite/?p=2120">http://www.panoramadaaquicultura.com.br/novosite/?p=2120</a>

Os animais parasitados apresentam pontos brancos e comportamento característico, como desequilíbrio no nado e lesões provocadas pelos próprios animais quando os mesmos debatem-se em pedras ou fundos de aquários, na tentativa de esfregar-se em decorrência da irritação, podendo também desencadear hipóxia, agitação intensa, perda do apetite e eventual morte, por tornarem-se suscetíveis a infecções secundárias, tais como as bacterianas (POST, 1987; STOSKOPF, 1993; FAIRFIELD, 2000; FAO, 2010; FAO, 2011; CEPAL/ FAO/ IICA, 2011; INFOPESCA, 2012).

A solução para controlar o parasito é relativamente fácil quando o tratamento é direcionado para tanques e aquários, sendo realizado conforme as alternativas apresentadas pela literatura, como controle da temperatura e higienização dos taques (FAIRFIELD, 2000). Porém, na maioria das vezes este recurso não se mostra eficaz nas pisciculturas, apenas em âmbito laboratorial ou em aquários. Em vista, das perdas na aquicultura ocasionada pelo Ich, tornou-se necessária a busca pelo tratamento focado no animal e não só nos reservatórios (POST, 1987; STOSKOPF., 1993; FAO, 2010; FAO, 2011; CEPAL/ FAO/ IICA, 2011; INFOPESCA, 2012).

Estudos relatam algumas intervenções frente ao parasito, como a utilização de 4g.L<sup>-1</sup> de sal, o que segundo os autores, melhora a sobrevivência de jundiás parasitados, podendo

ser utilizada então como um tratamento eficaz (GARCIA *et al.*, 2007). Contudo, a maioria dos tratamentos disponíveis é a base de produtos químicos de reconhecida toxicidade, administrados na forma de banhos, como formaldeído, o sulfato de cobre, permanganato de potássio, ácidos orgânicos, ferrato de potássio (VI), bronopol e produtos à base de ácido peracético. Entretanto, estes tratamentos requerem altas concentrações de produtos por um longo período, desencadeando elevados custos e impactos ambientais, bem como o risco de aumentar a resistência do parasito a estes recursos (PICÓN-CAMACHO *et al.*, 2012). Portanto, os tratamentos mencionados são limitados pela toxicidade, rentabilidade e difícil controle em pisciculturas de grande porte (SUTILI *et al.*, 2014). Em vista deste problema, um estudo abordou a modificação do pH da água e dureza, sendo então constado que essa infecção é menos grave em *R. quelen* quando a água for mantida em pH 5 e dureza igual a 20 mg L<sup>-1</sup> de CaCO<sub>3</sub> (GARCIA *et al.*, 2011).

#### 2.4 Rhamdia quelen

A espécie *Rhamdia quelen* (Figura 3) pertence à família Heptapteridae e é popularmente conhecida jundiá, jundiá-tinga, jandiá, jandiá-tinga, mandi, sapipoca, bagre, bagre negro, bagre-sapo, bagre sulamericano, nhurundia e mandi-Guaru. O jundiá tem coloração que varia de marrom avermelhado claro a cinza ardósia e possui característica ectodérmica e tem ampla distribuição geográfica, sendo encontrado desde o sudeste do México até o sul da Argentina. Habita preferencialmente fundos arenosos de lagos e rios de águas calmas, possuindo hábitos noturnos, alimentação omnívora, com tendência piscívora e podendo atingir cerca de 50 cm de comprimento e 3Kg de peso corporal (GOMES *et al.*, 2000).

O jundiá é a espécie nativa mais cultivada no RS, uma vez que é tolerante às baixas temperaturas do inverno. Ainda, apresenta facilidade de adaptação em âmbito laboratorial e vem sendo utilizada há anos pelo nosso grupo de pesquisas. Os diversos trabalhos têm como objetivo testar modelos experimentais que envolvem pesquisa para melhoramento de dieta, avaliação de fatores de crescimento, reprodução, prevenção/tratamento de patologias que acometem peixes de água doce, desenvolvimento de sedativos e anestésicos para uso em

piscicultura, entre outros (SUTILI et al., 2014; GOMES et al., 2000; GARCIA et al., 2011 SILVA et al., 2013; LAZZARI et al., 2011; BARCELLOS et al., 2014).



Figura 3 – Foto de um exemplar de *Rhamdia quelen* 

Fonte: Internet

Os peixes teleósteos são os primeiros da classe de vertebrados com imunidade inata e adaptativa (WHYTE, 2007). Estas respostas são moduladas por fatores intrínsecos e extrínsecos, como a idade, temperatura, contato com herbicidas e alimentação, entre outros fatores. Nesses animais, os macrófagos teciduais são as principais células do sistema imune inato e representam uma das primeiras e mais importantes barreiras frente à micro-organismos patogênicos. Também têm a capacidade de fornecer sinalização para os linfócitos, que atuam na imunidade adquirida (GUEDES, 1980; SILFVERGRIP, 1996; FRACALOSSI *et al.*, 2002; KREUTZ *et al.*, 2010; AOKI *et al.*, 2013). Entretanto, o jundiá possui susceptibilidade frente ao protozoário conhecido como "ich" (*Ichthyophthirius multifiliis*), sendo que o cultivo desta espécie em sistemas de aquicultura, como tanques e aquários, por vezes pode desencadear uma contaminação em massa por agentes infectantes ou até mesmo uma co-infecção por parasitos ou bactérias, ocasionando uma perda considerável na produção (SUTILI *et al.*, 2013).

A espécie *Rhamdia quelen* (jundiá) não possui seus parâmetros hematológicos e bioquímicos bem elucidados, pois de acordo com a literatura esses parâmetros são bastante variáveis e ainda não possuem valores normais e intervalos de confiança bem estabelecidos. Ainda embasando-se na literatura, tem-se que através do sangue podem ser evidenciadas várias alterações metabólicas. Também pode ser avaliado o estado de saúde dos animais e eventuais enfermidades nos mesmos, além de contribuir para a compreensão da fisiologia,

relação filogenética interespécies e outros parâmetros ecológicos provenientes da intervenção de produtos comerciais/ naturais (TAVARES-DIAS *et al.*, 2000; LAZZARI *et al.*, 2011; BARCELLOS *et al.*, 2014).

# 3 MANUSCRITO

The essential oil *Hyptis mutabilis* against *Ichthyophthirius multifiliis* and its effect on hematological, biochemical and immunological parameters in silver catfish (*Rhamdia quelen*)

Cunha, J.A.; Sutili, F.J.; Motta, A.O.; Gressler, L.T.; Scheeren, C.A; Silva, L.L.; Almeida, R.V.; Baldisserotto, B.; Heinzmann, B.M

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The essential oil *Hyptis mutabilis* against *Ichthyophthirius multifiliis* and its effect on hematological, biochemical and immunological parameters in silver catfish (*Rhamdia quelen*)

Cunha, J.A<sup>1</sup>.; Sutili, F.J<sup>1</sup>.; Motta, A.O<sup>2</sup>.; Gressler, L.T<sup>3</sup>.; Scheeren, C.A<sup>2</sup>; Silva, L.L<sup>4</sup>.; Almeida, R.V<sup>5</sup>.; Baldisserotto, B<sup>1,6</sup>.; Heinzmann, B.M<sup>1,2,7</sup>.

<sup>1</sup>Programa de Pós-Graduação / PPG em Farmacologia, Federal University of Santa Maria / UFSM, Campus Universitário, Prédio 21, BR-97105-900, Santa Maria, RS, Brazil.

Phone: (55) 32209674; email: <a href="mailto:berta.heinzmann@gmail.com">berta.heinzmann@gmail.com</a>; Address: UFSM/ CCS /DFI, Prédio 26, Campus Universitário, BR-97105-900, Santa Maria, RS, Brazil.

#### **ABSTRACT**

The aim of this study was to evaluate the activity of leaves essential oil (EO) of *Hyptis mutabilis*, as well as its major constituent (-)-globulol against the fish parasite *Ichthyophthirius multifiliis* (Ich) and their effects on hematological, biochemical and immunological parameters in silver catfish (*Rhamdia quelen*). In the first experiment lasting four days, naturally infected fish were treated with EO (0, 10 and 20 mg.L<sup>-1</sup> and 200 μL.L<sup>-1</sup> ethanol) by means of different exposure methods (a single application at the beginning of the experiment, twice with a 48 h interval, and 1 h daily baths). Fish mortality and the number of parasites (trophonts per fish) were assessed after 48 and 96 h. The 1 h daily baths provided the best survival when compared to a single application and two applications with a 48 h interval, and this methodology was set for the second experiment, in which infected animals were exposed to (-)-globulol at 2.5 and 5 mg.L<sup>-1</sup>. The most effective concentrations in the experiments 1 and 2 were chosen for experiment 3, in which healthy animals were subjected to 1 h daily baths with EO (20 mg.L<sup>-1</sup>) or (-)-globulol (2.5 mg.L<sup>-1</sup>). Four days later, the

<sup>&</sup>lt;sup>2</sup> Curso de Graduação em Farmácia, UFSM, Santa Maria, RS, Brazil.

<sup>&</sup>lt;sup>3</sup> PPG em Medicina Veterinária, UFSM, Santa Maria, RS, Brazil.

<sup>&</sup>lt;sup>4</sup> Universidade Regional Integrada do Alto Uruguai e das Missões/ URI, Santiago, RS, Brazil.

<sup>&</sup>lt;sup>5</sup> PPG em Análises Clínicas, Centro Universitário Franciscano, Santa Maria, RS, Brazil.

<sup>&</sup>lt;sup>6</sup> Departamento de Fisiologia e Farmacologia, UFSM, Santa Maria, RS, Brazil.

<sup>&</sup>lt;sup>7</sup> Departamento de Farmácia Industrial, UFSM, Santa Maria, RS, Brazil.

animals had hematological, biochemical, and immunological parameters assessed. After exposure of healthy animals to EO, significant increase was detected in hematocrit (28.42 %), erythrocytes (2.11 x  $10^6 \ \mu L^{-1}$ ) and leukocytes (48.82 x  $10^3 \ \mu L^{-1}$ ), while the total count of thrombocytes (20.50 x  $10^3 \ \mu L^{-1}$ ) decreased in comparison to previous studies with *Rhamdia quelen*. However exposure to (-)-globulol increased only the leukocyte number (70.87 x  $10^3 \ \mu l^{-1}$ ) related to the default values already reported to this species. Total cholesterol showed elevated levels (226.66 mg.dL<sup>-1</sup>) in fish exposed to EO, while the group treated with (-)-globulol presented higher LDL (244.66 mg.dL<sup>-1</sup>) and lower HDL (87.66 mg.dL<sup>-1</sup>) values than the control and ethanol groups. EO and (-)-globulol increased survival of fish infected with ich and altered some hematological and biochemical parameters, such as such as albumin, total cholesterol, HDL, LDL, hematocrit, erythrocytes, leucocytes, thrombocytes and MCV. No significant differences were detected in nonspecific immune assay between the treated groups and control, indicating that the EO and (-)-globulol did not increase innate immunity of fish.

Key-words: organic aquaculture, biological control, (-)-globulol, parasitic disease

#### 1. INTRODUCTION

The silver catfish (*Rhamdia quelen*) is the most raised native species from southern Brazil (Baldisserotto *et al.*, 2009) because presents good food conversion, growth rate and absence of intramuscular spines (Gomes *et al.*, 2000). However, this species is very susceptible to the protozoan parasite *Ichthyophthirius multifiliis* (ich) (Miron *et al.*, 2003; Garcia *et al.*, 2007). Farming of silver catfish in aquaculture systems such as ponds and aquaria sometimes may trigger a massive contamination by infectious agents or even a co-infection with bacteria (*A. hydrophila*) or parasites (*I. multifiliis*) (Sutili *et al.*, 2013). The infected animals present white spots and characteristic behavior, which includes swimming imbalance and injuries caused when the fish scratch the tank walls as a result of irritation, and can also lead to respiratory distress, severe agitation, loss of appetite and death as a result of susceptibility to secondary infections (Matthews, 2005; Garcia *et al.*, 2007; Sutili *et al.*, 2013).

Some treatments were used against ich, such as the addition of salt (Miron *et al.*, 2003), copper sulfate (Straus, 2008; Straus *et al.*, 2009), potassium permanganate (Straus & Griffin, 2002) and increasing temperature. However, these treatments are limited by toxicity

and/or low profitability and control in large fish farms (Carneiro *et al.*, 2005; Picón-Camacho *et al.*, 2012). In this context, an alternative increasingly considered are essential oils (EO), complex mixtures of volatile compounds from plants that are a source of pharmacologically active constituents and may trigger analgesic, anti-inflammatory, antibiotic and repellant effects, among others (Curtis *et al.*, 1989; Gillij *et al.*, 2007; Nerio *et al.*, 2010).

Plants from the Lamiaceae family are known to be sources of EO, among which stands out *Hyptis mutabilis* (Rich.) Briq., used in folk medicine due to its antimicrobial, larvicide, sedative, anti-inflammatory and antinociceptive effects (Fachinetto,Tedesco, 2009; Ximenes *et al.*, 2013; Silva *et al.*, 2013). The EO of this species collected in southern Brazil has in its composition mono- and sesquiterpenoids and as the major constituent (-)-globulol (26.61%) when the EO is derived from leaves, and Germacrene D (14.97%) when from inflorescences (Silva *et al.*, 2013). This study aimed to evaluate the *in vivo* antiparasitic activity of the EO of *Hyptis mutabilis* leaves and of (-)-globulol against ich using different methodologies. Hematological, biochemical and immunological parameters of healthy animals were also evaluated, after treatment by the methodology that promoted the highest survival of parasitized animals during the tests.

#### 2. MATERIALS AND METHODS

#### 2.1. Plant material and phytochemical analysis

Leaves of *Hyptis mutabilis* were collected in March 2012 (at the end of summer) in Santa Maria (Rio Grande do Sul state, southern Brazil). A voucher specimen was deposited at the Herbarium of the Departamento de Biologia, Universidade Federal de Santa Maria (N: SMDB 13076). The EO was extracted and analyzed as described by Silva *et al.* (2013), which also reported the isolation of (-)-globulol from the EO by column chromatography, identification of this constituent by EI-MS, <sup>1</sup>H- and <sup>13</sup>C-NMR. The measure of optical rotation by these authors defined the constituent as the levorotatory isomer.

#### 2.2. Fish and water quality

Silver catfish (R. quelen) juveniles (Exp 1:  $5.3 \pm 0.40$  g, Exp 2:  $25 \pm 0.50$  g and Exp 3:  $5.6 \pm 0.30$  g) were used to evaluate survival and immunological, biochemical and hematological parameters after exposure. All fish were transferred from a local fish culture to the laboratory, where they were maintained in continuously aerated 250 L tanks, with controlled water parameters until experiments were performed. The acclimation tanks had

70% of water exchanged each 48 h. Dissolved oxygen and temperature were measured with an YSI oxygen meter (Model Y5512) and the pH by a DMPH-2 pH meter (Digimed). Total ammonia levels were determined according to Verdouw *et al.* (1978) and un-ionized ammonia (NH3) levels were calculated according to Ostrensky (1997). Fish were acclimated for seven days and were fed once a day with commercial feed (28.0% crude protein). Juveniles were fasted for a period of 24 h prior to the experiments.

The mean water quality parameters in experiment 1 were: temperature:  $21.0 \pm 2$  °C, pH:  $6.8 \pm 0.11$ , dissolved oxygen levels:  $7.98 \pm 0.12$  mg.L<sup>-1</sup>, total ammonia levels:  $2.08 \pm 0.21$  mg.L<sup>-1</sup>, non-ionized ammonia:  $0.0279 \pm 0.0008$  mg.L<sup>-1</sup>; experiment 2 were: temperature:  $21.0 \pm 2$  °C, pH:  $7.1 \pm 0.08$ , dissolved oxygen levels:  $5.8 \pm 1.65$  mg.L<sup>-1</sup>, total ammonia levels:  $1.20 \pm 0.15$  mg.L<sup>-1</sup>, non-ionized ammonia:  $0.0279 \pm 0.0006$  mg.L<sup>-1</sup>; and in experiment 3 were: temperature:  $21.0 \pm 2$  °C, pH:  $7.3 \pm 0.05$ , dissolved oxygen levels:  $7.2 \pm 1.15$  mg.L<sup>-1</sup>, total ammonia levels:  $1.80 \pm 0.3$  mg.L<sup>-1</sup>, non-ionized ammonia:  $0.0242 \pm 0.0007$  mg.L<sup>-1</sup>.

#### 2.3. Experiment 1

All experiments were conducted with water (control) and ethanol (at the same concentration used for dilution of the highest concentration of EO). The EO was diluted in 95% ethanol (1:10) and added separately to 1.5 L aquaria, while (-)-globulol was added directly to the water. Fish infected with *I. multifiliis* (in quadruplicate with n = 5 fish for each replicate and about 20 white spots per fish) were exposed for four days to EO at 0, 10 and 20 mg.L<sup>-1</sup> and 200 μL.L<sup>-1</sup> ethanol using three different methodologies: a single application at the beginning of the experiment (A), twice with a 48 h interval (B) and 1 h daily baths (C). To evaluate the antiparasitic activity of EO, lower concentrations than those reported as sedative were chosen (Silva *et al.*, 2013). Parasites were counted in the dorsolateral region above the lateral line with a magnifying glass (x 10) at the beginning of the experiment, after 48 h, and at the end of the experiment (96 h). Daily mortality in each group was also assessed through the same period.

#### 2.4. Experiment 2

Silver catfish infected with ich (in quadruplicate, with n = 5 fish for each replicate and about 18 white spots per fish) were exposed for four days to (-)-globulol at 2.5 and 5 mg.L<sup>-1</sup> through 1 h daily baths. The globulol concentrations chosen for the experiment 2 were lower

than those which showed sedative effect in silver catfish (Silva *et al.*, 2013). The counting of parasites and the assessment of mortality were performed as described in experiment 1.

#### 2.5. Experiment 3

According to the results of the first and second experiments (survival experiments), the optimum exposure method and the most effective concentrations of EO and (-)-globulol were chosen in order to investigate possible effects on fish metabolism. Thus, healthy animals were subjected to 1 h daily baths for four days at 20 mg.L<sup>-1</sup> EO or 2.5 mg.L<sup>-1</sup> (-)-globulol. The methodology of all these experiments was approved by the Ethical and Animal Welfare Committee of the Universidade Federal de Santa Maria (Process N: 046/2010).

#### 2.6. Blood collection-Experiment 3

At the end of the fourth day of the experiment 3, fish from each aquarium were anesthetized with eugenol (50 mg.L<sup>-1</sup>), blood was collected with heparinized syringes and stored in eppendorf tubes for later hematological analysis. The material directed to biochemical and immunological analysis was stored in another eppendorf tubes, which were centrifuged (1500 x g, room temperature,1 min) to separate serum and plasma.

#### 2.7. Hematological analysis

Hematological parameters were evaluated at the end of experiment 3, using an automatic counter XS-800i (Symex®). Total leukocytes, total erythrocytes, hematocrit (Ht), hemoglobin concentration (Hb), mean corpuscular volume (MCV), mean corpuscular hemoglobin concentration (MCHC) and platelets were determined. Blood smears were fixed in methanol and stained with Instant-Prov (NewProv®) for determination of the differential white blood cell (WBC) count. At least 200 WBCs were counted for differential WBC determinations.

#### 2.8. Serum biochemistry

The serum levels of glucose, total proteins, albumin, total bilirubin, creatinine, urea, total cholesterol, triglycerides, HDL and LDL cholesterol were evaluated at the end of experiment 3 in an automated Vitros 250 (Ortho – Clinical Diagnostics) using Johnson & Johnson kits dry chemistry method. All tests were carried out in duplicate.

#### 2.9. Non-specific immune assays

The complement system activity was determined at the end of experiment 3, according to Castro *et al.* (2008) using fresh plasma (without freezing) from each fish. Rabbit red blood cells were added to a normal plasma and incubated at 20 °C, after 1 h ice-cold saline was added to stop the complement activity, cells were pelleted by centrifugation and the absorbance of supernatant was measured in a microplate reader (405 nm). The percent hemolysis was calculated by comparison between total hemolysis (100%) and no-hemolysis (0%) controls.

The natural agglutination activity of fish plasma was investigated using "U"-shaped 96-well plates. Plasma was diluted two-fold in PBS (pH 7.4, Ca<sup>+2</sup> and Mg), and an equal volume of washed *Aeromonas hydrophila* (ATCC 7966) in suspension (0.4 OD 600 nm) was added to each well. The plates were incubated for 2 h at 25 °C and then overnight at 4 °C; the titer was defined as the logarithm of the highest dilution of the plasma that caused complete agglutination of the bacterial cell (Sutili *et al.*, 2014).

Plasma bactericidal activity was assessed against *A. hydrophila* (ATCC 7966) by agar diffusion method, using paper discs (5 mm diameter). The bacteria (McFarland  $0.5 \times 10^8$  cells.mL<sup>-1</sup>) were seeded in Petri dishes (solid medium Müller Hinton) and plasma was soaked on the discs (20  $\mu$ L). The plates were incubated at 30 °C for 24 h, and afterwards the diameter of the halo formed around the discs was assessed (Sutili *et al.*, 2014).

#### 2.10. Statistical analysis

The homogeneity of variances between groups was determined by Levene test. Comparisons between different groups were performed using one-way ANOVA and Tukey's test (Statistica 7.0 Software). Fish survival was compared using Kaplan-Meier survival analysis with the Log-rank test (SPSS 18 Software). The minimum significance level was set at  $P \leq 0.05$ . Hematological, biochemical and immunological parameters are presented as mean values and standard error.

#### 3. RESULTS

### 3.1. Effect of EO of *H. mutabilis* and (-)-globulol against *Ichthyophthirius multifiliis* and fish survival

Experiment 1 - control and ethanol groups showed survival of 60% and 40% after 96 h of exposure, respectively. The group treated with a single exposure to 10 mg.L<sup>-1</sup> EO presented equivalent survival as the control, but fish submitted to 20 mg.L<sup>-1</sup> EO had significantly higher

survival (P  $\leq$  0.01) (100%) than the control (Fig 1A). The control group of the two applications methodology showed 20% survival at the end of 96 h of exposure. Fish treated with EO and ethanol showed higher survival compared to the control (P  $\leq$  0.01) (Fig. 1B). The survival of the control and ethanol groups were 50 and 80%, respectively, after 96 h exposure with daily baths. Both groups treated with EO presented higher survival (P  $\leq$  0.01) than the control and ethanol group (Fig. 1C).

The number of trophonts per fish was calculated considering the average values (n=20) for the intervals of 0, 48 and 96 hours, for each of the treatments. In methodology A, ethanol group showed averages of 8.2, 10 and 75 at the above mentioned times, while for the control group, the corresponding averages were 8.4, 11.2 and 53.3. At the concentration of 10 mg.L<sup>-1</sup> EO, averages of 6.3, 5.4 and 17.3 were found, and averages of 7.9, 8.2 and 39.2 were observed for the concentration of 20 mg.L<sup>-1</sup> EO. However, for methodology B, the ethanol group had means of 8.7, 5, 47.33, respectively, while for the control averages of 9.6, 1.25 and 1 were observed. The averages detected at the concentration of 10 mg.L<sup>-1</sup> EO were 8.7, 13.9 and 35.3 and at 20 mg L<sup>-1</sup> EO the corresponding values were 9.1, 5.3 and 41.66. In the methodology C, the ethanol group showed values of 15.8, 24.33 and 11.5, while for the control 9.3, 3.5 and 11 were observed. However, at the concentration of 10 mg.L<sup>-1</sup> EO, 13.8, 48.2 and 20.2 were found, while at 20 mg.L<sup>-1</sup> EO, 13.1, 24.66 and 19.8 were observed (Fig. 2A-C).

When a single application of EO was used, the control group differed from ethanol, 10 and 20 mg.L-1 EO after 48h; at the same time, 10 mg.L-1 EO was different from ethanol, control and 20 mg.L-1. However, after 96 h, control group and 10 mg.L-1 EO differed from ethanol, while 20 mg.L-1 EO differed from control and 10 mg.L-1 EO (Fig. 2A). The treatment that received two applications within 48 h, after the first 48 h 10 mg.L-1 EO differed from ethanol, control group and 20 mg.L-1 EO (Fig. 2B). The 1 h daily baths treatment showed statistic difference between 10 mg.L-1 EO and ethanol after 48 h (Fig. 2C).

Statistical differences were also observed for the same methodology at different times. In methodology A, differences were observed among the control, ethanol and 20 mg.L<sup>-1</sup> EO after 96 h when compared to the other two evaluated times (Fig. 2A). In the methodology B, no difference was detected after 96 h when compared to the other two times evaluated in ethanol group, 10 and 20 mg.L<sup>-1</sup> EO. There was also no difference in the control group between the times 0 and 48 h (Fig. 2B). In methodology C, the group subjected to a concentration of 10 mg.L<sup>-1</sup> EO showed statistic difference among different times (Fig. 2C).

Experiment 2 - After 96 h from the beginning of the experiment, the survival observed for fish submitted to 1 h daily baths of (-)-globulol 2.5 mg.L<sup>-1</sup> was significantly highest ( $P \le 0.01$ ) than the other treatments (Fig. 1D). The fishes which were exposed to (-)-globulol at 2.5 and 5 mg.L<sup>-1</sup> through 1 h daily baths (Fig. 1D) revealed a difference between 5 mg.L<sup>-1</sup> and ethanol, while water and 2.5 mg.L<sup>-1</sup> showed to be different after 48 hours. For 96 hours interval, (-)-globulol 5 mg.L<sup>-1</sup> differed from ethanol and control group; ethanol and control group were different from all other treatments, including each other.

Concerning the number of trophonts per fish, the average numbers after 0, 48 and 96 h observed for the ethanol group were 18.7, 57 and 5.3, whereas for the control group the corresponding data were 15.4, 51.2 and 71.3. For 2.5 mg.L<sup>-1</sup> (-)-globulol, averages of 16.3, 73.4 and 50.3 were obtained, while for 5 mg.L<sup>-1</sup> (-)-globulol, 20, 37.2 and 39.2 were found.

Considering the three evaluated time periods, a significant statistic difference were observed for the control group and 2.5 mg.L<sup>-1</sup> (-)-globulol at time 0 h when compared to 48 and 96 h; at 0 and 96 h ethanol group was different when compared to 48 h (Fig. 2D).

Significant difference between treatments after 48 h was detected among the group subjected to 5 mg.L<sup>-1</sup> (-)-globulol in relation to the control, ethanol and 2.5 mg.L<sup>-1</sup> (-)-globulol groups within the same time. After 96 h, there was a difference of 5 mg L<sup>-1</sup> (-)-globulol when compared to the control and ethanol groups, and at this same time the ethanol group differed from all other treatments (Fig 2D).

Experiment 3 – Daily baths of EO (20 mg.L<sup>-1</sup>) and (-)-globulol (2.5 mg.L<sup>-1</sup>) with respect to infected fish survival showed no significant difference between the two treatments, however both groups presented higher survival than the control and ethanol groups (Fig. 1E).

## **3.2.** Effect of essential oil of *H. mutabilis* and (-)-globulol on hematological and biochemical parameters and non-specific immune assays

Of the ten biochemical parameters evaluated in serum, HDL presented significantly lower values in fish subjected to baths with 2.5 mg.L<sup>-1</sup> (-)-globulol compared to control fish. Albumin showed high values for ethanol compared to the control group. The LDL parameter presented lower values for the control group and ethanol when compared with 20 mg.L<sup>-1</sup> EO. For total cholesterol, higher values were obtained for the animals treated with 20 mg.L<sup>-1</sup> EO in comparison to other treatments with ethanol and 2.5 mg.L<sup>-1</sup> (-)-globulol, and to the control group (Table 1).

Considering the analyzed hematological parameters, hematocrit showed higher levels in fish exposed to EO (20 mg.L<sup>-1</sup>) compared to control group. The treatment with (-)-globulol (2.5 mg.L<sup>-1</sup>) showed a significant increase in the total number of erythrocytes and leukocytes when compared to control group. The number of thrombocytes were lower in fish exposed to 20 mg.L<sup>-1</sup> EO compared to control, ethanol and groups, (-)-globulol (2.5 mg.L<sup>-1</sup>). However, MCV values were higher in the control group in comparison to 2.5 mg.L<sup>-1</sup> (-)-globulol and 20 mg.L<sup>-1</sup> EO (Table 2).

The complement system activity represented by hemolysis of fish erythrocytes showed no significant differences for all treated groups compared to control (Table 2). Titer and halo formation in the plasma agglutination and plasma bactericidal assays were not verified in non-specific immune assays avaliable.

#### 4. DISCUSSION

*Ichthyophthirius multifiliis* has the ability to infect all freshwater fish species and is considered an important research field due to the significant economic impact on fish farming. It is also a good model for the study of innate and acquired immunity against fish parasites in teleost (Dickerson, Clark, 1998; Dickerson, 2012; Matthews, 2005).

In the present study, a decrease of the number of white spots could not be observed. However animals treated with EO or (-)-globulol showed a higher survival compared to untreated fish. Effective alternatives to combat the great losses caused by ich are the use of 4 g. L<sup>-1</sup> of common salt (Miron *et al.*, 2003) or 1.1 mg.L<sup>-1</sup> CuSO<sub>4</sub> (Straus, 2008). Salt prevents the fish deaths and progressively reduces the number of trophonts, however its use is not feasible in large aquaculture systems. Although CuSO<sub>4</sub> has showed efficacy in a study (Straus, 2008) it also presented high toxicity (Straus, Griffin, 2002; Straus *et al.*, 2009).

Research with EO have focused on this problem, since according to the Food and Agriculture Organization the use of plant extractives as EO are allowed to control parasites in organic fish farming (Prein *et al.*, 2012). Reports of *in vitro* activity of *Melaleuca alternifolia*, *Lavandula angustifolia* and *Mentha piperita* EO purchased commercially against *I. multifiliis* have been found, and the EO of *M. alternifolia* was effective to treat ichthyophthiriasis in pacu (*Piaractus mesopotamicus*) (Valladão, 2014).

The blood may show various metabolic changes and also provides an animal health condition assessment and detection of any diseases. These parameters can also contribute to understand the physiology, interspecies phylogenetic relationship and other ecological

parameters from intervention commercial/ natural products (Tavares-Dias et al., 2000; Lazzari et al., 2011; Barcellos et al., 2014). In this study the control group values of serum glucose, albumin, total proteins, total bilirubin, total cholesterol, creatinine and urea were similar to basal values reported for the species (Borges et al., 2004). However, for triglycerides and LDL levels were lower, while HDL was higher when compared to those already reported (Borges et al., 2004). There are no literature data addressing a possible explanation for the increase in LDL and HDL observed in R. quelen. The obtained results for the hematological parameters erythrocytes, hemoglobin, hematocrit, MCHC, MCV, heterophils and eosinophils were similar to those described by Tavares-Dias, et al. (2000). Regarding to total thrombocyte number, a decrease of this parameter in animals exposed to EO and also an increase of total leucocyte number in fish treated with EO and globulol were observed. The thrombocytes represent an important linkage between innate and adaptive immunity in fish, which interaction is involved in protection against pathogenic and environmental factors (Passantino et al., 2005). The role of thrombocytes in fish defense mechanisms has been quite studied but is still controversial because there is no evidence of phagocytosis by thrombocytes (Meseguer et al., 2002). Leukocytes are responsible for defending the organism against infections and parasites, acting in the production of antibodies (Misra et al., 2006; Ranzani-Paiva et al., 2004). Although the intervention with the OE and globulol was not statistically different from the control in the immunoassay, hematology results corroborate with the increased survival.

A wide range of EO from aromatic medicinal plants consists of many active substances that are known to trigger inhibitor or stimulating effects on the immune system and may also have immunomodulatory properties in fish (Sutilli *et al.*, 2014). However, in this study there was no significant change in the immunological test to determine the hemolysis percentage of fish erythrocytes of treated fish when compared to control. Fish immune system is classified into innate or nonspecific defense, which is the first line of protection against pathogens, and specific defense, characterized by the immune memory (Biller-Takahashi *et al.*, 2012). The complement system activity is widely used as an immune indicator (Bayne, Gerwick, 2001), and it was evaluated to measure the hemolytic activity of the alternative pathway from complement system, as an indicator of the innate immunity in silver catfish after being and submitted to 1 h daily baths with EO or (-)-globulol. Despite the increased survival of infected animals, hemolysis of treated fish erythrocytes showed no significant difference from control. In addition, the significant differences detected in the

number of trophonts per fish in relation to the control group could not be assigned to an antiparasitic activity, since the reduction in the number of trophonts was directly related to mortality of the specimens under study. However, sedative and anesthetic activities detected for *H. mutabilis* EO and (-)-globulol may be contributing to the increased survival of infected animals. Since (-)-globulol showed positive interaction with diazepam regarding the depressor effects, (Silva *et al.*, 2013), its sedative effect may be preventing the decline of the immune defenses resulting from stress.

It is also noteworthy that factors such as nutritional status, seasonality, maturation, sex and genetic variation can also significantly influence the hematological variables. Furthermore, differences may occur in the blood collection methodology, regarding the type of anticoagulant used, which can also act as a source of variation in fish hematology results (Tavares-Dias *et al.*, 2000). Thus, a clear explanation for the obtained results is still missing. However, the lack of significant difference in the survival increase of infected fish treated with EO (20 mg.L<sup>-1</sup>) or with globulol (2.5 mg.L<sup>-1</sup>) allows us to suggest that the tested products can be an alternative for the control of parasites in both organic as conventional aquacultures.

#### 5. CONCLUSION

Daily baths of the EO of *H. mutabilis* (20 mg.L<sup>-1</sup>) and its major constituent (-)-globulol (2.5 mg.L<sup>-1</sup>) increased the survival rate of silver catfish infected with ich. The number of trophonts per fish changed over time and among treatments. However, the detected differences could be attributed to mortality of animals, excluding a possible antiparasitic activity of EO and its major constituent at the concentrations used in this study. Changes in some hematological and biochemical parameters were observed in healthy fish, but no clear relationship between them and higher survival of infected fish was found.

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- **Table 1.** Biochemical serum parameters of healthy silver catfish after 1 h daily baths with OE of *Hyptis mutabilis* or (-)-globulol for four days, n=20.
- **Table 2.** Hematological and immunological parameters of silver catfish after 1 h daily baths with EO of *Hyptis mutabilis* or (-)-globulol for four days, n=20.
- **Figure 1.** Fish survival exposed to *Hyptis mutabilis* EO submitted to three different methodologies for four days: a single application at the beginning of the experiment (A), twice with an interval of 48 h (B) and 1 h daily baths (C). Fish survival after exposure to (-)-globulol through 1 h daily baths in four consecutive days (D). Comparison between EO 20 mg.L<sup>-1</sup> and (-)-globulol 2.5 mg.L<sup>-1</sup> (E). \* Significant difference from control; #: Significant difference from the group treated with ethanol (Kaplan-Meier (Lag-Rank),  $P \le 0.01$ ), n=20.
- **Figure 2.** Number of white spots in the dorsolateral region of silver catfish exposed to EO of *Hyptis mutabilis* submitted to three different methodologies for four days: a single application at the beginning of the experiment (A), twice with an interval of 48 h (B) and 1 h daily baths (C). Number of white spots in fish exposed to (-)-globulol through 1 h daily baths for four days (D). Lower case letters indicate significant differences between treatments evaluated in the same time. Capital letters indicate significant differences of the same treatment in different time periods (P < 0.05), n = 20.

Table 1

Parameters	Treatment					
	Control	Ethanol	Globulol 2.5 mg.L <sup>-1</sup>	EO 20 mgL <sup>-1</sup>		
Glucose	$45.66 \pm 3.48$	$48.33 \pm 7.35$	$42.66 \pm 0.88$	48.33 ± 5.24		
$(mg.dL^{-1})$						
Albumin	$2.80 \pm 0.35^{b}$	$4.17 \pm 0.33^{a}$	$2.96 \pm 0.18^{a,b}$	$3.30 \pm 0.20^{a,b}$		
$(g.dL^{-1})$						
Total Bilirubin	$0.16 \pm 0.06$	$0.17 \pm 0.02$	$0.23 \pm 0.02$	$0.24 \pm 0.05$		
$(mg.dL^{-1})$						
Triglycerides	$192.66 \pm 23.77$	$187.66 \pm 11.89$	$185.66 \pm 33.34$	$227.66 \pm 70.34$		
$(mg dL^{-1})$						
<b>Total Proteins</b>	$3.23 \pm 0.18$	$3.00\pm0.21$	$2.76 \pm 0.26$	$4.33 \pm 0.64$		
$(g.dL^{-1})$						
Total Cholesterol	$193.66 \pm 9.49^{b}$	$187.66 \pm 5.36^{b}$	$183.00 \pm 5.68^{b}$	$226.66 \pm 5.48^{a}$		
$(mg.dL^{-1})$						
HDL	$148.00 \pm 6.24^{a}$	$119.66 \pm 19.17^{a,b}$	$87.66 \pm 4.81^{b}$	$100.00 \pm 10.11^{a,b}$		
$(mg.dL^{-1})$						
LDL	$98.00 \pm 17.43^{b}$	$85.66 \pm 10.98^{b}$	$244.66 \pm 52.71^{a,b}$	$463.00 \pm 97.82^{a}$		
$(mg.dL^{-1})$						
Creatinine	$0.11 \pm 0.006$	$0.09 \pm 0.003$	$0.09\pm0.04$	$0.09 \pm 0.009$		
$(mg.dL^{-1})$						
Urea	$3.76 \pm 0.93$	$2.96\pm0.58$	$4.46 \pm 0.48$	$4.76\pm0.58$		
$(mg.dL^{-1})$						

Table 2

Parameters	Treatment			
	Control	Ethanol	Globulol 2.5 mg.L <sup>-1</sup>	OE 20 mg.L <sup>-1</sup>
Hematocrit (%)	$22.40 \pm 0.73^{b}$	$26.12 \pm 1.65^{a,b}$	$26.78 \pm 0.99^{a,b}$	$28.42 \pm 0.95^{a}$
Hemoglobin (g%)	$7.52 \pm 0.48$	$7.07 \pm 0.47$	$6.95 \pm 0.29$	$7.34 \pm 0.30$
Erythrocytes (106 μL-1)	$1.64 \pm 0.07^{b}$	$2.01 \pm 0.12^{a}$	$2.06\pm0.08^a$	$2.11 \pm 0.08^{a}$
Leucocytes $(10^3  \mu \text{L}^{-1})$	$12.01 \pm 5.00^{\circ}$	$57.61 \pm 10.00^{a,b}$	$70.87 \pm 15.00^{a,b}$	$48.82 \pm 7.00^{a,b,c}$
Thrombocytes $(10^3  \mu L^{-1})$	$103.00 \pm 12.00^{a}$	$95.80 \pm 11.20^{a}$	$108.90 \pm 12.30^{a}$	$20.50 \pm 3.40^{b}$
MCV (fL)	$137.50 \pm 4.45^{a}$	$129.90 \pm 3.09^{a,b}$	$129.87 \pm 1.55^{b}$	$134.56 \pm 1.29^{b}$
MCHC (g .L <sup>-1</sup> )	$27.00\pm0.11$	$33.95 \pm 3.54$	$25.75 \pm 0.33$	$25.95 \pm 0.58$
Monocytes (%)	$10.66 \pm 4.48$	$8.33 \pm 3.52$	$9.33 \pm 2.33$	$12.00 \pm 0.57$
Lymphocytes (%)	$85.33 \pm 4.91$	$86.33 \pm 4.97$	$87.00 \pm 2.51$	$85.33 \pm 1.20$
Heterophils (%)	$3.33 \pm 0.33$	$4.66 \pm 1.45$	$3.33 \pm 0.88$	$2.33 \pm 0.88$
Eosinophils (%)	$0.66 \pm 0.33$	$0.66 \pm 0.33$	$0.33 \pm 0.33$	$0.33 \pm 0.33$
Hemolysis of fish (%)	$91.66 \pm 3.08$	88.69± 1.97	$78.38 \pm 3.88$	83.44± 5.25

Figure 1

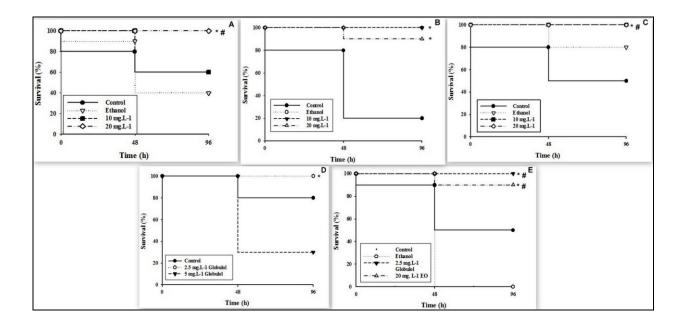
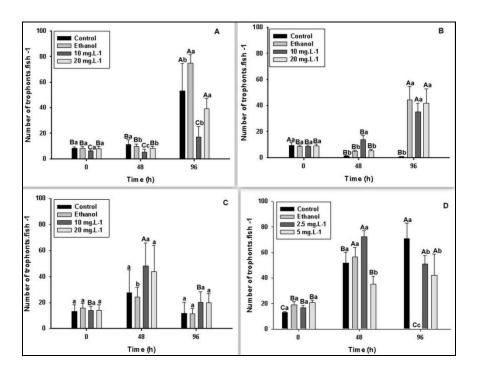


Figure 2



## 4 CONCLUSÃO

- O óleo essencial (OE) de folhas de *Hyptis mutabilis* e constituinte (-)-globulol não foram eficazes na diminuição do número de trofontes decorrentes da parasitose por *Ichthyophthirius multifiliis* em jundiás;
- Dentre as três metodologias avaliadas, a metodologia que apresentou melhor desempenho considerando-se a sobrevivência dos animais parasitados foi banhos diários de 1 hora por 4 dias consecutivos;
- A sobrevivência dos animais parasitados com *Ichthyophthirius multifiliis* foi aumentada quando os mesmos foram expostos ao OE (20 mg.L<sup>-1</sup>) e ao seu constituinte majoritário, (-)-globulol (2,5 mg.L<sup>-1</sup>) através de banhos diários de 1h por 4 dias consecutivos;
- O OE de *Hyptis mutabilis* e o (-)-globulol não inibiram a atividade hemolítica de *A. hydrophila* em eritrócitos de peixes sadios;
- A adição do OE de *Hyptis mutabilis* e do (-)-globulol à água não apresentou correlação clara entre a alteração dos parâmetros hematológicos e bioquímicos em jundiás sadios e o aumento da sobrevivência de parasitados.

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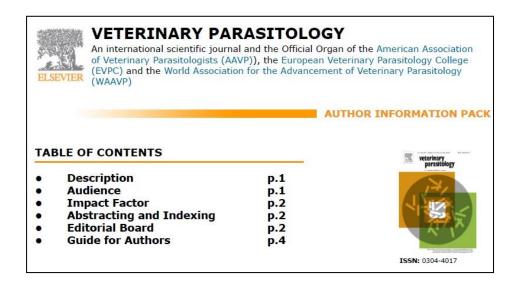
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# **ANEXO**

Anexo A - Normas para publicação no periódico Veterinary Parasitology

## **VETERINARY PARASITOLOGY**



#### **DESCRIPTION**

This journal is concerned with those aspects of helminthology, protozoology and entomology which are of interest to animal health investigators, veterinary practitioners and others with a special interest in parasitology. Papers of the highest quality dealing with all aspects of disease prevention, pathology, treatment, epidemiology, and control of parasites in all domesticated animals, fall within the scope of the journal. Papers of geographically limited (local) interest which are not of interest to an international audience will not be accepted. Authors who submit papers based on local data will need to indicate why their paper is relevant to a broader readership.

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Studies dealing with parasite control by means of natural products, both in vivo and in vitro, fall within the scope of the journal, but only if well documented and with therapeutically relevant minimum inhibitory concentrations of the active compound(s) being clearly demonstrated.

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- 3. Rapid Communications
- 4. Short Communications
- 5. Letters to the Editor

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